



UNIVERSIDADE
ESTADUAL DE LONDRINA

VICTOR FATTORI

**MECANISMOS DE INIBIÇÃO DA INFLAMAÇÃO E DOR
PELA CURCUMINA NOS MODELOS DE ARTRITE INDUZIDA
POR DIÓXIDO DE TITÂNIO E INFLAMAÇÃO INDUZIDA
POR ÂNION SUPERÓXIDO EM CAMUNDONGOS**

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Dissertação de mestrado apresentada ao programa de pós-graduação em Patologia Experimental da Universidade Estadual de Londrina, como requisito parcial à obtenção do título de Mestre em Patologia Experimental

Orientador: Prof. Dr. Waldiceu Aparecido Verri Junior.

Londrina
2016

Ficha de identificação da obra elaborada pelo autor, através do Programa de Geração Automática do Sistema de Bibliotecas da UEL

Fattori, Victor.

Mecanismos de inibição da inflamação e dor pela curcumina nos modelos de artrite induzida por dióxido de titânio e inflamação induzida por ânion superóxido em camundongos / Victor Fattori. - Londrina, 2016.
120 f. : il.

Orientador: Waldiceu Aparecido Verri Junior.

Dissertação (Mestrado em Patologia Experimental) - Universidade Estadual de Londrina, Centro de Ciências Biológicas, Programa de Pós-Graduação em Patologia Experimental, 2016.

Inclui bibliografia.

1. artrite - Teses. 2. curcumina - Teses. 3. dor - Teses. 4. inflamação - Teses. I. Verri Junior, Waldiceu Aparecido. II. Universidade Estadual de Londrina. Centro de Ciências Biológicas. Programa de Pós-Graduação em Patologia Experimental. III. Título.

VICTOR FATTORI

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CURCUMINA NOS MODELOS DE ARTRITE INDUZIDA POR DIÓXIDO
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EM CAMUNDONGOS**

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Londrina, 14 de março de 2016.

Dedico a minha esposa Natasha, meu irmão
Vinicius e meus pais.

AGRADECIMENTOS

Agradeço, primeiramente, aos meus pais. Meus objetivos foram alcançados graças aos seus incomensuráveis esforços.

Ao meu irmão Vinicius, por fazer parte da minha vida. Agradeço pelo suporte contínuo, por compartilhar sua visão de mundo e expandir a minha. Sua presença e auxílio léxico são indispensáveis. É, de longe, a pessoa mais inteligente que conheço.

À minha esposa Natasha, por me fazer uma pessoa melhor e nunca desistir de mim. É gratificante saber que não estou e nunca estive sozinho nessa jornada. Atribuo grande parte do que consegui à sua ajuda perene.

Ao meu orientador, Prof^o Dr^o Waldiceu Aparecido Verri Junior, pela confiança, paciência e oportunidade de trabalhar em seu laboratório. É um privilégio ser orientado por alguém que tanto admiro.

À minha tia Maria. Gostaria que estivesse presente para testemunhar minha conquista.

Aos meus familiares, por se fazerem presentes.

A Felipe Almeida de Pinho Ribeiro e Sérgio Marques Borghi, pelos incontáveis finais de semana no laboratório, amizade e ajuda. Agradeço por contribuírem para meu crescimento científico.

Aos colegas de laboratório, pelo auxílio nas demais atividades diárias, independente da participação no projeto.

Aos técnicos Zui e Pedro, pela conversa, pelo café, auxílio na pesagem de reagentes e com cortes histológicos.

Aos membros da Banca Examinadora, pela paciência, disposição e contribuição fundamental para construção dessa dissertação.

Ao café Pilão e Red Bull, por afastarem o sono e permitir a conclusão do texto que ora pode ser lido

“Demore o tempo que for para você decidir o que quer da vida, e depois que decidir, não recue ante nenhum pretexto, pois o mundo tentará te dissuadir”

Profeta Zaratustra, de Friedrich Nietzsche.

FATTORI, Victor. **Mecanismos de inibição da inflamação e dor pela curcumina nos modelos de artrite induzida por dióxido de titânio e inflamação induzida por ânion superóxido em camundongos**. 2016. 120 f. Dissertação (Mestrado em Patologia Experimental) – Universidade Estadual de Londrina, Londrina, 2016.

RESUMO

Os turmericos (*Curcuma longa*) têm sido utilizados pela Medicina Ayevedica e chinesa por milhares de anos para o tratamento de diversas doenças, como asma, úlcera e artrite. Curcumina, o principal constituinte derivado do rizoma da planta, é uma molécula pleiotrópica e possui atividade anti-inflamatória, em parte, através da inibição do NF- κ B. Tendo em vista essas características, curcumina é uma alternativa promissora para o tratamento da dor. Desse modo, nosso objetivo foi avaliar a atividade da curcumina em modelo agudo de dor induzido pelo superóxido de potássio (KO₂), um doador de ânion superóxido (O₂^{•-}) e modelo de artrite induzido por dióxido de titânio. Para avaliação da dor manifesta, foi contabilizado o número total de contorções abdominais (durante 20 minutos) ou número total de sacudidas de patas e o tempo lambida (durante 30 minutos). A hiperalgesia mecânica e térmica foi avaliada 0,5; 1; 3; 5 e 7 h após o estímulo utilizando um analgesímetro digital e placa quente, respectivamente. A produção de citocinas (IL-1 β , TNF- α e IL-10) e atividade do NF- κ B foram avaliados por ELISA, 3 h após o estímulo. A produção de O₂^{•-} foi avaliada através do ensaio nitroblue tetrazolium, e a atividade antioxidante total pelo ensaio ABTS, ambos 3 h após o estímulo. A atividade da mieloperoxidase e recrutamento de leucócitos foram avaliados 7 h após o estímulo. A expressão do mRNA de Nrf2, HO-1 e gp91^{phox} foram avaliados por RT-qPCR. O tratamento com curcumina 10 mg/kg reduziu dor manifesta induzida por O₂^{•-} de maneira dose dependente, além de inibir a hiperalgesia mecânica e térmica em todos os intervalos avaliados. A produção de citocinas induzidas pelo O₂^{•-} (IL-1 β e TNF- α) bem como a atividade do NF- κ B foram reduzidas pela curcumina. Esses efeitos foram acompanhados pelo aumento da produção da citocina anti-inflamatória IL-10. Nós demonstramos, que o tratamento com curcumina 10 mg/kg foi capaz de aumentar a expressão do mRNA para o fator de transcrição Nrf2 e seu alvo HO-1. Além disso, o tratamento com curcumina aboliu a produção do O₂^{•-}, restaurou a atividade antioxidante total e inibiu o recrutamento de leucócitos. Em um segundo momento, avaliamos o efeito da curcumina em modelo de artrite induzido dióxido de titânio. A hiperalgesia mecânica e edema foram avaliados 1, 3, 5, 7 e 24h após o estímulo e de dois em dois dias até o 30º dia após o estímulo. Nós observamos que o tratamento com curcumina na dose de 100 mg/kg foi efetiva na redução da dor, edema, recrutamento de neutrófilos. Como resultado disso houve menor degradação de proteoglicanos e maior preservação da articulação. Tendo isso como esteio, nós demonstramos que a curcumina apresenta atividades anti-inflamatórias e analgésica em modelos de dor crônica e dessa forma contribuímos para o entendimento do mecanismo de ação dessa molécula reforçando sua atividade pleiotrópica.

Palavras chaves: Ânion superóxido. Artrite. Curcumina. Dor. Dióxido de Titânio. IL-33. NF- κ B. Nrf2. Proteoglicanos

FATTORI, Victor. **Curcumin inhibits pain and inflammation in superoxide anion-induced acute pain and titanium dioxide-induced arthritis**. 2015. 120 p. Dissertation (Master's degree in Experimental Pathology) – Universidade Estadual de Londrina, Londrina, 2016.

ABSTRACT

Ayurvedic and Chinese medicine have used turmeric (*Curcuma longa*) as a medicinal herb for thousands of years to treat different diseases, such as asthma, ulcer, and arthritis. Curcumin, the main bioactive constituent derived from the rhizome of turmeric, is pleiotropic molecule and has its anti-inflammatory properties, at least in part, through inhibition of NF- κ B. Due these features, curcumin is a promising alternative to treat inflammatory pain. Therefore, we aim of to evaluate the activity of curcumin in two models: the first model was acute pain – potassium superoxide (KO₂)–induced – and the second model was arthritis – titanium dioxide-induced – in mice. Experimental pain was induced by administration of 30 μ g (i.pl.) or 1 mg (i.p.) of radical superoxide anion (O₂^{•-}) donor KO₂. Overt pain-like behaviours were quantified by counting the number of abdominal writhings over 20 minutes, or the number of paw flinches and the time spent licking the paw over 30 minutes. Mechanical and thermal hyperalgesia was evaluated in the paw 0.5, 1, 3, 5 and 7 h after stimulus. Cytokine levels (IL-1 β , TNF- α , and IL-10) and NF- κ B activity in the paw tissue were measured by ELISA 3 h after stimulus. O₂^{•-} production was evaluated by nitroblue tetrazolium (NBT) assay, and the antioxidant capacity was evaluated by measuring the ABTS radical scavenging ability, both in samples collected 3 h after stimulus. Myeloperoxidase activity and total leukocyte recruitment was measured collected 7 h after stimulus in paw skin samples and peritoneal wash, respectively. Nrf2, HO-1 and gp91^{phox} mRNA expression in paw skin was determined 3 h after stimulus by real time PCR. We observed that curcumin inhibited overt pain-like behaviours induced by O₂^{•-} in a dose-dependent manner, and inhibited mechanical and thermal hyperalgesia in all mesuared intervals as well. O₂^{•-}-induced production of pro-inflammatory cytokines (IL-1 β and TNF- α) and activity of NF- κ B were suppressed by curcumin, and these effects were accompanied by an enhanced production of the anti-inflammatory cytokine IL-10. We found increased expression of mRNA for transcription factor Nrf2 and HO-1 in curcumin-treated animals. Curcumin restored antioxidant capacity, abolished O₂^{•-} production and myeloperoxidase activity, and reduced gp91^{phox} mRNA expression. In arthritis model, mechanical threshold and edema were evaluated after 1, 3, 5, 7, 24h and then each two days until 30th day after titanium dioxide. We observed that curcumin at 100 mg/kg reduced titanium dioxide-induced pain, edema and neutrophils recruitment. As consequence of that, curcumin reduced proteoglycan degradation and avoid joint lesion. Taking this into account, we demonstrate that curcumin provided anti-inflammatory and analgesic properties in models of acute and chronic pain, and this may contribute to understanding the mechanisms of action of this molecule reinforcing, its pleiotropic activity and supporting its use in clinical trials.

Keywords: Arthritis. Curcumin. IL-33. NF- κ B. Nrf2 – Pain. Proteoglycan. Superoxide anion. Titanium Dioxide

LISTA DE ILUSTRAÇÕES

Figura 1 – Vias de sinalização do NF- κ B e Nrf2	19
Figura 2 – Descrição de René Descartes do processo de percepção da dor	24
Figura 3 – Alvos moleculares da curcumina	31

LISTA DE TABELA

Tabela 1 – Principais mediadores e estímulos pró-nociceptivos e seus receptores.....	24
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LISTA DE ABREVIATURAS E SIGLAS

ABTS	Ácido 2,2'-azino-bis-3-ethylbenzthiazolino-6-sulphonico
AEC	Antes da Era Comum
ASIC	Canal Iônico Sensível a Ácidez
cGMP	Guanosina Monofosfato
COX	Ciclo-oxigenase
DAMP	Molécula Padrão Associada a Dano
EC	Era Comum
ELAM	Molécula de Adesão Leucocitária Endotelial
ERN	Espécies Reativas de Nitrogênio
ERO	Espécies Reativas de Oxigênio
FoxO3a	Fator de Transcrição Forkhead subfamília O
fMLP	N-Formilmetil-Leucyl-Fenilalanina
GABA	Ácido Gama-Aminobutírico
GSH	Glutathiona Reduzida
HO-1	Heme Oxigenase-1
ICAM-1	Molécula de Adesão Intracelular-1
IL	Interleucina
Keap-1	Proteína 1 associada a ECH similar a Kelch
KO ₂	Superóxido de Potássio
MPO	Mieloperoxidase
mRNA	RNA mensageiro
NADPH	Fosfato de Dinucleotídio de Nicotinamida e Adenina
Nav	Canal de Sódio Voltagem-Resistente
NBT	Nitrozul de Tetrazólio
NF-κB	Fator Nuclear de Cadeia Leve kappa de Células B Ativadas (Fator de Transcrição Nuclear kappa B)
Nrf2	Fator 2 Relacionado ao Fator de Transcrição Nuclear Eritróide 2
NLR	Receptor Semelhante a Nod
PAMP	Molécula Padrão Associada à Membrana de Patógenos
PGE ₂	Prostaglandina E ₂
PKA	Proteína Kinase A
PKC	Proteína Kinase C

PKG	Proteína Kinase G
PRR	Receptor de Reconhecimento de Padrão
RANK	Receptor Ativador do NF- κ B
RANKL	Ligante do Receptor Ativador do NF- κ B
SOD	Superóxido Dismutase
RLR	Receptor Semelhante a Rig
SPM	Mediadores Lipídicos Pró-Resolução
TLR	Receptor Semelhante a Toll
TNF- α	Fator de Necrose Tumoral alfa
TRPA1	Receptor de Potencial Transitório Subfamília A, membro 1
TRPV1	Receptor de Potencial Transitório Subfamília V, membro 1
VAS	Escala Analógica Visual
VCAM-1	Molécula de Adesão Vascular-1

SUMÁRIO

1	INTRODUÇÃO	15
1.1	INFLAMAÇÃO	15
1.1.1	NF-κB	16
1.1.2	Nrf2.....	18
1.2	DOR.....	19
1.2.1	Fibras Nociceptivas	20
1.2.2	Fisiologia Da Dor – Aspectos Gerais.....	21
1.2.2.1	Dor inflamatória	22
1.3	ESPÉCIES REATIVAS DE OXIGÊNIO E NADPH OXIDASE	25
1.3.1	Relação Entre Ânion Superóxido E Dor	26
1.4	ARTRITE	27
1.5	CURCUMINA.....	28
2	OBJETIVOS	32
2.1	OBJETIVO GERAL.....	32
2.1.1	Objetivos Específicos (Modelo Crônico).....	32
2.1.2	Objetivos Específicos (Modelo Agudo).....	32
	REFEÊNCIAS	33
3	ARTIGO 1 PUBLICADO (PHARMACOLOGICAL RESEARCH)	39
4	ARTIGO PARA PUBLICAÇÃO (JOURNAL OF NATURAL PRODUCTS)	78
5	ARTIGO 2 PUBLICADO (INFLAMMATION RESEARCH)	96
6	CONCLUSÃO	120

1 INTRODUÇÃO

1.1 INFLAMAÇÃO

Inflamação é uma das condições médicas com registros mais antigos. Isso é evidenciado com a descoberta de hierógrafos datados 2700 AEC desenvolvidos pela civilização sumeriana, e que representavam os sinais e sintomas da inflamação: vermelhidão, calor, edema e dor. Esses mesmos sinais e sintomas foram descritos e eternizados pelas letras do romano Aulus Cornelius Celsus, no primeiro século EC (ROCHA E SILVA, 1978). A inflamação aguda é uma resposta apropriada para uma defesa efetiva do hospedeiro e manutenção da homeostase, e é hodiernamente subdividida em duas fases: iniciação e resolução (MADERNA; GODSON, 2009). Entretanto, se ocorrer de forma excessiva pode levar a destruição do tecido, fibrose e com eventual perda de função do órgão afetado (MADERNA; GODSON, 2009). Em face de limitar tal resposta, mecanismos compensatórios foram desenvolvidos e mantidos ao longo do processo evolutivo, tais como: produção de citocinas anti-inflamatórias e agentes antioxidantes (LAWRENCE; WILLOUGHBY; GILROY, 2002; NITURE; KHATRI; JAISWAL, 2014). O NF- κ B e Nrf2 são fatores de transcrição que têm papel chave nessa regulação, e serão discutidos com mais detalhes posteriormente.

O processo inflamatório é iniciado, geralmente, após o reconhecimento de PAMPs ou DAMPs por PRRs (TLR, NLR e RLR, por exemplo) presentes em células residentes, especialmente macrófagos (IWASAKI; MEDZHITOV, 2010; TAKEUCHI; AKIRA, 2010). Esse reconhecimento tem como resultado final a ativação do NF- κ B e consequente produção de citocinas pró-inflamatórias, tais como IL-1 β , TNF- α , e IL-6 (IWASAKI; MEDZHITOV, 2010; TAKEUCHI; AKIRA, 2010). Outros mediadores como quimiocinas (CXCL1 [KC em murinos] e CXCL2), proteína do complemento C5a LTB₄, PGE₂ e endotelinas (CUNHA, F. Q.; CACINI; FERREIRA, 1986; FERREIRA; ROMITELLI; DE NUCCI, 1989; GUERRERO et al., 2008; RIBEIRO et al., 1997; TING et al., 2008; VERRI; CUNHA; PARADA; WEI; et al., 2006), colaboram para amplificação do processo. Em conjunto, esses mediadores são reponsáveis por ativar o endotélio e guiar precisamente, inicialmente, neutrófilos para o foco inflamatório. Na última década, o processo de recrutamento de neutrófilos tem sido estudado elegantemente (MCDONALD et al., 2010; SREERAMKUMAR et al., 2014). É um processo que depende da

35 característica inicial do estímulo nócico e segue uma cascata temporal, espacial e
36 hierárquica de mediadores (MCDONALD et al., 2010; SREERAMKUMAR et al.,
37 2014).O recrutamento de neutrófilos em sinusóides do fígado em modelo de
38 inflamação estéril são dependentes da integrina Mac-1 e ICAM-1 (MCDONALD et
39 al., 2010). Em contraste, após estímulo com *E. coli* o recrutamento de neutrófilos é
40 dependente de CD44 em detrimento de Mac-1, revelando diferentes mecanismos de
41 recrutamento de neutrófilos em modelos de inflamação estéril e não estéril
42 (MCDONALD et al., 2010). Neutrófilos tendem a responder melhor à moléculas
43 quimioatraentes finais como fMLP e C5a quando comparado à moléculas iniciais
44 como IL-8 e LTB₄ (FOXMAN; CAMPBELL; BUTCHER, 1997). Corroborando esse
45 fato, em modelo de inflamação estéril camundongos knockout para receptor de
46 fMLP, mas não CXCR2, apresentaram redução significativa de neutrófilos no foco
47 necrótico, o que demonstra que o receptor de fMLP guia localização precisa de
48 neutrófilos em áreas de necrose, fortificando a ideia de hierarquia de mediadores
49 para o recrutamento de neutrófilos (MCDONALD et al., 2010). Os neutrófilos na
50 circulação normalmente têm uma vida útil curta e rapidamente sofrem apoptose
51 constitutiva. No entanto, o microambiente inflamatório estimula a expressão de
52 genes relacionados com a longevidade, como FoxO3a (JONSSON; ALLEN; PENG,
53 2005), além de promover ativação do NF-κB e contribuir para produção de mais
54 citocinas pró-inflamatórias e EROs (WRIGHT et al., 2010).

55 A questão temporal também é relevante na resolução do processo
56 inflamatório, o qual é visto como ativo e altamente regulado (SERHAN et al., 2015).
57 A presença de neutrófilos apoptóticos associada a produção de PGE₂ inicia a troca
58 na classe de mediadores lipídicos de pró-inflamatórios para pró-resolução (LEVY et
59 al., 2001). Esses neutrófilos apoptóticos são fagocitados por macrófagos não
60 flogísticos, em um processo conhecido como eferocitose. Os principais mediadores
61 pró-resolução produzidos são as lipoxinas, maresinas e resolvinas (SERHAN et al.,
62 2015). Em conjunto, essas e outras moléculas atuam na redução e no reparo dos
63 danos provocados durante a inflamação aguda e promovem a resolução do
64 processo de forma ativa.

65

66 1.1.1 NF-κB

67 A cascata de sinalização do NF-κB consiste, basicamente, em uma série de
68 elementos regulatórios que culminam na ativação kinases responsáveis pela

69 fosforilação, ubiquitinação e degradação de proteínas inibidoras desse fator de
70 transcrição. O NF- κ B é composto pelo agrupamento de complexos homo e
71 heterodiméricos (HAYDEN; GHOSH, 2008). Nove dos 15 potenciais dímeros
72 apresentam atividade transcripcional, ou seja, são capazes de se ligar na região
73 promotora de genes transcritos pelo NF- κ B, região essa denominada locais κ B. O
74 dímero de maior importância é o formado pelas subunidades p50-p65 (SEN; SMALE,
75 2010). Assim como toda via de sinalização, sua modulação negativa é essencial
76 para manutenção da homeostase. O inibidor do NF- κ B é o I κ B, o qual é composto
77 das subunidades α , β e ϵ . Esse fator tem a função de inibir os sinais de localização
78 nuclear do heterodímero p50-p65 e impedir que ocorra a transcrição de genes
79 relacionados ao NF- κ B. A ativação desse fator de transcrição pode ocorrer por duas
80 vias: canônica e não-canônica, e para efeito de discussão, iremos focar na primeira
81 devido seu mecanismo *upstream* de sinalização ser dependente da transdução de
82 sinal via PRR, TNFR e/ou IL-1R (HAYDEN; GHOSH, 2008). Para que haja ativação
83 do NF- κ B é necessário que seu inibidor (I κ B) seja degradado. Para tanto, a presença
84 de kinases é indispensável. Como citado anteriormente, a estimulação dos PRR que
85 ocorre pelos sinais produzidos durante a lesão tecidual e/ou infecção leva à ativação
86 do NF- κ B através da degradação de I κ B. O complexo de kinases do I κ B (IKK),
87 formado pelas subunidades catalíticas IKK α , IKK β , e pela subunidade regulatória
88 IKK γ (NEMO), inicia o processo de fosforilação do I κ B, em sua subunidade α em
89 dois resíduos específicos de serina (KARIN; BEN-NERIAH, 2000), propiciando a
90 ubiquitinação desse inibidor e consequente sua degradação via proteossomo. Isso
91 permite que o heterodímero p50-p65 seja translocado até o núcleo da célula e
92 induza a transcrição de genes pró-inflamatórios (Figura 1 A) (HAYDEN; GHOSH,
93 2008; SEN; SMALE, 2010).

94 As EROs são capazes de oxidar diversas proteínas, especialmente com
95 resíduos de cisteína (tióis), essa característica permite a ativar diversas vias de
96 sinalização, entre elas a do NF- κ B (GLOIRE; PIETTE, 2009). De fato, a via do NF- κ B
97 é conhecida como sensível ao estado redox da célula (TOLEDANO; LEONARD,
98 1991). Após ubiquitinação e degradação do I κ B α , resíduos específicos de serina da
99 subunidade p65 são fosforilados por PKA. Isso permite a interação de p65 com a
100 molécula coativadora do NF- κ B (mas não apenas) CBP (proteína ligadora de CREB).
101 Essa fosforilação por PKA é elevada na presença de EROs e isso possibilita o
102 aumento da atividade do NF- κ B. De fato, tratamento com antioxidantes diminui a

103 associação de p65 com CBP na região promotora do gene da IL-8, por exemplo
104 (JAMALUDDIN et al., 2007). Focando no ânion superóxido, tratamento com M40403
105 (molécula mimética da enzima SOD) inibiu a ativação do NF- κ B, relacionada com
106 diminuição da degradação do I κ B, sugerindo a participação do ânion superóxido da
107 regulação redox desse fator de transcrição (NDENGELE et al., 2005)

108 De modo a controlar a resposta inflamatória, o NF- κ B estimula a transcrição
109 de genes anti-inflamatórios, como por exemplo a citocina IL-10 (HAYDEN; GHOSH,
110 2008; SEN; SMALE, 2010), proteínas com função ubiquitina-ligase (A20) (CHEN,
111 2005), seu inibidor I κ B (LAWRENCE, 2009), moléculas antioxidantes como
112 catalase, GSH e SOD, entre outros (HAYDEN; GHOSH, 2008; SEN; SMALE, 2010).

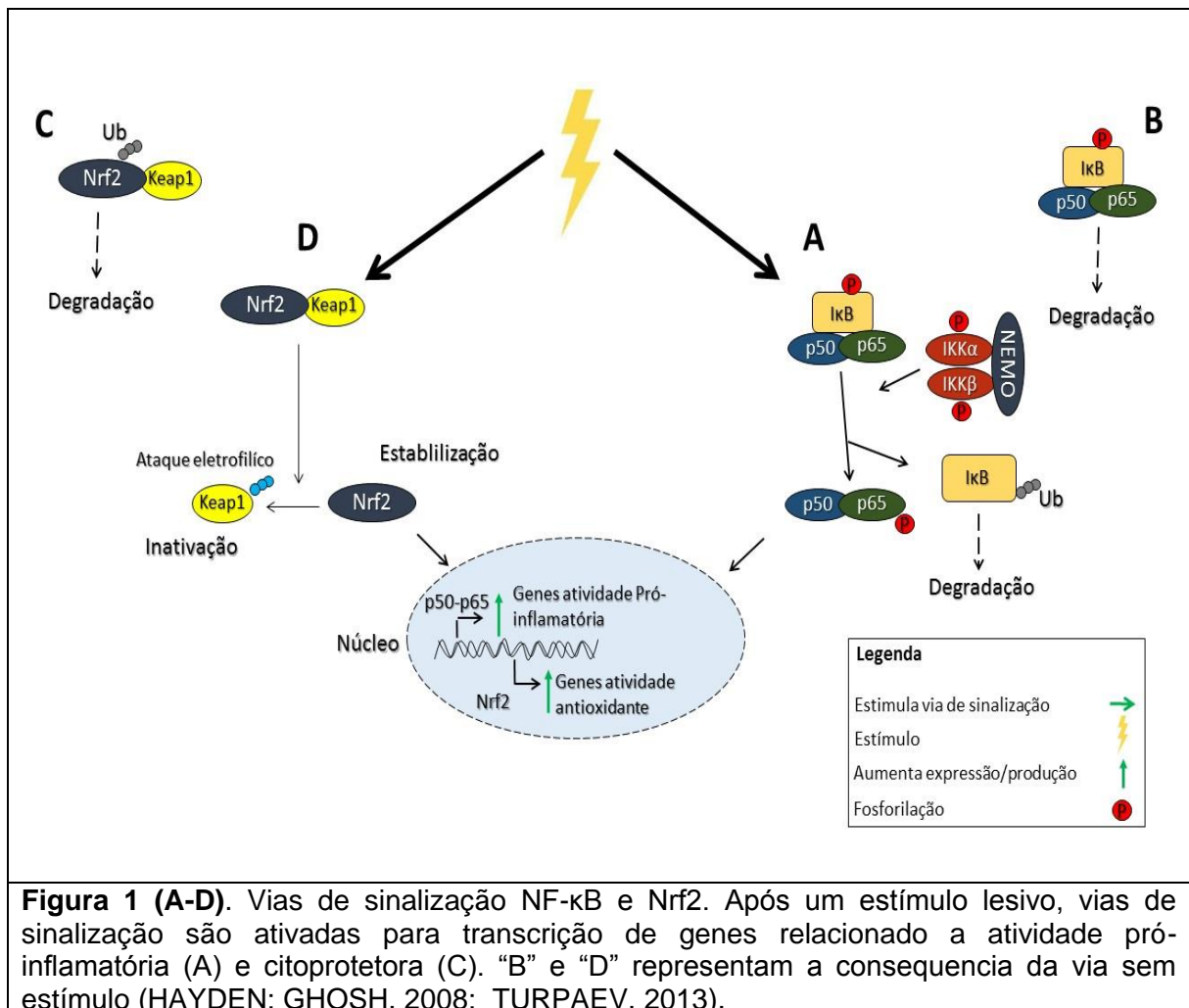
113

114 1.1.2 Nrf2

115 O fator de transcrição Nrf2 é um dos sistemas citoprotetores mais importantes
116 adquirido pelos vertebrados ao longo da evolução (GAO, B.; DOAN; HYBERTSON,
117 2014). De fato, a substituição de um único nucleotídeo (SNP) na região promotora de
118 genes transcritos pelo Nrf2 é capaz de conferir ao indivíduo maior susceptibilidade a
119 lesão pulmonar (MARZEC et al., 2007). O Keap-1 é uma molécula rica em
120 resíduos de cisteína e com atividade ubiquitina-ligase e conseqüentemente é o
121 inibidor dessa via de sinalização (ITOH et al., 1997). Portanto, sob condições
122 fisiológicas o Nrf2 está associado ao Keap-1, constantemente ubiquitinado e degradado
123 via proteassomo (Figura 1C). Entretanto, após modificações em Keap-1 decorrentes
124 de ataque eletrofílico em resíduos tióis específicos de cisteína, associado a
125 fosforilação em resíduos específicos de serina do Nrf2, ocorre a dissociação do
126 Keap-1 e estabilização do fator de transcrição. O Nrf2 estável é capaz de se
127 dimerizar com proteína sMaf (*small Maf*) e com conseqüente translocação para
128 núcleo (HUANG; NGUYEN; PICKETT, 2002; LI et al., 2008; NITURE; JAIN;
129 JAISWAL, 2009) em genes com regiões promotoras denominadas ARE (Elementos
130 de Resposta Antioxidante) para que haja aumento da expressão de genes
131 relacionados a resposta citoprotetora (Figura 1D) (TURPAEV, 2013).

132 Um dos alvos mais interessantes do Nrf2 é a enzima HO-1 (TURPAEV, 2013).
133 HO-1 apresenta atividade anti-inflamatória, antioxidante e antinociceptiva (STEINER
134 et al., 2001; TURPAEV, 2013). Para exercer sua atividade antioxidante, HO-1
135 cataliza a reação do heme que leva a formação de monóxido de carbono, ferro e
136 biliverdina. A enzima biliverdina redutase é responsável pela conversão da

137 biliverdina em bilirrubina, a qual tem potente atividade antioxidante, e com
 138 participação das EROs há a conversão de bilirrubina em biliverdina novamente.
 139 Desse modo esse ciclo garante a neutralização das EROs e provê atividade
 140 antioxidante para HO-1 (PAE et al., 2010). Essa via ainda é capaz de inibir a
 141 migração e adesão de neutrófilos para o foco inflamatório (FREITAS et al., 2006),
 142 que são fundamentais para o desenvolvimento e manutenção de estados
 143 hiperalgésicos (CUNHA, T. M. et al., 2008). Além disso, HO-1 é capaz de ativar a via
 144 de canais de potássio sensíveis a cGMP/PKG/ATP (KAPITULNIK, 2004). Essa via é
 145 responsável por aumentar o limiar de ativação dos nociceptores e consequentemente
 146 diminuir a despolarização provendo atividade antinociceptiva (STAURENGO-
 147 FERRARI et al., 2014; ZARPELON; SOUZA; et al., 2013).



148

149 1.2 DOR

150 O sistema nervoso detecta e interpreta uma vasta gama de estímulos
 151 térmicos e mecânicos, bem como irritantes químicos ambientais ou endógenos.

152 Quando intenso, esses estímulos são capazes de gerar dor aguda. A dor é um
153 sintoma presente em muitas doenças e é uma das maiores causas de procura ao
154 atendimento médico. Durante o processo evolutivo, a percepção da dor, ou
155 nocicepção, foi um mecanismo que se manteve por conferir ao indivíduo a
156 capacidade de autopreservação, identificando situações que possam causar danos
157 ou mesmo controlar as consequências de uma lesão já ocorrida. Esse caráter
158 evolutivo é evidenciado em indivíduos portadores da doença conhecida como
159 “insensibilidade congênita à dor”, ou “analgesia congênita”, já que raramente
160 atingem a idade adulta, por não disporem desses mecanismos inatos protetores. A
161 compreensão dos mecanismos envolvendo a dor, especialmente as crônicas, é
162 essencial, tendo em vista seu impacto na qualidade de vida das pessoas afetadas.
163 Tal a influência da dor na qualidade de vida, sua presença constante é um fator de
164 risco consistente para pensamentos e comportamentos suicidas (CALATI et al.,
165 2015).

166

167 1.2.1 Fibras Nociceptivas

168 A percepção de um estímulo como doloroso só é possível graças a presença
169 de neurônios sensoriais, denominados de nociceptores. Nesse sentido, a
170 nocicepção pode ser considerada uma modalidade sensorial tanto quanto visão ou
171 olfato, já que um determinado estímulo de determinada intensidade e proporção
172 pode ser detectado por células especializadas para tanto. Existem dois tipos de
173 fibras nociceptivas: A e C, sendo que a primeira se subdivide em $A\alpha$, $A\beta$ e $A\delta$ (BRAZ
174 et al., 2014). As fibras do tipo $A\alpha$ e $A\beta$ apresentam axônios com maior quantidade de
175 mielina e respondem a estímulos inócuos na pele, músculos e articulações. As fibras
176 do tipo $A\delta$ são finamente mielinizadas e respondem a estímulos mecânicos, térmicos
177 e químicos. As fibras do tipo C não apresentam mielina e respondem também a
178 estímulos térmicos, químicos e mecânicos, são denominadas de fibras “silenciosas” e
179 são as responsáveis pela resposta a estímulos como capsaicina (presente na
180 pimenta) (TODD, 2010). O corpo celular desses neurônios estão no gânglio da raiz
181 dorsal (DRG) e suas projeções vão para lâminas do corno dorsal da medula espinal,
182 variando de acordo com o tipo de fibra (BRAZ et al., 2014; TODD, 2010). As fibras C
183 e $A\delta$ conduzem informações para áreas nociceptivas específicas nas lâminas I e II
184 da superfície do corno dorsal da medula espinal e também para neurônios de
185 espectro dinâmico amplo na lâmina V, os quais codificam tanto informações de

186 estímulos inócuos quanto nocivos. Por outro lado, fibras amplamente mielinizadas
187 A β transmitem informações como toque leve ou estímulos mecânicos inócuos para
188 estruturas no corno dorsal da medula espinal nas lâminas III e IV (TODD, 2010).

189

190 1.2.2 Fisiologia Da Dor – Aspectos Gerais

191 O entendimento da fisiologia da dor no mundo moderno se inicia com René
192 Descartes em seu livro “Treatise of Man”, 1664. Para isso, Descartes utilizou seu
193 icônico diagrama de um garoto próximo a uma fogueira (Figura 2). Ele propôs que o
194 fogo ativa fibras em locais periféricos e tal informação seria transmitida para o
195 cérebro, onde há o reconhecimento de tal estímulo como doloroso. Essa informação
196 desencadeia uma resposta de retirada do pé de próximo do estímulo nócico (Figura
197 2). Para Descartes, a dor era apenas uma consequência de uma ativação linear
198 periférica para o cérebro (MOAYEDI; DAVIS, 2013), definição essa que não é
199 errônea, apenas incompleta. Muito se evoluiu desde então. Isso é exemplificado com
200 a definição de dor pelo comitê de taxonomia da Associação Internacional para o
201 Estudo da Dor (IASP): “uma experiência sensorial e emocional desagradável que é
202 associada a lesões reais ou potenciais ou descrita em termos de tais lesões”. Desse
203 modo, cada indivíduo interpreta a dor de acordo com suas experiências, o que torna
204 inconcebível a dissociação do caráter emocional para seu entendimento. Outros
205 fatores interferem no limiar nociceptivo e sua magnitude, e que não podem ser
206 ignorados ou subjulgados, dentre eles estão estado de humor, atenção
207 (BUSHNELL; CEKO; LOW, 2013) e expectativas em relação à intensidade do
208 estímulo (WIECH et al., 2014).

209 O entendimento contemporâneo da fisiologia da dor inclui sensibilização
210 periférica de nociceptores, sensibilização espinal, plasticidade neuronal e mudança
211 de fenótipo de células do sistema imune (MOGIL; YU; BASBAUM, 2000;
212 REICHLING; LEVINE, 2009; SCHOLZ; WOOLF, 2002; WOOLF; SALTER, 2000).
213 Após ser gerado periféricamente por um estímulo nociceptivo, o mecanismo
214 ascendente envolve o impulso nervoso percorrer o neurônio nociceptivo de primeira
215 ordem até o corno dorsal da medula espinal, ou até o núcleo trigeminal, onde estes
216 neurônios fazem sinapses com as terminações dos neurônios nociceptivos de
217 segunda ordem. A informação é transmitida ao neurônio nociceptivo de segunda
218 ordem por neurotransmissores excitatórios (peptídeo relacionado ao gene da
219 calcitonina [CGRP], glutamato e a substância P, por exemplo) que são liberados pelo

220 neurônio nociceptivo primário na fenda sináptica (BRAZ et al., 2014; SCHOLZ;
221 WOOLF, 2002). O mecanismo inibitório da dor envolve a liberação de mediadores
222 inibitórios (GABA, canabinóides e β -endorfina, por exemplo) de modo a limitar a
223 despolarização dos nociceptores ativados (MILLAN, 2002). Basicamente o processo
224 de nocicepção envolve transdução, condução, transmissão e percepção. A
225 transdução é a conversão de um estímulo nódico em atividade elétrica decorrente da
226 despolarização periférica dos nociceptores. Esse processo é estimulado por canais
227 iônicos expressos na membrana dos nociceptores (Tabela 1). A condução é a
228 passagem do potencial de ação pelos axônios para terminações centrais, e a
229 transmissão é a transferência sináptica de neurotransmissores na fenda sináptica
230 de um neurônio nociceptivo para o outro. Por fim, a percepção é o fornecimento da
231 informação referente ao estímulo inicial (através de sinapses no córtex
232 somatossensorial) sobre a localização e a intensidade do estímulo nódico. Outras
233 projeções envolvem sinapses na região da amígdala, contribuindo para o componente
234 emocional do processo doloroso (MILLAN, 1999; MOGIL et al., 2000; REICHLING;
235 LEVINE, 2009; SCHOLZ; WOOLF, 2002; TODD, 2010; WOOLF; SALTER, 2000).

236

237 1.2.2.1 Dor inflamatória

238 A dor de origem inflamatória resulta do aumento da excitabilidade
239 (sensibilização) dos nociceptores periféricos decorrente da estimulação de
240 mediadores pró-inflamatórios. Diversos mediadores podem contribuir para a
241 sensibilização dos nociceptores (Tabela 1). Nesse sentido, alguns termos utilizados
242 na prática clínica relacionada à dor inflamatória como alodinia (dor decorrente de um
243 estímulo inócua) e hiperalgesia (resposta exacerbada a um estímulo doloroso) que
244 também são utilizados na prática experimental em animais. A exacerbação ou
245 persistência da experiência da dor está presente em diversas doenças e representa
246 a principal causa da procura por atendimento médico.

247 A presença de neutrófilos é um denominador comum no que se refere a dor
248 inflamatória. Eles tem papel fundamental na manutenção da dor inflamatória
249 (CUNHA, T. M. et al., 2008). Neutrófilos recrutados produzem citocinas pró-
250 inflamatórias como IL-1 β , TNF- α e IL-33 (VERRI; CUNHA; PARADA; POOLE; et al.,
251 2006). Após um estímulo lesivo uma cascata de citocinas pró-inflamatórias precede
252 a liberação de aminas simpáticas e PGE₂ que levam a sensibilização dos
253 nociceptores. Essa cascata inicia com a liberação da alarmina IL-33 (ZARPELON;

254 CUNHA; et al., 2013) a qual estimula a produção sequencial de $\text{TNF-}\alpha \rightarrow \text{IL6} \rightarrow \text{IL-}$
255 $1\beta \rightarrow \text{PGE}_2$; e $\text{TNF-}\alpha \rightarrow \text{CXCL1} \rightarrow \text{IL-1}\beta \rightarrow$ aminas simpáticas (CUNHA, T. M. et al.,
256 2005; VERRI; CUNHA; PARADA; POOLE; et al., 2006). Essas moléculas, ao se
257 ligarem em seus receptores ativam kinases (PKA e PKC, por exemplo), que por sua
258 vez fosforilam canais iônicos (TRPV1, TRPA1, Nav1.8; por exemplo) e ativam os
259 nociceptores (BRAZ et al., 2014; SCHOLZ; WOOLF, 2002). Outros mediadores
260 como endotelina (VERRI et al., 2009; ZARPELON; CUNHA; et al., 2013;
261 ZARPELON et al., 2012), LTB_4 (GUERRERO et al., 2008), proteína do complemento
262 C5a (TING et al., 2008) e EROs, em especial ânion superóxido (JANES; NEUMANN;
263 SALVEMINI, 2012; MAIOLI et al., 2015; WANG et al., 2004) liberados por
264 neutrófilos contribuem de forma significativa para a hiperalgesia. Desse modo, a
265 depleção de neutrófilos (anticorpo anti-neutrófilo), inibição da migração dessas
266 células (fucoidan), ou utilização de antagonistas de endotelinas reduzem a migração
267 de neutrófilos e a hiperalgesia (CUNHA, T. M. et al., 2008; GUERRERO et al., 2008;
268 TING et al., 2008; VERRI et al., 2009).

269 A compreensão atual ainda envolve os nociceptores como moléculas ativas
270 na defesa do hospedeiro. Um fato que possibilitou um grande avanço nesse sentido
271 foi a descoberta da expressão de PRR (TLR 2, 3, 4 e 7; e NLR por exemplo)
272 associado à capacidade de liberar DAMP (CHIU et al., 2013; LIU et al., 2010; QI et
273 al., 2011). Isso sugere que os nociceptores são capazes de reconhecer estímulos
274 lesivos como bactérias, responder a esses estímulos e auxiliar na ativação de
275 células do sistema imune. Um dos principais DAMPs liberados e reconhecidos por
276 nociceptores é o ATP (Tabela 1). Seu mecanismo *downstream* envolve a ativação de
277 componentes da plataforma inflamassoma neuronal e de células do sistema imune,
278 que culmina na liberação da forma ativa da $\text{IL-1}\beta$ e IL-18 (MARIATHASAN et al.,
279 2006). Interessantemente, neurônios expressam os componentes dessa plataforma,
280 sendo capaz, não só de reconhecer, mas também de liberar essas citocinas em sua
281 forma ativa (DE RIVERO VACCARI et al., 2008). Além disso, o fato de *S. aureus* ser
282 capaz de modular a resposta de nociceptores e causar dor (CHIU et al., 2013),
283 reforça a ideia de os nociceptores participarem ativamente da montagem da
284 resposta imune.



Figura 2. Descrição de René Descartes do processo de percepção da dor. "Partículas de calor" (letra "A") ativam um local da pele (letra "B") conectado a uma válvula no cérebro (letras "d" e "e") por uma fibra fina (letra "C"). Tal atividade abre a válvula, permitindo que o espírito do animal possa fluir a partir de uma cavidade (letra "F") para os músculos, levando-os a recuar do estímulo, virar a cabeça e os olhos para a parte do corpo afetada, e mover a mão e girar o corpo de forma protetora afastando-se do fogo (Figura adaptada de (MOAYEDI; DAVIS, 2013).

285

Tabela 1. Principais mediadores e estímulos pró-nociceptivos e seus receptores

Mediador/Estímulo	Receptor
IL-1 β	IL-1R
TNF- α	TNFR
Bradicinina	B1 e B2
LTB ₄	BLT1 e BLT2
PGE ₂	EP ₃
ATP	P2X ₂ , P2X ₃ e P2X ₇
C5a	C5aR
Endotelina-1	ET _A e ET _B
Calor/ pH baixo	TRPV1
Mecânico/Irritantes	TRPA1
Frio	TRPM8
Frio	Nav1.8
pH baixo	ASIC

(BRAZ et al., 2014; BURNSTOCK, 2009; GUERRERO et al., 2008; MOGIL et al., 2000; NAKAMURA; FERREIRA, 1987; SCHOLZ; WOOLF, 2002; VERRI; CUNHA; PARADA; POOLE; et al., 2006; WOOLF; SALTER, 2000)

286

287 1.3 ESPÉCIES REATIVAS DE OXIGÊNIO E NADPH OXIDASE

288 O oxigênio é uma molécula fundamental para existência de vida complexa no
289 planeta. Seu acúmulo na atmosfera levou ao desenvolvimento de organismos
290 chamados aeróbios. Apesar de indispensável, seus efeitos acompanham uma
291 paradoxal toxicidade decorrente da formação de EROs. De fato, após o
292 aparecimento do oxigênio diversos micro-organismos sucumbiram frente essa
293 adversidade por não disporem de mecanismos de defesa adequados para tal.
294 Mantendo o paradoxo, as EROs desempenham papel fundamental na homeostase,
295 apresentando toxicidade quando não há controle adequado sobre sua produção ou
296 detoxificação. As EROs são formadas após a transferência de elétrons na cadeia
297 fosforilativa (mitocôndria) ou através do sistema da NADPH oxidase (JONES, 2008),
298 o qual é composto por proteínas que se encontram dissociadas na célula em
299 repouso. Esses componentes são a p40^{phox}, a p47^{phox} e a p67^{phox}, agrupadas em um
300 sistema protéico citoplasmático de 240kDa. Também há o citocromo b558, composto
301 pelas proteínas p22^{phox} e gp91^{phox} localizadas nas membranas das vesículas
302 secretórias e dos grânulos específicos citoplasmáticos. Outras proteínas de baixa
303 massa molecular, ligantes de nucleotídeo guanina: a Rac 1 e 2 e Rap1a, também
304 participam do processo (BEDARD; KRAUSE, 2007; JONES, 2008). Quando ativo, o
305 sistema multienzimático NADPH oxidase é responsável pela transferência de
306 elétrons do NADPH para o oxigênio molecular, formando o ânion superóxido
307 (BEDARD; KRAUSE, 2007), o qual é produzido em grande escala por neutrófilos e
308 tem proeminente atividade microbicida (BABIOR; KIPNES; CURNUTTE, 1973). Em
309 condições fisiológicas, o ânion superóxido é dismutado pela SOD em peróxido de
310 hidrogênio e esse é transformado em duas (2) molécula de água e uma (1) de
311 oxigênio (JONES, 2008).

312 As EROs são capazes de oxidar diversas proteínas, especialmente com
313 resíduos de cisteína (tióis), essa característica permite ativar diversas vias de
314 sinalização, entre elas a do NF-κB (GLOIRE; PIETTE, 2009). De fato, a via do NF-κB
315 é conhecida como sensível ao estado redox da célula (TOLEDANO; LEONARD,
316 1991). Após ubiquitinação e degradação do IκBα, resíduos específicos de serina da
317 subunidade p65 são fosforilados por PKA. Isso permite a interação de p65 com a
318 molécula coativadora do NF-κB (mas não apenas) CBP (proteína ligadora de CREB).
319 Essa fosforilação por PKA é elevada na presença de EROs e isso possibilita o
320 aumento da atividade do NF-κB. De fato, tratamento com antioxidantes diminui a

321 associação de p65 com CBP na região promotora do gene da IL-8, por exemplo
322 (JAMALUDDIN et al., 2007). Focando no ânion superóxido, tratamento com M40403
323 (molécula mimética da enzima SOD) inibiu a ativação do NF- κ B, relacionada com
324 diminuição da degradação do I κ B, sugerindo a participação do ânion superóxido da
325 regulação redox desse fator de transcrição (NDENGELE et al., 2005)

326

327 1.3.1 Relação Entre Ânion Superóxido E Dor

328 Existe uma linha tênue entre ânion superóxido e dor. Desse modo, é
329 necessário que haja uma sintonia fina para o controle adequado da produção de
330 EROs, especialmente do ânion superóxido. Em relação a dor, o ânion superóxido é
331 importante para o desenvolvimento de hiperalgesia térmica associada com
332 inflamação aguda e crônica (NDENGELE et al., 2008; WANG et al., 2004); e
333 promoção de hiperalgesia mecânica (KIM; CHUNG; CHUNG, 2008). Associado a
334 isso, o ânion superóxido é capaz de ativar canais TRPA1 (FERNANDES et al.,
335 2013), que são essenciais para dor (Tabela 1). Corroborando, o tratamento com
336 Tempol (molécula mimética da enzima SOD) foi capaz de inibir hiperalgesia
337 mecânica e inflamação em modelo de dor induzido por carragenina (KHATTAB,
338 2006).

339 Adicionalmente à sua atividade hiperalgésica, nosso grupo demonstrou –
340 utilizando o superóxido de potássio como um doador de ânion superóxido – que o
341 ânion superóxido é capaz de produzir comportamentos de dor manifesta, como
342 contorções abdominais e sacudidas de pata (MAIOLI et al., 2015). Além disso, o
343 ânion superóxido (injetado periféricamente) é capaz contribuir para sensibilização
344 espinal por aumentar a expressão de endotelina-1, TNF- α e IL-1 β (SERAFIM et al.,
345 2015; YAMACITA-BORIN et al., 2015). Corroborando esse fato, inibição desses
346 mediadores utilizando o bosentan (antagonista misto de receptor de endotelina),
347 etanercept (receptor solúvel de TNF- α) e camundongos knockout para TNFR1
348 apresentaram redução de comportamentos relacionados a dor, recrutamento de
349 neutrófilos, estresse oxidativo e produção de citocinas pró-inflamatórias (SERAFIM
350 et al., 2015; YAMACITA-BORIN et al., 2015). Esses dados apontam para a
351 interação fundamental entre o ânion superóxido e diversos mediadores e fatores de
352 transcrição, o que sugere o papel crucial dessa molécula no desenvolvimento de dor
353 e inflamação. Assim, estratégias que visem sua inibição extremamente atrativas.

354

355 1.4 ARTRITE

356 Artrite é um nome generalizado para doenças que afetam as articulações,
357 dentre elas se destacam: osteoartrite, artrite reumatoide (RA), artrite séptica e gota
358 (MARTIN; HARPER, 2010; SMOLEN; ALETAHA; REDLICH, 2012). No Brasil, a
359 prevalência de RA varia entre 0,2% e 1%, dependendo da região (DE AZEVEDO;
360 FERRAZ; CICONELLI, 2008). As doenças artríticas impactam significativamente nos
361 cofres públicos. De fato, em um estudo envolvendo 192 pacientes com RA, 47 deles
362 se aposentaram precocemente devido incapacidade físicas provenientes da doença.
363 Para essa população, o impacto orçamentário público foi de US\$ 2.423,51 por
364 paciente por ano (DE AZEVEDO et al., 2008). A dor, especificamente alodinia, é o
365 sintoma que converge a artrite. Desse modo, pacientes acometidos com os diversos
366 tipos de artrites apresentam redução brusca da qualidade de vida, já que atividades
367 normais diárias, como se movimentar, são excruciantes.

368 A revolução industrial foi o salto inicial para uma série de transformações
369 tecnológicas que sepultou regimes arcaicos de produção entorno do mundo. Isso
370 possibilitou uma nova era tecnológica. Com o avanço tecnológico, o prolongamento
371 da vida humana foi possível, devido a cura/erradicação de várias doenças
372 (poliomielite por exemplo) e com o paradoxal, porém inevitável, surgimento de
373 doenças que outrora não havia. Acompanhado a isso, o avanço tecnológico
374 propiciou a implementação de intervenções médicas para aumentar a qualidade de
375 vida de pacientes afetados com diversas doenças; dentre elas o uso de próteses
376 para substituir completamente ou parcialmente um membro. Em particular para
377 artrite, um desses procedimentos é a artroplastia – substituição total da articulação
378 por próteses (LEE; GOODMAN, 2008) – uma intervenção necessária devido
379 destruição das articulações acometidas. De fato, pacientes submetidos a artroplastia
380 apresentam melhor qualidade de vida quando comparados ao que não foram
381 submetidos à intervenção (DA SILVA et al., 2014; MARICONDA et al., 2011). Para
382 fabricação das próteses é utilizada uma variedade de materiais, incluindo aço
383 inoxidável, ligas de cobalto molibdênio, titânio e ligas de titânio (tal como titânio
384 alumínio e vanádio), polietileno entre outros; em uma tentativa de encontrar
385 materiais que são biocompatíveis e possuem as propriedades mecânicas
386 necessárias para suportar as forças aplicadas para a prótese. Muitos destes
387 materiais permanecem em uso hoje em dia (LEE; GOODMAN, 2008).

388 Infelizmente, 10-15% dos pacientes que utilizam próteses apresentam
389 osteólise decorrente de detritos liberados pelos materiais prostéticos (GOODMAN,
390 2007; HARRIS, 2001). Os relatos de osteólise datam de 1970 (CHARNLEY, 1975) e
391 tem sido associada com todos materiais utilizados em materiais prostéticos
392 (GOODMAN, 2007; HARRIS, 2001). A inflamação crônica oriunda do uso de
393 prótese está relacionada com a fagocitose de partículas liberadas por esses
394 materiais, causando necrose e osteólise (GOODMAN; MA, 2010). Dois principais
395 mecanismos colaboram para a destruição óssea: (I) a osteoclastogênese que ocorre
396 decorrente da sinalização RANK-RANKL (HAYNES et al., 2001) e (II) a fagocitose de
397 partículas liberadas pelos materiais prostéticos, causando liberação de enzimas
398 proteolíticas e citocinas pró-inflamatórias (GOODMAN; MA, 2010; TAKEI et al.,
399 2000).

400 Em particular, uma paciente, sem histórico de artrite familiar, desenvolveu
401 artrite relacionada a implante após a implementação de uma prótese. O biomaterial
402 em questão era uma liga de titânio e vanádio (ISO standard 5832-3:1996) (DORNER
403 et al., 2006). Após incubação de células mononucleares sanguíneas derivadas da
404 paciente com dióxido de titânio, foi observado aumento da produção de TNF- α , uma
405 citocina fundamental na fisiopatologia da artrite (DORNER et al., 2006). De fato, a
406 exposição pulmonar a dióxido de titânio em ratos, leva ao aumento da produção de
407 citocinas pró-inflamatórias (como IL-1 β , TNF- α e IL-6) acompanhado com maior
408 influxo de leucócitos e destruição tecidual (GUSTAFSSON et al., 2011).
409 Corroborando esses resultados, incubação com dióxido de titânio é capaz de
410 produzir dano no DNA e aumentar a atividade do NF- κ B (PRASAD, R. Y. et al.,
411 2014), além de ser capaz de aumentar liberação do citocromo C no citoplasma das
412 células culminando na ativação de caspase-3 e consequente apoptose dos
413 hepatócitos (EL-SAID et al., 2014). Esses resultados sugerem que as partículas de
414 dióxido de titânio são capazes de induzir estresse oxidativo, citotoxicidade,
415 genotoxicidade, inflamação crônica e em última instância apoptose e necrose
416 celular; associado a isso um relato de artrite induzida por implante. Contudo, os
417 benefícios decorrentes da utilização de próteses são extraordinários, necessitando,
418 portanto, de um controle da quantidade de dióxido de titânio implementado nesses
419 biomateriais acompanhado do desenvolvimento ou aplicação de moléculas com
420 atividade biológica capazes de controlar os efeitos deletérios provenientes da
421 liberação dessas partículas.

422 1.5 CURCUMINA

423 A Medicina Tradicional ainda é tida como uma fonte próspera de compostos
424 com atividades farmacológicas para a medicina contemporânea. Os extratos de
425 plantas têm sido utilizado por milhares de anos por praticantes de Medicina
426 Tradicional Chinesa e Indiana (Ayurvedica), mas apesar disso a aplicação desses
427 compostos ainda é vista com ceticismo por parte de praticantes da medicina
428 ocidental. Uma das principais característica e que tornam os compostos derivados
429 de produtos naturais extremamente atrativos, reside no fato de eles terem sidos
430 testados “clanicamente” durante milhares de anos por civilizações praticantes de
431 medicinas alternativas. A curcumina apresenta mais de 100 diferentes alvos
432 moleculares (AGGARWAL, B. B.; HARIKUMAR, 2009), e é um dos compostos que
433 se destaca dentre os derivados de produtos naturais. Sua importância farmacológica
434 é evidenciada através de inúmeras moléculas análogas e derivadas de sua estrutura
435 (AGRAWAL; MISHRA, 2010; MARTINEZ-CIFUENTES et al., 2015; PRASAD, S.;
436 TYAGI, 2015; ZHAO; LIU; LIANG, 2013).

437 A curcumina [(E,E)-1,7-bis(4-hidroxi-3-metoxifenil)-1,6-heptadieno-3,5-diona]
438 (Figura 3) é um composto de coloração amarela e o constituinte presente em maior
439 concentração no extrato do rizoma da planta *Curcuma longa* (açafrão). O extrato da
440 cúrcuma comercialmente disponível contém cerca de 77% de curcumina pura, 17%
441 dimetoxicurcumina e 3% bis-dimetoxicurcumina (SANDUR et al., 2007). Além de ser
442 utilizada como condimento alimentar, tem sido amplamente utilizada por praticante
443 da Medicina Tradicional Chinesa e Ayuvedica para o tratamento de diversas
444 enfermidades, como disenterias, flatulência, úlcera, feridas e artrite (AGGARWAL, B.
445 B.; SUNG, 2009). Essa molécula possui atividade anti-inflamatória, antioxidante,
446 anticancerígena, antiviral e antibacteriana (AGGARWAL, B. B.; HARIKUMAR, 2009;
447 AGGARWAL, B. B.; SUNG, 2009), além de ser extremamente segura. Administração
448 de curcumina por 4 semanas nas doses de 1-2 g/kg não demonstrou efeitos tóxicos
449 significativos. A LD₅₀ foi de 12,2 g/kg (ARORA et al., 1971). Em humanos, a dose
450 máxima utilizada e que não apresentou efeitos tóxicos foi de 8000 mg por dia
451 (CHENG et al., 2001). A curcumina tem atividade pleiotrópica, e aparentemente, não
452 apresenta um alvo específico. Ao invés disso, seu efeito farmacológico está
453 relacionado a um sinergismo envolvendo modulação de diversas moléculas em
454 diferentes vias de sinalização (Figura 3) (ZHOU, H.; BEEVERS; HUANG, 2011).
455 Esse pleiotropismo é proporcionado por 3 importantes funcionalidades em sua

456 molécula: (1) o anel aromático orto-metilado com um grupamento fenol, (2) a
457 presença de uma dicetona α , β insaturada e que possibilita (3) a formação de uma
458 molécula tautômera (co-existente) ceto-enol (ANAND et al., 2008). Essas
459 características únicas propiciam a ligação da curcumina com uma gama de alvos
460 moleculares. Sua atividade anti-inflamatória está relacionada interação direta com
461 diversos mediadores, como TNF- α , COX-1 e COX-2, por exemplo. A curcumina foi
462 capaz de reduzir a sinalização entre o TNF- α e seu receptor (WUA et al., 2010).
463 Essa interação foi possível já que a curcumina se liga (através de ligações iônicas)
464 em diversos resíduos específicos de aminoácidos (cisteína [Cys129], leucina
465 [Leu89] e ácido aspártico [Asp105]) presentes na molécula do TNF- α (WUA et al.,
466 2010). Em relação a COX-1 e COX-2, a curcumina se liga, entre outros, em resíduo
467 específico de serina (Ser535) e tirosina (Tyr355), respectivamente (SELVAM et al.,
468 2005). Além disso, a curcumina inibe atividade do NF- κ B e conseqüentemente
469 produção de citocinas pró-inflamatórias (AGGARWAL, S. et al., 2006), reduz a
470 expressão de moléculas de adesão (SUYENAGA et al., 2014), tem capacidade de
471 inibir a polimerização de actina em neutrófilos (LARMONIER et al., 2011) e reduz
472 estresse oxidativo, aumentando por exemplo atividade do fator de transcrição Nrf2 e
473 conseqüentemente HO-1 (GAO, S. et al., 2013; HEEBA; MAHMOUD; EL HANAFY,
474 2012). Além disso, a curcumina é capaz de reduzir osteoclastogênese através da
475 inibição da sinalização entre RANK e RANKL (OH; KYUNG; CHOI, 2008; ZHOU, T.
476 et al., 2013) (Figura 3).

477 A curcumina tem sido amplamente utilizada em diversos ensaios clínicos
478 (GOEL; KUNNUMAKKARA; AGGARWAL, 2008), porém iremos nos ater nos ensaios
479 clínicos que avaliaram sua atividade analgésica. O primeiro ensaio clínico
480 demonstrando sua atividade analgésica foi em 1980 em pacientes com artrite
481 reumatóide (DEODHAR; SETHI; SRIMAL, 1980). O tratamento com curcumina foi
482 capaz de reduzir edema e rigidez matinal das articulações, e aumentar o tempo de
483 caminhada dos pacientes sem queixas de dor (DEODHAR et al., 1980). Em outro
484 ensaio clínico, também em pacientes com artrite reumatóide, a curcumina foi capaz
485 de reduzir a dor reportada pelos pacientes, edema das articulações e sensibilidade no
486 local (CHANDRAN; GOEL, 2012). Nesse sentido, um medicamento derivado de
487 curcumina foi desenvolvido: Meriva[®]. Em diferentes ensaios clínicos em pacientes
488 com osteoartrite, o Meriva[®] reduziu a dor reportada pelos pacientes (escala de
489 VAS), rigidez matinal das articulações, produção de IL-6, IL-1 β e sVCAM (BELCARO

490 et al., 2010; DI PIERRO et al., 2013). Essas características (Figura 3) nos
 491 conduziram a avaliar a eficácia da curcumina frente a dois modelos: um modelo dor
 492 dor agudo induzido por ânion superóxido e em segundo momento em um modelo de
 493 dor crônica de artrite induzido por dióxido de titânio (TiO₂).
 494

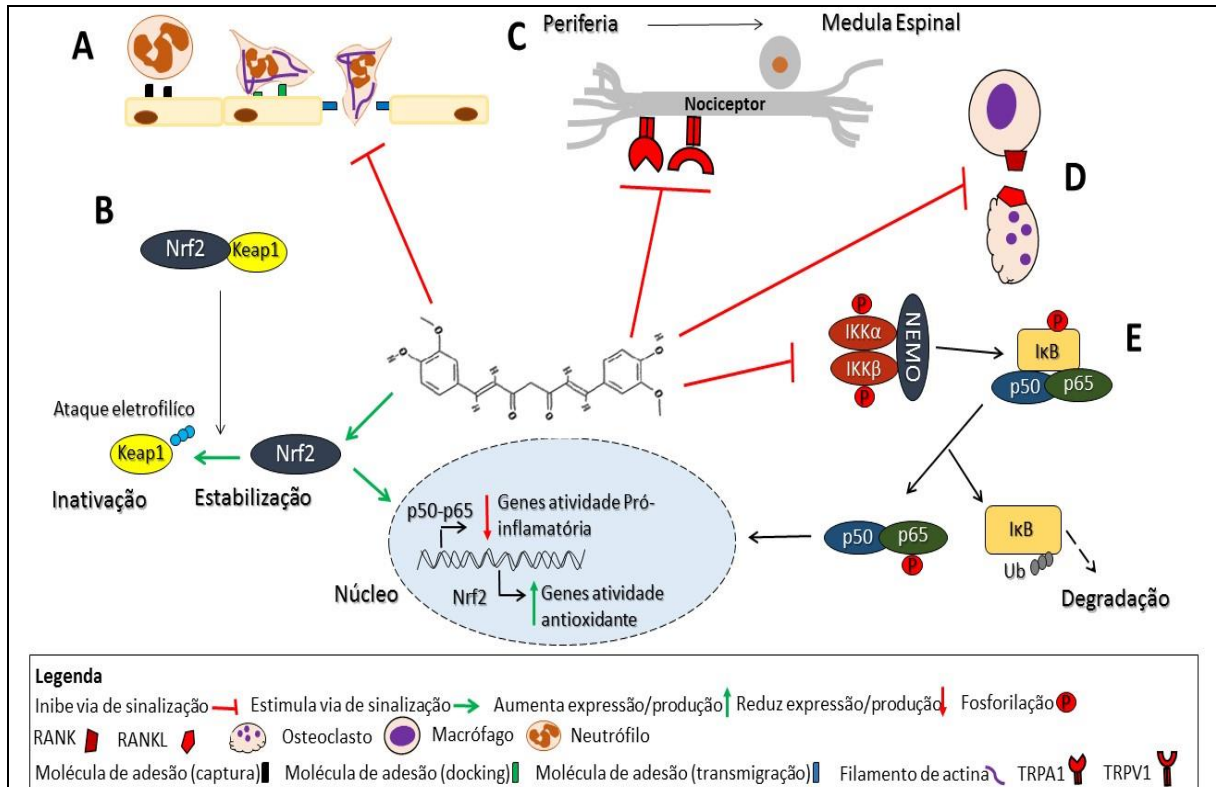


Figura 3. Alvos moleculares da curcumina. A curcumina inibe o recrutamento de neutrófilos e a expressão de moléculas de adesão (A). Além disso é capaz de aumentar a atividade do Nrf2 e seus genes alvos, como a HO-1 (B); e inibir a sinalização entre RANK e RANKL, diminuindo a osteólise (D); e inibir a atividade do NF-κB e conseqüentemente reduz a produção de mediadores pró-inflamatórios (E). Adicionalmente, a curcumina tem a capacidade de dessensibilizar os nociceptores (C) por inibir a atividade de canais iônicos associados a reconhecimento de estímulos dolorosos (Tabela 1). Em conjunto, essas características propiciam uma proeminente atividade anti-inflamatória e anti-nociceptiva para a curcumina (AGGARWAL, B. B.; HARIKUMAR, 2009; AGGARWAL, B. B.; SUNG, 2009; LEAMY et al., 2011; OH et al., 2008; YEON et al., 2010).

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501 **2 OBJETIVOS**

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503 2.1 OBJETIVO GERAL

504 Avaliar o efeito da curcumina em modelos de dor agudo (induzido por ânion
505 superóxido) e dor crônica (artrite induzida por dióxido de titânio).

506

507 2.1.1 Objetivos Específicos (Modelo Crônico)

508 -Determinar a melhor dose de curcumina em modelo de hiperalgesia
509 mecânica e edema; e degradação de proteoglicanos

510 -Avaliar o efeito da curcuina sobre atividade da mieloperoxidase

511

512 2.1.2 Objetivos Específicos (Modelo Agudo)

513 -Avaliar o efeito da curcumina, através de ensaio de dose-resposta, em
514 modelo de dor manifesta

515 -Avaliar o efeito da curcumina em modelo de hiperalgesia mecânica e térmica

516 -Avaliar o efeito da curcumina sobre o recrutamento de leucócitos

517 -Avaliar o efeito da curcumina sobre parâmetros relacionados ao estresse
518 oxidativo, Nrf2 e HO-1

519 -Avaliar o efeito da curcumina sobre produção de citocinas e atividade do NF-

520 κ B

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REFERÊNCIAS

- 529 AGGARWAL, B. B.; HARIKUMAR, K. B. Potential therapeutic effects of curcumin, the
530 anti-inflammatory agent, against neurodegenerative, cardiovascular, pulmonary,
531 metabolic, autoimmune and neoplastic diseases. **Int J Biochem Cell Biol**, v. 41, n.
532 1, p. 40-59, Jan 2009.
- 533 AGGARWAL, B. B.; SUNG, B. Pharmacological basis for the role of curcumin in
534 chronic diseases: an age-old spice with modern targets. **Trends Pharmacol Sci**, v.
535 30, n. 2, p. 85-94, Feb 2009.
- 536 AGGARWAL, S. et al. Curcumin (diferuloylmethane) down-regulates expression of
537 cell proliferation and antiapoptotic and metastatic gene products through suppression
538 of IkappaBalpha kinase and Akt activation. **Mol Pharmacol**, v. 69, n. 1, p. 195-206,
539 Jan 2006.
- 540 AGRAWAL, D. K.; MISHRA, P. K. Curcumin and its analogues: potential anticancer
541 agents. **Med Res Rev**, v. 30, n. 5, p. 818-60, Sep 2010.
- 542 ANAND, P. et al. Biological activities of curcumin and its analogues (Congeners)
543 made by man and Mother Nature. **Biochem Pharmacol**, v. 76, n. 11, p. 1590-611,
544 Dec 1 2008.
- 545 ARORA, R. B. et al. Anti-inflammatory studies on *Curcuma longa* (turmeric). **Indian**
546 **J Med Res**, v. 59, n. 8, p. 1289-95, Aug 1971.
- 547 BABIOR, B. M.; KIPNES, R. S.; CURNUTTE, J. T. Biological defense mechanisms.
548 The production by leukocytes of superoxide, a potential bactericidal agent. **J Clin**
549 **Invest**, v. 52, n. 3, p. 741-4, Mar 1973.
- 550 BEDARD, K.; KRAUSE, K. H. The NOX family of ROS-generating NADPH oxidases:
551 physiology and pathophysiology. **Physiol Rev**, v. 87, n. 1, p. 245-313, Jan 2007.
- 552 BELCARO, G. et al. Efficacy and safety of Meriva(R), a curcumin-
553 phosphatidylcholine complex, during extended administration in osteoarthritis
554 patients. **Altern Med Rev**, v. 15, n. 4, p. 337-44, Dec 2010.
- 555 BRAZ, J. et al. Transmitting pain and itch messages: a contemporary view of the
556 spinal cord circuits that generate gate control. **Neuron**, v. 82, n. 3, p. 522-36, May 7
557 2014.
- 558 BURNSTOCK, G. Purinergic receptors and pain. **Curr Pharm Des**, v. 15, n. 15, p.
559 1717-35, 2009.
- 560 BUSHNELL, M. C.; CEKO, M.; LOW, L. A. Cognitive and emotional control of pain
561 and its disruption in chronic pain. **Nat Rev Neurosci**, v. 14, n. 7, p. 502-11, Jul 2013.
- 562 CALATI, R. et al. The impact of physical pain on suicidal thoughts and behaviors:
563 Meta-analyses. **J Psychiatr Res**, v. 71, p. 16-32, Dec 2015.

- 564 CHANDRAN, B.; GOEL, A. A randomized, pilot study to assess the efficacy and
565 safety of curcumin in patients with active rheumatoid arthritis. **Phytother Res**, v. 26,
566 n. 11, p. 1719-25, Nov 2012.
- 567 CHARNLEY, J. Fracture of femoral prostheses in total hip replacement. A clinical
568 study. **Clin Orthop Relat Res**, n. 111, p. 105-20, Sep 1975.
- 569 CHEN, Z. J. Ubiquitin signalling in the NF-kappaB pathway. **Nat Cell Biol**, v. 7, n. 8,
570 p. 758-65, Aug 2005.
- 571 CHENG, A. L. et al. Phase I clinical trial of curcumin, a chemopreventive agent, in
572 patients with high-risk or pre-malignant lesions. **Anticancer Res**, v. 21, n. 4B, p.
573 2895-900, Jul-Aug 2001.
- 574 CHIU, I. M. et al. Bacteria activate sensory neurons that modulate pain and
575 inflammation. **Nature**, v. 501, n. 7465, p. 52-7, Sep 5 2013.
- 576 CUNHA, F. Q.; CACINI, A. T.; FERREIRA, S. H. Inhibition of the release of a
577 neutrophil chemotactic factor from macrophages partially explains the anti-
578 inflammatory action of glucocorticoids. **Agents Actions**, v. 17, n. 3-4, p. 314-7, Jan
579 1986.
- 580 CUNHA, T. M. et al. Crucial role of neutrophils in the development of mechanical
581 inflammatory hypernociception. **J Leukoc Biol**, v. 83, n. 4, p. 824-32, Apr 2008.
- 582 CUNHA, T. M. et al. A cascade of cytokines mediates mechanical inflammatory
583 hypernociception in mice. **Proc Natl Acad Sci U S A**, v. 102, n. 5, p. 1755-60, Feb 1
584 2005.
- 585 DA SILVA, R. R. et al. Quality of life after total knee arthroplasty: systematic review.
586 **Rev Bras Ortop**, v. 49, n. 5, p. 520-7, Sep-Oct 2014.
- 587 DE AZEVEDO, A. B.; FERRAZ, M. B.; CICONELLI, R. M. Indirect costs of
588 rheumatoid arthritis in Brazil. **Value Health**, v. 11, n. 5, p. 869-77, Sep-Oct 2008.
- 589 DE RIVERO VACCARI, J. P. et al. A molecular platform in neurons regulates
590 inflammation after spinal cord injury. **J Neurosci**, v. 28, n. 13, p. 3404-14, Mar 26
591 2008.
- 592 DEODHAR, S. D.; SETHI, R.; SRIMAL, R. C. Preliminary study on antirheumatic
593 activity of curcumin (diferuloyl methane). **Indian J Med Res**, v. 71, p. 632-4, Apr
594 1980.
- 595 DI PIERRO, F. et al. Comparative evaluation of the pain-relieving properties of a
596 lecithinized formulation of curcumin (Meriva((R))), nimesulide, and acetaminophen. **J
597 Pain Res**, v. 6, p. 201-5, 2013.
- 598 DORNER, T. et al. Implant-related inflammatory arthritis. **Nat Clin Pract
599 Rheumatol**, v. 2, n. 1, p. 53-6; quiz 57, Jan 2006.

- 600 EL-SAID, K. S. et al. Molecular mechanism of DNA damage induced by titanium
601 dioxide nanoparticles in toll-like receptor 3 or 4 expressing human hepatocarcinoma
602 cell lines. **J Nanobiotechnology**, v. 12, p. 48, 2014.
- 603 FERNANDES, E. S. et al. Superoxide generation and leukocyte accumulation: key
604 elements in the mediation of leukotriene B(4)-induced itch by transient receptor
605 potential ankyrin 1 and transient receptor potential vanilloid 1. **FASEB J**, v. 27, n. 4,
606 p. 1664-73, Apr 2013.
- 607 FERREIRA, S. H.; ROMITELLI, M.; DE NUCCI, G. Endothelin-1 participation in overt
608 and inflammatory pain. **J Cardiovasc Pharmacol**, v. 13 Suppl 5, p. S220-2, 1989.
- 609 FOXMAN, E. F.; CAMPBELL, J. J.; BUTCHER, E. C. Multistep navigation and the
610 combinatorial control of leukocyte chemotaxis. **J Cell Biol**, v. 139, n. 5, p. 1349-60,
611 Dec 1 1997.
- 612 FREITAS, A. et al. Heme oxygenase/carbon monoxide-biliverdin pathway down
613 regulates neutrophil rolling, adhesion and migration in acute inflammation. **Br J**
614 **Pharmacol**, v. 149, n. 4, p. 345-54, Oct 2006.
- 615 GAO, B.; DOAN, A.; HYBERTSON, B. M. The clinical potential of influencing Nrf2
616 signaling in degenerative and immunological disorders. **Clin Pharmacol**, v. 6, p. 19-
617 34, 2014.
- 618 GAO, S. et al. Curcumin attenuates arsenic-induced hepatic injuries and oxidative
619 stress in experimental mice through activation of Nrf2 pathway, promotion of arsenic
620 methylation and urinary excretion. **Food Chem Toxicol**, v. 59, p. 739-47, Sep 2013.
- 621 GLOIRE, G.; PIETTE, J. Redox regulation of nuclear post-translational modifications
622 during NF-kappaB activation. **Antioxid Redox Signal**, v. 11, n. 9, p. 2209-22, Sep
623 2009.
- 624 GOEL, A.; KUNNUMAKKARA, A. B.; AGGARWAL, B. B. Curcumin as "Curecumin":
625 from kitchen to clinic. **Biochem Pharmacol**, v. 75, n. 4, p. 787-809, Feb 15 2008.
- 626 GOODMAN, S. B. Wear particles, periprosthetic osteolysis and the immune system.
627 **Biomaterials**, v. 28, n. 34, p. 5044-8, Dec 2007.
- 628 GOODMAN, S. B.; MA, T. Cellular chemotaxis induced by wear particles from joint
629 replacements. **Biomaterials**, v. 31, n. 19, p. 5045-50, Jul 2010.
- 630 GUERRERO, A. T. et al. Involvement of LTB4 in zymosan-induced joint nociception
631 in mice: participation of neutrophils and PGE2. **J Leukoc Biol**, v. 83, n. 1, p. 122-30,
632 Jan 2008.
- 633 GUSTAFSSON, A. et al. Lung exposure of titanium dioxide nanoparticles induces
634 innate immune activation and long-lasting lymphocyte response in the Dark Agouti
635 rat. **J Immunotoxicol**, v. 8, n. 2, p. 111-21, Jun 2011.
- 636 HARRIS, W. H. Wear and periprosthetic osteolysis: the problem. **Clin Orthop Relat**
637 **Res**, n. 393, p. 66-70, Dec 2001.

- 638 HAYDEN, M. S.; GHOSH, S. Shared principles in NF-kappaB signaling. **Cell**, v. 132,
639 n. 3, p. 344-62, Feb 8 2008.
- 640 HAYNES, D. R. et al. The osteoclastogenic molecules RANKL and RANK are
641 associated with periprosthetic osteolysis. **J Bone Joint Surg Br**, v. 83, n. 6, p. 902-
642 11, Aug 2001.
- 643 HEEBA, G. H.; MAHMOUD, M. E.; EL HANAFY, A. A. Anti-inflammatory potential of
644 curcumin and quercetin in rats: Role of oxidative stress, heme oxygenase-1 and
645 TNF-alpha. **Toxicol Ind Health**, Sep 28 2012.
- 646 HUANG, H. C.; NGUYEN, T.; PICKETT, C. B. Phosphorylation of Nrf2 at Ser-40 by
647 protein kinase C regulates antioxidant response element-mediated transcription. **J**
648 **Biol Chem**, v. 277, n. 45, p. 42769-74, Nov 8 2002.
- 649 ITOH, K. et al. An Nrf2/small Maf heterodimer mediates the induction of phase II
650 detoxifying enzyme genes through antioxidant response elements. **Biochem**
651 **Biophys Res Commun**, v. 236, n. 2, p. 313-22, Jul 18 1997.
- 652 IWASAKI, A.; MEDZHITOV, R. Regulation of adaptive immunity by the innate
653 immune system. **Science**, v. 327, n. 5963, p. 291-5, Jan 15 2010.
- 654 JAMALUDDIN, M. et al. TNF-alpha-induced NF-kappaB/RelA Ser(276)
655 phosphorylation and enhanceosome formation is mediated by an ROS-dependent
656 PKAc pathway. **Cell Signal**, v. 19, n. 7, p. 1419-33, Jul 2007.
- 657 JANES, K.; NEUMANN, W. L.; SALVEMINI, D. Anti-superoxide and anti-peroxynitrite
658 strategies in pain suppression. **Biochim Biophys Acta**, v. 1822, n. 5, p. 815-21, May
659 2012.
- 660 JONES, D. P. Radical-free biology of oxidative stress. **Am J Physiol Cell Physiol**, v.
661 295, n. 4, p. C849-68, Oct 2008.
- 662 JONSSON, H.; ALLEN, P.; PENG, S. L. Inflammatory arthritis requires Foxo3a to
663 prevent Fas ligand-induced neutrophil apoptosis. **Nat Med**, v. 11, n. 6, p. 666-71, Jun
664 2005.
- 665 KAPITULNIK, J. Bilirubin: an endogenous product of heme degradation with both
666 cytotoxic and cytoprotective properties. **Mol Pharmacol**, v. 66, n. 4, p. 773-9, Oct
667 2004.
- 668 KARIN, M.; BEN-NERIAH, Y. Phosphorylation meets ubiquitination: the control of
669 NF-[kappa]B activity. **Annu Rev Immunol**, v. 18, p. 621-63, 2000.
- 670 KHATTAB, M. M. TEMPOL, a membrane-permeable radical scavenger, attenuates
671 peroxynitrite- and superoxide anion-enhanced carrageenan-induced paw edema and
672 hyperalgesia: a key role for superoxide anion. **Eur J Pharmacol**, v. 548, n. 1-3, p.
673 167-73, Oct 24 2006.
- 674 KIM, H. Y.; CHUNG, J. M.; CHUNG, K. Increased production of mitochondrial
675 superoxide in the spinal cord induces pain behaviors in mice: the effect of

- 676 mitochondrial electron transport complex inhibitors. **Neurosci Lett**, v. 447, n. 1, p.
677 87-91, Dec 5 2008.
- 678 LARMONIER, C. B. et al. Modulation of neutrophil motility by curcumin: implications
679 for inflammatory bowel disease. **Inflamm Bowel Dis**, v. 17, n. 2, p. 503-15, Feb
680 2011.
- 681 LAWRENCE, T. The nuclear factor NF-kappaB pathway in inflammation. **Cold
682 Spring Harb Perspect Biol**, v. 1, n. 6, p. a001651, Dec 2009.
- 683 LAWRENCE, T.; WILLOUGHBY, D. A.; GILROY, D. W. Anti-inflammatory lipid
684 mediators and insights into the resolution of inflammation. **Nat Rev Immunol**, v. 2, n.
685 10, p. 787-95, Oct 2002.
- 686 LEAMY, A. W. et al. Curcumin ((E,E)-1,7-bis(4-hydroxy-3-methoxyphenyl)-1,6-
687 heptadiene-3,5-dione) activates and desensitizes the nociceptor ion channel TRPA1.
688 **Neurosci Lett**, v. 503, n. 3, p. 157-62, Oct 10 2011.
- 689 LEE, K.; GOODMAN, S. B. Current state and future of joint replacements in the hip
690 and knee. **Expert Rev Med Devices**, v. 5, n. 3, p. 383-93, May 2008.
- 691 LEVY, B. D. et al. Lipid mediator class switching during acute inflammation: signals
692 in resolution. **Nat Immunol**, v. 2, n. 7, p. 612-9, Jul 2001.
- 693 LI, W. et al. Heterodimerization with small Maf proteins enhances nuclear retention
694 of Nrf2 via masking the NESzip motif. **Biochim Biophys Acta**, v. 1783, n. 10, p.
695 1847-56, Oct 2008.
- 696 LIU, T. et al. Toll-like receptor 7 mediates pruritus. **Nat Neurosci**, v. 13, n. 12, p.
697 1460-2, Dec 2010.
- 698 MADERNA, P.; GODSON, C. Lipoxins: revolutionary road. **Br J Pharmacol**, v. 158,
699 n. 4, p. 947-59, Oct 2009.
- 700 MAIOLI, N. A. et al. The superoxide anion donor, potassium superoxide, induces
701 pain and inflammation in mice through production of reactive oxygen species and
702 cyclooxygenase-2. **Braz J Med Biol Res**, v. 0, p. 0, Feb 13 2015.
- 703 MARIATHASAN, S. et al. Cryopyrin activates the inflammasome in response to
704 toxins and ATP. **Nature**, v. 440, n. 7081, p. 228-32, Mar 9 2006.
- 705 MARICONDA, M. et al. Quality of life and functionality after total hip arthroplasty: a
706 long-term follow-up study. **BMC Musculoskelet Disord**, v. 12, p. 222, 2011.
- 707 MARTIN, W. J.; HARPER, J. L. Innate inflammation and resolution in acute gout.
708 **Immunol Cell Biol**, v. 88, n. 1, p. 15-9, Jan 2010.
- 709 MARTINEZ-CIFUENTES, M. et al. Heterocyclic Curcumin Derivatives of
710 Pharmacological Interest: Recent Progress. **Curr Top Med Chem**, v. 15, n. 17, p.
711 1663-72, 2015.

- 712 MARZEC, J. M. et al. Functional polymorphisms in the transcription factor NRF2 in
713 humans increase the risk of acute lung injury. **FASEB J**, v. 21, n. 9, p. 2237-46, Jul
714 2007.
- 715 MCDONALD, B. et al. Intravascular danger signals guide neutrophils to sites of
716 sterile inflammation. **Science**, v. 330, n. 6002, p. 362-6, Oct 15 2010.
- 717 MILLAN, M. J. The induction of pain: an integrative review. **Prog Neurobiol**, v. 57, n.
718 1, p. 1-164, Jan 1999.
- 719 _____. Descending control of pain. **Prog Neurobiol**, v. 66, n. 6, p. 355-474, Apr
720 2002.
- 721 MOAYEDI, M.; DAVIS, K. D. Theories of pain: from specificity to gate control. **J**
722 **Neurophysiol**, v. 109, n. 1, p. 5-12, Jan 2013.
- 723 MOGIL, J. S.; YU, L.; BASBAUM, A. I. Pain genes?: natural variation and transgenic
724 mutants. **Annu Rev Neurosci**, v. 23, p. 777-811, 2000.
- 725 NAKAMURA, M.; FERREIRA, S. H. A peripheral sympathetic component in
726 inflammatory hyperalgesia. **Eur J Pharmacol**, v. 135, n. 2, p. 145-53, Mar 17 1987.
- 727 NDENGELE, M. M. et al. Cyclooxygenases 1 and 2 contribute to peroxynitrite-
728 mediated inflammatory pain hypersensitivity. **FASEB J**, v. 22, n. 9, p. 3154-64, Sep
729 2008.
- 730 NDENGELE, M. M. et al. Superoxide potentiates NF-kappaB activation and
731 modulates endotoxin-induced cytokine production in alveolar macrophages. **Shock**,
732 v. 23, n. 2, p. 186-93, Feb 2005.
- 733 NITURE, S. K.; JAIN, A. K.; JAISWAL, A. K. Antioxidant-induced modification of INrf2
734 cysteine 151 and PKC-delta-mediated phosphorylation of Nrf2 serine 40 are both
735 required for stabilization and nuclear translocation of Nrf2 and increased drug
736 resistance. **J Cell Sci**, v. 122, n. Pt 24, p. 4452-64, Dec 15 2009.
- 737 NITURE, S. K.; KHATRI, R.; JAISWAL, A. K. Regulation of Nrf2-an update. **Free**
738 **Radic Biol Med**, v. 66, p. 36-44, Jan 2014.
- 739 OH, S.; KYUNG, T. W.; CHOI, H. S. Curcumin inhibits osteoclastogenesis by
740 decreasing receptor activator of nuclear factor-kappaB ligand (RANKL) in bone
741 marrow stromal cells. **Mol Cells**, v. 26, n. 5, p. 486-9, Nov 30 2008.
- 742 PAE, H. O. et al. Role of heme oxygenase in preserving vascular bioactive NO.
743 **Nitric Oxide**, v. 23, n. 4, p. 251-7, Dec 15 2010.
- 744 PRASAD, R. Y. et al. Cellular interactions and biological responses to titanium
745 dioxide nanoparticles in HepG2 and BEAS-2B cells: role of cell culture media.
746 **Environ Mol Mutagen**, v. 55, n. 4, p. 336-42, May 2014.
- 747 PRASAD, S.; TYAGI, A. K. Curcumin and its analogues: a potential natural
748 compound against HIV infection and AIDS. **Food Funct**, v. 6, n. 11, p. 3412-9, Nov 4
749 2015.

- 750 QI, J. et al. Painful pathways induced by TLR stimulation of dorsal root ganglion
751 neurons. **J Immunol**, v. 186, n. 11, p. 6417-26, Jun 1 2011.
- 752 REICHLING, D. B.; LEVINE, J. D. Critical role of nociceptor plasticity in chronic pain.
753 **Trends Neurosci**, v. 32, n. 12, p. 611-8, Dec 2009.
- 754 RIBEIRO, R. A. et al. Role of resident mast cells and macrophages in the neutrophil
755 migration induced by LTB₄, fMLP and C5a des arg. **Int Arch Allergy Immunol**, v.
756 112, n. 1, p. 27-35, Jan 1997.
- 757 ROCHA E SILVA, M. A brief survey of the history of inflammation. **Agents Actions**,
758 v. 8, n. 1-2, p. 45-9, Jan 1978.
- 759 SANDUR, S. K. et al. Curcumin, demethoxycurcumin, bisdemethoxycurcumin,
760 tetrahydrocurcumin and turmerones differentially regulate anti-inflammatory and anti-
761 proliferative responses through a ROS-independent mechanism. **Carcinogenesis**, v.
762 28, n. 8, p. 1765-73, Aug 2007.
- 763 SCHOLZ, J.; WOOLF, C. J. Can we conquer pain? **Nat Neurosci**, v. 5 Suppl, p.
764 1062-7, Nov 2002.
- 765 SELVAM, C. et al. Design, synthesis, biological evaluation and molecular docking of
766 curcumin analogues as antioxidant, cyclooxygenase inhibitory and anti-inflammatory
767 agents. **Bioorg Med Chem Lett**, v. 15, n. 7, p. 1793-7, Apr 1 2005.
- 768 SEN, R.; SMALE, S. T. Selectivity of the NF- κ B response. **Cold Spring Harb**
769 **Perspect Biol**, v. 2, n. 4, p. a000257, Apr 2010.
- 770 SERAFIM, K. G. et al. Bosentan, a mixed endothelin receptor antagonist, inhibits
771 superoxide anion-induced pain and inflammation in mice. **Naunyn Schmiedeberg's**
772 **Arch Pharmacol**, Aug 6 2015.
- 773 SERHAN, C. N. et al. Lipid mediators in the resolution of inflammation. **Cold Spring**
774 **Harb Perspect Biol**, v. 7, n. 2, p. a016311, Feb 2015.
- 775 SMOLEN, J. S.; ALETAHA, D.; REDLICH, K. The pathogenesis of rheumatoid
776 arthritis: new insights from old clinical data? **Nat Rev Rheumatol**, v. 8, n. 4, p. 235-
777 43, Apr 2012.
- 778 SREERAMKUMAR, V. et al. Neutrophils scan for activated platelets to initiate
779 inflammation. **Science**, v. 346, n. 6214, p. 1234-8, Dec 5 2014.
- 780 STAURENGO-FERRARI, L. et al. The ruthenium nitric oxide donor,
781 [Ru(HEDTA)NO], inhibits acute nociception in mice by modulating oxidative stress,
782 cytokine production and activating the cGMP/PKG/ATP-sensitive potassium channel
783 signaling pathway. **Naunyn Schmiedeberg's Arch Pharmacol**, Aug 13 2014.
- 784 STEINER, A. A. et al. Role of the haeme oxygenase/carbon monoxide pathway in
785 mechanical nociceptor hypersensitivity. **Br J Pharmacol**, v. 132, n. 8, p. 1673-82,
786 Apr 2001.

- 787 SUYENAGA, E. S. et al. Beyond organoleptic characteristics: the pharmacological
788 potential of flavonoids and their role in leukocyte migration and in L-selectin and
789 beta2-integrin expression during inflammation. **Phytother Res**, v. 28, n. 9, p. 1406-
790 11, Sep 2014.
- 791 TAKEI, I. et al. Messenger ribonucleic acid expression of 16 matrix
792 metalloproteinases in bone-implant interface tissues of loose artificial hip joints. **J**
793 **Biomed Mater Res**, v. 52, n. 4, p. 613-20, Dec 15 2000.
- 794 TAKEUCHI, O.; AKIRA, S. Pattern recognition receptors and inflammation. **Cell**, v.
795 140, n. 6, p. 805-20, Mar 19 2010.
- 796 TING, E. et al. Role of complement C5a in mechanical inflammatory
797 hypernociception: potential use of C5a receptor antagonists to control inflammatory
798 pain. **Br J Pharmacol**, v. 153, n. 5, p. 1043-53, Mar 2008.
- 799 TODD, A. J. Neuronal circuitry for pain processing in the dorsal horn. **Nat Rev**
800 **Neurosci**, v. 11, n. 12, p. 823-36, Dec 2010.
- 801 TOLEDANO, M. B.; LEONARD, W. J. Modulation of transcription factor NF-kappa B
802 binding activity by oxidation-reduction in vitro. **Proc Natl Acad Sci U S A**, v. 88, n.
803 10, p. 4328-32, May 15 1991.
- 804 TURPAEV, K. T. Keap1-Nrf2 signaling pathway: mechanisms of regulation and role
805 in protection of cells against toxicity caused by xenobiotics and electrophiles.
806 **Biochemistry (Mosc)**, v. 78, n. 2, p. 111-26, Feb 2013.
- 807 VERRI, W. A., JR. et al. Targeting endothelin ETA and ETB receptors inhibits
808 antigen-induced neutrophil migration and mechanical hypernociception in mice.
809 **Naunyn Schmiedebergs Arch Pharmacol**, v. 379, n. 3, p. 271-9, Mar 2009.
- 810 VERRI, W. A., JR. et al. Hypernociceptive role of cytokines and chemokines: targets
811 for analgesic drug development? **Pharmacol Ther**, v. 112, n. 1, p. 116-38, Oct 2006.
- 812 VERRI, W. A., JR. et al. IL-15 mediates immune inflammatory hypernociception by
813 triggering a sequential release of IFN-gamma, endothelin, and prostaglandin. **Proc**
814 **Natl Acad Sci U S A**, v. 103, n. 25, p. 9721-5, Jun 20 2006.
- 815 WANG, Z. Q. et al. A newly identified role for superoxide in inflammatory pain. **J**
816 **Pharmacol Exp Ther**, v. 309, n. 3, p. 869-78, Jun 2004.
- 817 WIECH, K. et al. Influence of prior information on pain involves biased perceptual
818 decision-making. **Curr Biol**, v. 24, n. 15, p. R679-81, Aug 4 2014.
- 819 WOOLF, C. J.; SALTER, M. W. Neuronal plasticity: increasing the gain in pain.
820 **Science**, v. 288, n. 5472, p. 1765-9, Jun 9 2000.
- 821 WRIGHT, H. L. et al. Neutrophil function in inflammation and inflammatory diseases.
822 **Rheumatology (Oxford)**, v. 49, n. 9, p. 1618-31, Sep 2010.

- 823 WUA, S. T. et al. Docking Prediction for Tumor Necrosis Factor- α and Five Herbal
824 Inhibitors. **International Journal of Engineering Science and Technology**, v. 2, n.
825 9, p. 4263-4277, 2010.
- 826 YAMACITA-BORIN, F. Y. et al. Superoxide anion-induced pain and inflammation
827 depends on TNF α /TNFR1 signaling in mice. **Neurosci Lett**, v. 605, p. 53-8, Sep
828 25 2015.
- 829 YEON, K. Y. et al. Curcumin produces an antihyperalgesic effect via antagonism of
830 TRPV1. **J Dent Res**, v. 89, n. 2, p. 170-4, Feb 2010.
- 831 ZARPELON, A. C. et al. IL-33/ST2 signalling contributes to carrageenin-induced
832 innate inflammation and inflammatory pain: role of cytokines, endothelin-1 and
833 prostaglandin E2. **Br J Pharmacol**, v. 169, n. 1, p. 90-101, May 2013.
- 834 ZARPELON, A. C. et al. Endothelin-1 induces neutrophil recruitment in adaptive
835 inflammation via TNF α and CXCL1/CXCR2 in mice. **Can J Physiol Pharmacol**,
836 v. 90, n. 2, p. 187-99, Feb 2012.
- 837 ZARPELON, A. C. et al. The nitroxyl donor, Angeli's salt, inhibits inflammatory
838 hyperalgesia in rats. **Neuropharmacology**, v. 71, p. 1-9, Aug 2013.
- 839 ZHAO, C.; LIU, Z.; LIANG, G. Promising curcumin-based drug design: mono-
840 carbonyl analogues of curcumin (MACs). **Curr Pharm Des**, v. 19, n. 11, p. 2114-35,
841 2013.
- 842 ZHOU, H.; BEEVERS, C. S.; HUANG, S. The targets of curcumin. **Curr Drug**
843 **Targets**, v. 12, n. 3, p. 332-47, Mar 1 2011.
- 844 ZHOU, T. et al. Curcumin inhibits inflammatory response and bone loss during
845 experimental periodontitis in rats. **Acta Odontol Scand**, v. 71, n. 2, p. 349-56, Mar
846 2013.
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859 3 ARTIGO 1 PUBLICADO (PHARMACOLOGICAL RESEARCH)

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861 O presente artigo de revisão foi realizado no Laboratório de Dor, Inflamação,
862 Neuropatia e Câncer, da Universidade Estadual de Londrina e segue as normas da
863 revista Pharmacological Research e estão descritos no artigo intitulado “Neutrophils
864 and Arthritis: Role in Disease and Pharmacological Perspectives” (doi:
865 10.1016/j.phrs.2016.01.027)

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886 **Neutrophils and Arthritis: Role in Disease and Pharmacological Perspectives**

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912 **Abstract**

913

914 The inflammatory response in the joint can induce an intense accumulation of leukocytes in
915 the tissue that frequently results in severe local damage and loss of function. Neutrophils are
916 essential cells to combat many pathogens, but their arsenal can contribute or aggravate
917 articular inflammation. Here we summarized some aspects of neutrophil biology, their role in
918 inflammation and indicated how the modulation of neutrophil functions could be useful for
919 the treatment of different forms of arthritis.

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922 **Keywords:** Neutrophil, phagocytosis; neutrophil extracellular traps; arthritis; neutrophil
923 recruitment; infection; tissue damage

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946 **1) Introduction**

947 Rheumatic diseases, represented by varied forms of arthritis and other musculoskeletal
948 disorders, affect millions of people around the world and are currently one of the most studied
949 diseases in many centers of research [1]. There are variable and well-established models to
950 study different types of arthritis, most of them in mice, which give us valuable information
951 about the mechanisms that regulate the production of mediators of inflammation, cellular
952 infiltration in the joint and tissue damage and dysfunction. Furthermore, these proof-of-
953 concept models are useful for pre-clinical studies during the development of new anti-
954 inflammatory and anti-rheumatic drugs [2]. However, although useful, these models do not
955 represent the authentic pathogenesis in humans, which can cause failures during translational
956 studies. Thus, there are many challenges for the study of arthritis, especially on basic
957 research, and being conscious of the limitation of arthritis models is only the first step for a
958 better interpretation of the findings.

959 Neutrophils are crucial cells that have significant roles in virtually all inflammatory
960 diseases, ranging from acute, chronic, autoimmune, infectious, and non-infectious conditions
961 [3]. The most known effector functions of neutrophils are related to their role in innate
962 immunity since different chemoattractants quickly recruit neutrophils from the bloodstream.
963 However, recent research points out neutrophils as active cells during adaptive immunity,
964 facilitating the recruitment and activation of antigen-presenting cells or due to a direct
965 interaction with T cells [4,5].

966 Neutrophils have an arsenal of antimicrobial molecules that are essential to combat
967 several microorganisms. In this regard, the neutropenia or malfunctioning of neutrophils is
968 associated with the development of opportunistic infectious diseases. On the other hand,
969 several of the neutrophil-derived molecules that are crucial for the control of pathogens are
970 detrimental to the host tissue. Especially in autoimmune and autoinflammatory disorders, the
971 accumulation of neutrophils causes tissue damage and dysfunction, sometimes irreversible
972 [4]. Thus, the development of compounds that interfere on neutrophil biology is useful for the
973 treatment of many inflammatory diseases. Particularly in rheumatology, the infiltration of
974 neutrophils is directly associated with the worsening of the clinical condition, including
975 cartilage and bone destruction and pain in the different forms of arthritis [6]. Here, we
976 explored the recent findings in the field of neutrophil biology, its association with arthritis and
977 possible pharmacological approaches focusing on the biology of neutrophils.

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979

980 2) Neutrophil biology

981 Neutrophils represent one of the most important effector cells during innate immune
982 response. The mechanisms by which neutrophils deal with infection and their contribution to
983 tissue damage are discussed below in separated sections. In humans, these cells represent the
984 most abundant leukocytes in the blood, a variable that changes considerably among the
985 species [7]. Importantly, there is a rapid mobilization of stored neutrophils from the bone
986 marrow to the circulation during infectious/inflammatory conditions [8,9]. Independent on the
987 species, neutrophils quickly migrate from blood to the tissue under different chemoattractants
988 during inflammation, where they have an important role in the control of infections and
989 contribute to tissue repair. During the maturation of neutrophils, much of their arsenal of
990 molecules filled different types of granules, including primary or azurophilic, secondary or
991 specific, and tertiary or gelatinase. Section 4 addresses the content of each granule in detail
992 (Killing mechanisms of neutrophils in host defense) [For review see 10,11]. Adult humans
993 produce billions of neutrophils daily mainly by the actions of the cytokine Granulocyte-
994 Colony Stimulating Factor (G-CSF) on progenitor stem cells. In fact, the infusion of G-CSF is
995 clinically used to increase the neutrophil count in the peripheral blood of patients undergoing
996 myelosuppressive chemotherapy for acute myeloid leukemia or severe chronic neutropenia
997 [12]. Currently, a paradigm in the field of neutrophil biology is its lifespan, considered short
998 when compared to other leukocytes. Using *in vivo* labeling neutrophils, Pillay and colleagues
999 found that human neutrophils present a lifespan of 5.4 days on circulation while mouse
1000 neutrophils survive up to 12.5 hours in physiologic conditions [13].

1001 Some studies indicate that there is a heterogeneous population of neutrophils that
1002 could represent different phenotypes of these cells during an inflammatory response. Those
1003 variations depend on their morphology, cell surface markers, secreted molecules and density
1004 [For review see 14]. However, these subsets of neutrophils could be only a natural variation
1005 during their lifespan, considering the immature, mature and aged neutrophils and according to
1006 their state of activation. *In vitro* studies suggest that aged neutrophils decrease their ability to
1007 migrate and produce less pro-inflammatory molecules [15]. However, it was recently
1008 demonstrated that aged neutrophils in circulation display pro-inflammatory activity in mice
1009 [16]. Interestingly, neutrophil aging depends on the microbiota, since microbiota-depleted
1010 mice have reduced the circulation of aged neutrophils. Moreover, the decrease of a
1011 commensal population by the treatment with antibiotics ameliorates organ damage in mouse
1012 models of inflammation [16].

1013 Senescent neutrophils express high levels of CXCR4 (the ligand CXCL12 is highly
1014 expressed by bone marrow stromal cells), a process that is essential for the home back to the
1015 bone marrow where the apoptotic neutrophils are phagocytosed by macrophages, ending their
1016 life cycle [9] (Figure 1). *In vitro*, peripheral blood human neutrophils increase their expression
1017 of CXCR4 around 3 h at 37°C, an event that anticipates the apoptosis [17]. Importantly, there
1018 is an established concept that the apoptosis of neutrophils on inflammatory milieu is an
1019 important signal for the resolution of inflammation. The induction of neutrophil apoptosis by
1020 administration of H₂O₂ leads to the resolution of joint inflammation in antigen-induced
1021 arthritis in mice [18]. Mechanistically, the efferocytosis of apoptotic neutrophils by
1022 macrophages changes the profile of lipid mediators produced by these last cells, leading to the
1023 production of molecules with anti-inflammatory and pro-resolution properties, so-called
1024 specialized pro-resolving mediators (SPM) [For review see 19] (Figure 1).

1025 In addition to the neutrophil function in innate immunity, recent evidence points out
1026 their significant contribution to an adequate activation of adaptive immune response. Stored
1027 molecules in neutrophils, such as myeloperoxidase and proteinase-3, are potential
1028 autoantigens in some diseases and can lead to the development of anti-neutrophil cytoplasmic
1029 autoantibodies (ANCA) that mediate the chronic inflammation of some vasculitis [20].
1030 Moreover, some studies demonstrate that neutrophils can express MHC-II, the co-stimulatory
1031 molecules CD86, CD40, and CCR7, a chemokine receptor that enables cells to migrate
1032 towards lymph node through lymphatic vessels, acting as antigen presenting cells. These
1033 conditions support the idea of the existence of neutrophil-like antigen-presenting cells,
1034 suggesting that they could have a strong relationship with T cells [21-24]. On the other hand,
1035 TLR-activated neutrophils cross-talk with NK cells by promoting their activation, which, in
1036 turn, drive the maturation of dendritic cells for a proper antigen presentation to T cells [25].
1037 Moreover, the previous migration of neutrophils could facilitate the infiltration of T cells
1038 towards the site of infection. In a mouse model of Influenza virus infection, the early migrated
1039 neutrophils to trachea create a trail of CXCL12 in the tissue that attract CD8⁺ T lymphocytes
1040 locally, crucial cells to deal with this disease [5]. These findings strength the multirole of
1041 neutrophils in immunity, suggesting neutrophils may differentiate into different subsets
1042 defined by distinct phenotypic and functional profiles depending on the disease, a so-called
1043 neutrophils plasticity [26]. Altogether, deepening on the basic knowledge of neutrophil
1044 biology will certainly help the better understanding of the pathogenesis of several diseases
1045 and will favor the optimization of future treatments.

1046

1047 3) Mechanisms of neutrophil recruitment

1048 As previously mentioned, the physiological control of neutrophil availability in the
1049 circulation depends on the chemokine receptor CXCR4 expressed on their surface that
1050 regulates their maintenance, egress or regress from/to bone marrow [27] (Figure 1). However,
1051 during inflammation, the recruitment of neutrophils from the bloodstream to inflamed tissue is
1052 well orchestrated by a plethora of soluble molecules and by interaction with endothelial cells
1053 [For review see 28]. Among the chemoattractants, ELR⁺ (glutamic acid–leucine–arginine)
1054 CXC chemokines constitute the most important class of mediators involved in neutrophil
1055 recruitment [29] (Figure 2A). In this regard, several attempts have been performed to block
1056 their receptors, mainly CXCR2, in different types of inflammatory diseases [30]. However,
1057 the expression of chemokine receptor on the neutrophil surface can vary along the
1058 inflammatory response. For instance, in a mouse model of severe sepsis, neutrophils lose the
1059 expression of CXCR2 in a mechanism dependent on the signaling molecules PI3K γ and
1060 GRK2, impairing their recruitment to the site of infection by the previously produced
1061 chemokines resulting in a lessened control of bacterial growth [31,32]. In the same model,
1062 neutrophils can express CCR2, a non-canonical receptor for these cells, which is functional
1063 and causes neutrophil migration due to binding to its ligands [33]. In addition to chemokines,
1064 LTB₄, C5a, and especially formyl-peptide (fMLP) are fundamental to a precise guidance of
1065 neutrophils to inflammatory foci [34,35].

1066 Along the vascular bed in proximity to inflamed tissue, chemokines are concentrated
1067 on endothelial cells by binding to glycosaminoglycans (GAGs), such as heparin, heparan
1068 sulfate, and dermatan sulfate. GAGs permit a close interaction between chemokines and
1069 leukocytes during the rolling and adhesion processes, potentiating cell migration [29] (Figure
1070 2A). Thus, the blockade of GAG-binding sites for chemokines could be useful to regulate the
1071 migration of leukocytes. In mice, the injection of modified chemokines that are still able to
1072 bind to GAGs but lost their capacity to activate their receptors impaired the recruitment of
1073 neutrophils to the tissue when stimulated by exogenous or endogenous-produced chemokines.
1074 As a result, there was reduced tissue inflammation, including articular damage in mouse
1075 models of arthritis [36,37].

1076 Currently, much attention is given to the mechanisms that regulate the so-called
1077 reverse transmigration of neutrophils, where migrated neutrophils return to blood vessels
1078 (Figure 1). This phenomenon occurs in inflamed tissue and could contribute to inflammation
1079 in distant organs [38]. However, a possible contribution to the resolution of inflammation
1080 cannot be discarded, especially in acute inflammation, as observed in zebrafish models

1081 [39,40]. Mechanistically, the decreased expression of the junctional adhesion molecule C
1082 (JAM-C) between endothelial cells facilitates the reverse transmigration of neutrophils [38].
1083 A recent work showed that LTB₄ produced during cremaster muscle under ischemia and
1084 reperfusion injury drives the neutrophil elastase release by neutrophils that induce the
1085 proteolytic cleavage of JAM-C, promoting the reverse transmigration and potentiate remote
1086 tissue inflammation [41] (Figure 1).

1087

1088 **4) Killing mechanisms of neutrophils in host defense**

1089 Impairments in neutrophils quantity or function render the host susceptible to a variety
1090 of potentially life-threatening microbial agents, which highlight the fundamental role of these
1091 cells in immune system homeostasis. Although neutrophils have a well-recognized role as
1092 phagocytic cells, their vast microbicidal mechanisms and phenotypes demonstrate they are far
1093 away from only phagocytic cells. Neutrophils killing activity occurs by two distinct
1094 mechanisms. The first one is phagocytosis, which involves intracellular degranulation of
1095 vesicles containing proteases or ROS production into the phagolysosome. The second
1096 mechanism is the release of intracellular content (neutrophils extracellular traps, NETs)
1097 (Figure 2B). The process of forming NETs induces a unique form of death known as NETosis
1098 [3]. Nevertheless, neutrophils can continue living after forming NETs for an unknown period
1099 *in vivo* depending on NETosis rout.

1100 Neutrophils recognize opsonized targets to phagocyte through opsonin receptors such
1101 as Fcγ receptors, C-type lectins or complement receptors. A complex interplay of membrane
1102 lipids, intracellular signaling cascades, and cytoskeletal rearrangements mediate this process
1103 [10]. In opposition to macrophages, neutrophils contain a collection of pre-formed granules,
1104 and the lysosome that fuses with the phagosome even before internalization is complete [42].
1105 Neutrophils present three types of granules [For review see 10,11]: (a) azurophilic or primary
1106 granules that contain cathepsin G, elastase, and proteinase 3, and myeloperoxidase (MPO).
1107 MPO highlights among these enzymes because it reaches up to 5% of total weight of
1108 neutrophils and produces the microbicidal molecule hypochlorite, a strong oxidant, upon
1109 reaction with H₂O₂ and Cl⁻ [43]. Moreover, MPO is also required for NETosis since
1110 neutrophils of MPO-deficient patients do not release NETs. Nevertheless, even low MPO
1111 activity is sufficient for NETs formation because pharmacological inhibitors or neutrophils
1112 from patients with partial MPO activity undergo NETosis [44]. (b) The second type of granule
1113 is the specific or secondary that contain lactoferrin, neutrophil gelatinase-associated lipocalin
1114 and about two-thirds of the lysozyme and, the third one is (c) gelatinase or tertiary that are

1115 granules containing gelatinase [10,11]. Alongside with phagocytosis, NADPH oxidase
1116 activation is indispensable, which is a tightly regulated process and triggered upon recognition
1117 of agents in the neutrophils cell surface [42,45]. Assembly of an active oxidase on
1118 phagosomes is necessary for generation of superoxide anion and derivative ROS, which kill
1119 engulfed microorganisms [10]. Apart from the microbicidal mechanism, superoxide anion can
1120 activate transient receptor potential (TRP) channels [46] and it is crucial to induce pain-like
1121 behaviors [47-51] (Figure 2B).

1122 One killing mechanism in particular expanded the current view on the repertoire of
1123 neutrophils killing *modus operandi* and have been a focus of investigation over the decade
1124 since its discovery [52]. The composition of NETs includes chromatin, histones, and enzymes
1125 from neutrophils granules [52]. NETs trap microbial agents limiting their action and
1126 dissemination. Furthermore, an array of antimicrobial enzymes (e.g. MPO, elastase and serine
1127 proteases) from granules facilitates the extracellular destruction of microorganisms entrapped
1128 within the NETs. Neutrophils release NETs via two central mechanisms [53,54]. The major
1129 route is the slow lytic cell death mechanism (120–240 min) – so-called suicidal NETosis – in
1130 which there is release of intracellular content and cell death, a process requiring the activation
1131 of NADPH oxidase and production of ROS [52,55,56]. The other pathway – coined as non-
1132 suicidal NETosis – involves quick (5–60 min) and NADPH oxidase-independent release of
1133 vesicles containing uncondensed chromatin and granule proteins in the extracellular space,
1134 where they assemble into NETs. The non-suicidal NETosis induces NETs formation without
1135 signs of neutrophil lysis and death [57,58]. For instance, NETosis has protective role in gout
1136 [59], and in viral [60], bacterial [52,61] and fungal infections [62]. Therefore, this is a critical
1137 defense mechanism, although not always desirable since the release of intracellular contents
1138 may drawback an innate immune response and worsening of installed diseases (we will
1139 further discuss this issue in the next topic, focusing on RA) (Figure 2B; 2C).

1140

1141 **5) Relationship between neutrophils' granules enzymes and joint tissue** 1142 **destruction in arthritis**

1143 In addition to the protective role of neutrophils during infections, these cells also have
1144 a role in tissue destruction in arthritis. In this section, we will discuss the mechanisms of
1145 neutrophils that contribute to joint lesions in arthritis.

1146 Neutrophils can be activated in RA by immune complexes (such as rheumatoid factor)
1147 both within the synovial fluid and deposited on the articular cartilage surface [63]. These
1148 complexes engage Fc γ receptors and thereby trigger neutrophil activation, which releases

1149 ROS and extracellular granule contents due to frustrated phagocytosis. Increased
1150 degranulation, ROS production and delayed apoptosis of neutrophils correlate with the
1151 intensity of synovial inflammation and the destructive capacity of joint neutrophils in RA,
1152 summarizing that granules' components and ROS act synergistically to degrade articular
1153 cartilage [63-65]. In fact, the basal levels of total ROS and superoxide anion are significantly
1154 elevated in neutrophils from RA patients compared with neutrophils from healthy individuals
1155 [66,67]. Furthermore, the stimulation of joint tissue with neutrophilic products hypochlorite
1156 and H₂O₂ inhibit the synthesis of proteoglycan [68,69]. Hypochlorite also mediates the
1157 activation of neutrophilic collagenase and gelatinase, which contributes to joint destruction
1158 [69] (Figure 2C). In agreement with these findings, treatment with antioxidant molecules such
1159 as quercetin [70,71] and curcumin [72,73] ameliorate clinical index and joint destruction in
1160 experimental models of arthritis, in part, by inhibiting oxidative stress, and neutrophils
1161 enzymes activity.

1162 Initially, NETosis was seen as an antimicrobial mechanism, but recent data suggest
1163 that NETosis could contribute to the pathophysiology of autoimmune diseases. The
1164 citrullination of proteins constitutes the major posttranslational modification that generates
1165 novel antigens recognized by antibodies (anti-citrullinated protein autoantibodies) in patients
1166 with autoimmune diseases, especially in RA [74]. As aforementioned in the section above,
1167 histones are components of NETs and are targets of hypercitrullination catalyzed by
1168 peptidylarginine deiminase 4 in an irreversible reaction [75]. NETosis seems to be the
1169 primary source of citrullinated antigens [76] and as a matter of fact, NETosis aggravates RA
1170 disease [77,78] and systemic lupus erythematosus [79,80]. In synovial fluid of patients with
1171 RA, NETosis contributes to the release of peptidylarginine deiminase 4 and consequently
1172 generation of citrullinated antigens [81] (Figure 2C).

1173 NETs formation and its importance in diseases remain intriguing. Future discoveries
1174 will aid our knowledge in NETs biology and will contribute to the understanding of its role in
1175 the development of chronic inflammatory diseases. Apparently, NADPH oxidase-dependent
1176 mechanism that activates NETosis seems more relevant in chronic diseases than NADPH
1177 oxidase-independent mechanism. Of note, neutrophils from patients with the chronic
1178 granulomatous disease who carry mutations that impair the function of Nox2 enzyme, fail to
1179 form NETs, and gene therapy restores this ability in NADPH oxidase-competent neutrophils
1180 *in vitro* [82]. Likewise, mice lacking p47^{phox} (a subunit of NADPH oxidase) fail to form NETs
1181 and resolve *A. fumigatus*-induced pneumonia, one of the leading causes of morbidity and
1182 mortality in chronic granulomatous disease [83]. Despite that, it is still necessary to unveil

1183 which NETs formation pathway is involved to which disease, and then seek for new
1184 therapeutic interventions aiming to stimulate or not NETs formation and perhaps stimulate a
1185 particular pathway of NETs formation (“suicidal” or “nonsuicidal”), accordingly with the
1186 disease.

1187 It is also important to mention that neutrophils contribute to bone resorption through
1188 osteoclast activation (Figure 2C). For instance, lipopolysaccharide [84] activates TLR4
1189 inducing receptor activator of NF- κ B ligand (RANKL) expression by peripheral blood human
1190 neutrophils and neutrophils recruited to murine air pouch. The co-culture of these neutrophils
1191 with human monocyte-derived osteoclasts and murine RAW 264.7 cells stimulates bone
1192 resorption. Importantly, confocal microscopy reveals neutrophil-osteoclast contact and even
1193 invagination. Furthermore, synovial fluid neutrophils of patients with exacerbation of RA
1194 activates osteoclastogenesis in co-culture system, which corroborates their high expression of
1195 RANKL in the synovial fluid [85]. Further demonstrating the interactions between neutrophils
1196 and osteoclasts in RA, the neutrophil chemoattractant chemokine CXCL2 induces RANKL
1197 production by osteoclast precursors in a JNK- and NF- κ B-dependent manner as well as
1198 osteoclastogenesis. The CXCR1/2 receptor antagonist repertaxin reduced RANKL-induced
1199 osteoclastogenesis, and the administration of CXCL2 in mice induces bone erosion. In
1200 agreement with these *in vitro* and murine evidence, there are high titers of CXCL2 in the
1201 blood and synovial fluid of RA patients [86].

1202

1203 **6) Balancing the role of neutrophils in infectious and non-infectious arthritis**

1204 Inhibition of neutrophil recruitment is a double edge sword in arthritic diseases. In one
1205 hand, inhibiting neutrophil recruitment in infectious arthritis is detrimental. On the other hand,
1206 inhibiting neutrophil recruitment in non-infectious arthritis reduces the disease severity. For
1207 instance, in septic arthritis, neutrophils control bacterial load limiting infection. In RA,
1208 neutrophils are the main infiltrating cells in the synovial fluid, and their turnover can exceed
1209 10^9 cells per day in a 30 mL joint effusion [87]. In both non-infectious and infections arthritis
1210 neutrophils contribute to tissue lesion. In addition to this dichotomy, neutrophils also have a
1211 dual role in gout arthritis as discussed below.

1212 Neutrophils in the circulation usually have a short lifespan and rapidly undergo
1213 constitutive apoptosis. However, during migration toward inflammatory sites, activated
1214 neutrophils encounter changes in the inflammatory microenvironment that stimulate
1215 expression of genes related to longevity. One of these changes lies in the increased activity of
1216 the transcription factor Foxo3a that ensures neutrophil survival during inflammatory arthritis

1217 [64]. FOXO transcription factors have been found to play a critical role in the regulation of
1218 proliferation, apoptosis and oxidative stress [88]. A milder disease courses in patients carrying
1219 the *FOXO3A* minor allele (G, but not T haplotype) of rs12212067 [89] and increased
1220 expression of FoxO3 in polymorphonuclear cells in RA [90]. Accordingly, Foxo3a-deficient
1221 mice resist to K/BxN serum transfer-induced arthritis due enhanced FAS ligand expression
1222 and the consequent early neutrophil apoptosis [64]. This evidence suggests that delayed
1223 apoptosis of neutrophils is an important component in the development of RA, and therefore,
1224 inhibition of these pathway/or facilitating neutrophil apoptosis and inhibiting neutrophil
1225 recruitment are possible approaches to reduce inflammatory arthritis. Monosodium urate
1226 (MSU) crystal deposition in joints causes gouty arthritis with the participation of components
1227 of the innate immune system and mononuclear phagocytes, which are critical orchestrators of
1228 the inflammatory response. Nevertheless, neutrophils have a prominent role in the tissue
1229 lesion especially in gout attack in which the neutrophil counts increase drastically [91].
1230 Seminal studies in crystal-driven gout in canines demonstrated the fundamental role of
1231 neutrophils in the acute phase of gout. In these studies, neutrophil depletion significantly
1232 suppressed inflammation suggesting that its inhibition could be a new target for gouty arthritis
1233 [84,92]. The recruited neutrophils also produce IL-1 β , TNF- α , IL-8, and IL-6 contributing to
1234 increasing inflammation [91,93]. In this regard, the blockade of the chemokine receptor
1235 CXCR2 reduces the infiltration of neutrophil, joint inflammation and pain induced by MSU
1236 crystals in mice [94]. In fact, neutrophils contribute to inflammatory pain through the
1237 production of hyperalgesic molecules including leukotrienes and prostaglandins in response to
1238 a broad range of endogenous stimuli such as endothelin-1, LTB₄, IL-1 α and complement
1239 system molecules such as C5a [95-98] (Figure 2D). On the other hand, the inhibition of
1240 neutrophil recruitment may be prejudicial in gouty arthritis. NETs form dense aggregates that
1241 proteolytically degrade MSU crystals. Furthermore, the subsequent phagocytosis of MSU
1242 degraded components by macrophages ultimately lead to a reduction of inflammatory process
1243 [59]. This mechanism accounts for the resolution of the acute phase of gout. Therefore, in
1244 gouty arthritis neutrophils have a dual role of contributing to tissue lesion, but also to limiting
1245 inflammation triggers. It is likely that focusing only in inhibiting neutrophil chemotaxis in
1246 gouty arthritis is not the definitive therapeutic approach, but rather reducing the harmful
1247 actions maintaining the beneficial effects of neutrophils is more appropriate. For instance,
1248 determining the mechanisms regulating the fate of neutrophils as to whether they will form
1249 NETs or not seems an important approach to enhance the degradation of MSU crystals.

1250 Septic arthritis is one of the most dangerous joint diseases due to the rapid disease
1251 development course, severe joint lesion, sequels to the joint function and pain and even septic
1252 shock-induced death despite the use of antibiotics. The primary etiologic agent of septic
1253 arthritis is *Staphylococcus aureus*, an extracellular growing bacterium [99]. There is evidence
1254 that both drug resistant *S. aureus* such as methicillin-resistant *Staphylococcus aureus* (MRSA)
1255 and non-drug resistant *S. aureus* cause septic arthritis. The MRSA is a cause of septic arthritis
1256 in hospitalized patients, and non-drug resistant *S. aureus* causes septic arthritis in cases of
1257 spontaneous development within the community [100,101]. A contributing factor for *S.*
1258 *aureus*-induced arthritis is the immune suppression induced by drug, diseases and aging
1259 [100].

1260 Neutrophils depletion may increase pro-inflammatory cytokine production (e.g. TNF-
1261 α and IL-6) in septic arthritis, which would seem a contradiction if we consider the pro-
1262 inflammatory role of neutrophils. However, the increasing cytokine production by neutrophil
1263 depletion is related to an increase in disease severity related to increasing the bacterial load.
1264 Failure in controlling bacteria proliferation results in increasing activation of tissue resident
1265 cells that produce higher amounts of cytokines and chemokines. Therefore, neutrophils play a
1266 protective role due to their phagocytic ability and are critical cells for the outcome during the
1267 early stages of *S. aureus* infection although they also contribute to tissue lesion [102] (Figure
1268 2B, 2C). In agreement with that, mice develop more frequent and severe clinical septic
1269 arthritis upon anti-inflammatory therapy. Neither treatment with IL-1 receptor antagonist (IL-
1270 1Ra) [103] nor inhibition of the transcription factors NF- κ B and AP-1 alone or in combination
1271 with antibiotics ameliorate *S. aureus*-induced arthritis [104]. These data may be due to the
1272 counteracting effect of increased bacterial load due to diminished bactericidal response. A
1273 bias of these studies is that *S. aureus* was injected systemically causing septic arthritis
1274 alongside with sepsis, which is a condition with a fundamental role of neutrophil recruitment
1275 toward the inflammatory foci to host survival [32,105]. Therefore, remains the question
1276 whether the inhibition of neutrophil would benefit the host with *S. aureus* infection in the
1277 joint cavity representing the condition of established septic arthritis only and not the events
1278 that would lead to septic arthritis.

1279 The ability of the host to respond to bacterial infection is essential since neutrophils
1280 are an important part of the host protective system. In fact, patients with rheumatoid arthritis
1281 are 4- to 15-fold more susceptible to septic arthritis than the general population [100]. A
1282 likely explanation is that the presence of joint lesions facilitates its colonization together with
1283 a reduced immune response due chronic treatment with disease modifying drugs,

1284 corticosteroids, non-steroidal anti-inflammatory drugs and biologic therapies that cause
1285 patient immune suppression.

1286 An important point to keep in mind is that the role of neutrophils in disease context
1287 may vary as a consequence of the inflammatory milieu. For instance, we have shown that in a
1288 model of antigen-induced arthritis in mice, TNF- α induces the expression of ST2 (IL-33R)
1289 rendering neutrophils chemoattractable by IL-33. Corroborating, TNF- α induces ST2
1290 expression and IL-33 responsiveness of human neutrophils, and neutrophils of RA patients
1291 under infliximab treatment present less ST2 expression and IL-33 responsiveness [106].
1292 Therefore, determining the neutrophil phenotype under varied infectious and non-infectious
1293 inflammatory disease may open a venue to selectively targeting neutrophil phenotype related
1294 to a particular group of diseases without increasing the susceptibility to another group of
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1339 inflammatory disease may open a venue to selectively targeting neutrophil phenotype related
1340 to a particular group of diseases without increasing the susceptibility to another group of
1341 diseases.

1342

1343 **7) Pharmacological approaches to modulate the neutrophil recruitment and** 1344 **activity in animal models of arthritis**

1345 Varied animal models of inflammatory arthritis show that neutrophils are the first
1346 immune cells that migrate toward arthritic joint, and that early measures of joint inflammation
1347 correlate with neutrophil infiltration [64,106-109]. The pharmacological approaches that aim
1348 neutrophil inhibition (Table 1) highlight its importance in RA. Taking into account that the
1349 inhibition of neutrophil recruitment is prejudicial to the host in septic arthritis and sometimes
1350 in gout arthritis, in this section, we will highlight pharmacological approaches targeting
1351 neutrophil recruitment and activity presenting therapeutic benefit in RA. We also provided

1352 information on the pharmacological/genetics approaches related to infectious arthritis models
1353 and their consequence to the host (Table 2).

1354 Chemokines, especially CXCL1 and CXCL2 [110], provide a luminal gradient that
1355 guides neutrophils towards inflammatory foci. This role of chemokines supports chemokine
1356 receptors as useful targets to modulate neutrophil recruitment. In a model of antigen challenge
1357 in immunized mice, treatment with DF2162, an allosteric antagonist of CXCR1 and CXCR2,
1358 decreases neutrophils recruitment and MPO activity, and consequently hyperalgesia
1359 (exacerbated pain response) and TNF- α production [111,112]. Strengthening the therapeutic
1360 potential of antagonizing chemokines receptors, in two models of arthritis, two selective
1361 CXCR1/2 antagonists (SCH563705 and repertaxin) reduce disease activity, clinical score,
1362 paw swelling, pro-inflammatory cytokine production and neutrophil recruitment [113,114].

1363 Alongside with chemokines, C5a fragment (a peptide of complement system) bounds
1364 to C5aR expressed by neutrophils and acts as chemoattractant and activator of these cells
1365 depending on its concentration. At low concentration C5a acts as a chemoattractant while, at
1366 higher concentration, C5a can elicit neutrophil production of superoxide anion and enzyme
1367 release [115]. Inhibition of C5a-C5aR signaling through antibody anti-C5aR [116,117] or in
1368 C5aR knockout mice [118,119] reduces the clinical and histological score, neutrophil
1369 recruitment, neutrophils enzymes activity (MPO and MMP-9), pro-inflammatory cytokine
1370 production and bone erosion in models of experimental arthritis. Additionally, the importance
1371 of this signaling lies on C5aR and Fc γ R activation in neutrophils that induce the release of
1372 LTB₄ and IL-1 β . These mediators contribute to promotion and progression of arthritis [118].

1373 Considering the vast enzymatic arsenal of neutrophils, inhibition of their degranulation
1374 would be an interesting approach in RA. Pentoxifylline, a nonspecific phosphodiesterase
1375 inhibitor and a reducer of TNF- α mRNA half-life, attenuates neutrophil oxidative burst and
1376 decrease pro-inflammatory mediator synthesis [120]. Treatment with pentoxifylline
1377 diminishes neutrophil recruitment and TNF- α production in ovalbumin immunized and
1378 challenged mice [121] and in uric acid crystal-induced joint inflammation in mice [122]. In
1379 agreement with that, treatment with pentoxifylline reduces arthritic index, neutrophil
1380 recruitment, TNF- α production, and prevents proteoglycan loss in a model of antigen-induced
1381 arthritis [123].

1382 The murine K/BxN serum transfer model closely resembles human arthritis [124]. In
1383 this model, mice lacking 5-LOX and 15-LOX (key enzymes in leukotrienes and lipoxins
1384 production) or a 5-LOX pharmacological inhibitor (L-739,010) ameliorate arthritis by
1385 decreasing clinical index, bone and cartilage erosion alongside with neutrophils recruitment

1386 [125,126]. Likewise, neutrophil-derived LTB₄ signaling through either BLT1 or BLT2
1387 accounts for the development of K/BxN serum transfer model of arthritis [127,128]. Signaling
1388 of neutrophils through BLT1 drives the production of IL-1 β , which amplifies arthritis severity
1389 by inducing the production of neutrophil chemoattractants by synovial cells [129]. In
1390 agreement with that, antagonizing the LTB₄ receptor BLT1 [130] or using BLT12/2 or
1391 BLT1/BLT2 double-knockout mice result in protection from the development of collagen-
1392 induced arthritis [131]. In an inverse pathway, this leukotriene system can also be triggered by
1393 cytokines and modulate neutrophil recruitment. For instance, IL-15 induces IL-18 maturation
1394 in synovial fluid neutrophils and peripheral blood neutrophils of RA patients, but not in
1395 peripheral blood neutrophils of healthy human volunteers. IL-15 induces IL-18-dependent
1396 production of LTB₄ by RA neutrophils and induces neutrophil migration in ovalbumin
1397 immunized and challenged mice by the same mechanism [132]. Targeting IL-15 with IL-15
1398 soluble receptor also reduces pain in ovalbumin immunized and challenged mice [133]. In
1399 fact, HuMax-IL-15 (a human anti-IL-15 monoclonal antibody) reaches clinical trial and
1400 ameliorates disease activity, inhibits IL-15-induced release of IFN- γ and reduces CD69
1401 expression by synovial mononuclear cells in patients with RA [134].

1402 Regarding infectious arthritis such as Lyme's disease (a *Borrelia burgdorferi*
1403 infection), 5-LOX deficiency is prejudicial to the host, contributing to earlier joint swelling,
1404 inability to resolve arthritis and to worsening of arthritis score [135]. *B. burgdorferi* also
1405 induces IL-15 (which induces LTB₄ production [132]) production by neutrophils and
1406 mononuclear cells [136]. Importantly, targeting IL-15 with antibody and soluble receptor
1407 prevents *B. burgdorferi* vaccination and infection-induced paw edema, cellular infiltration in
1408 the joints (neutrophils, macrophages, and lymphocytes) and histopathologic indications of
1409 arthritis, including hyperplasia, hypertrophy, and villus formation of the synovium [137].
1410 These set of data demonstrate the fundamental role of neutrophils in animal models of
1411 arthritis and may provide insight into the human inflammatory disease. Figure 2 summarizes
1412 the molecules we discussed and additional molecules with a role in non-infectious arthritis
1413 models. It is also important to point out that are important molecules that modulate diseases
1414 involving joint inflammation, but that do not seem to have as central mechanism targeting
1415 neutrophils. For instance, there are therapies targeting activation of B cells, T cells, NK cells,
1416 innate lymphoid cells (known as group 3 ILCs) and osteoclasts, as reviewed by others [138-
1417 141].

1418

1419

1420 **8) Perspectives of therapies targeting neutrophils in arthritis**

1421 Neutrophils represent a promising pharmacological target in RA considering the role
1422 of innate immunity in addition to adaptive immunity in RA pathophysiology [142], the high
1423 counts of neutrophils in inflamed synovial fluid and the contribution of neutrophils to tissue
1424 lesion in RA. Of all cells implicated in the pathophysiology of arthritic diseases, neutrophils
1425 possess the greatest cytotoxic potential, owing to their ability to release degrading enzymes,
1426 the release of NETs-containing enzymes, pro-inflammatory cytokines, reactive oxygen
1427 species and regulation of osteoclastogenesis. Thus, to improve comprehension, we highlighted
1428 some examples in Figure 2 of potential targets of neutrophil function and recruitment, and in
1429 Table 2 examples of compounds undergoing clinical trials that target the neutrophil.

1430 The current treatment of RA patients includes glucocorticoids, non-steroidal anti-
1431 inflammatory drugs and disease modifying anti-rheumatic drugs. Biological agents such as
1432 monoclonal antibodies and recombinant proteins that target TNF- α , CD20, CTLA-4
1433 (cytotoxic T-lymphocyte-associated protein 4), IL-1 receptor, as well as therapies based on
1434 the blockade of T-cell and B-cell functions have shown efficacy to control physical signs and
1435 pain [143,144]. For instance, methotrexate is a compound that blocks folic acid metabolism
1436 and is widely used in clinical settings. Its benefits in RA include stimulation of neutrophil
1437 apoptosis [145], inhibition of NF- κ B pathway [146], reduction of adhesion molecules
1438 expression, LTB₄ production [147], and consequently neutrophil recruitment, and ROS
1439 production [148]. Anti-TNF- α therapies are also widely used for the treatment of RA patients.
1440 Its benefits include reduction in adhesion molecules [149,150], and neutrophil recruitment and
1441 activation, accompanied by a reduction of NF- κ B activity and pro-inflammatory cytokines
1442 such as IL-6, IL-1 β and IL-8 [149,151]. The current clinical practice has these molecules as
1443 targets, but their side effects and high costs usually limit their applicability in the chronic
1444 treatment of RA [143]. The development of adaptive immunity against the biologic therapies
1445 regularly occurs limiting their use. Some RA patients are refractory to even high doses of
1446 glucocorticoids or immunosuppressive agents [143,152]. Paradoxically, corticosteroids inhibit
1447 many neutrophil functions, but also delay its apoptosis, an effect that needs further
1448 investigation for a full understanding of its underlying mechanism and relevance *in vivo*
1449 [153]. Soluble ST2 and IL-33 are present in higher concentrations in plasma/serum of
1450 patients with RA compared to healthy volunteers. Furthermore, the IL-33/ST2 signaling
1451 pathway has a pro-inflammatory role in RA [106,154,155]. Assessing neutrophils in
1452 peripheral blood of patients with RA, we observed that neutrophils derived from RA patients

1453 treated only with methotrexate present higher expression of ST2 compared to healthy donors
1454 or patients under treatment with methotrexate plus infliximab. Thus, after TNF- α stimulus,
1455 neutrophils from healthy donors increase the expression of ST2. This finding supported the
1456 clinical relevance of IL-33 in RA and provided an important novel mechanism by which anti-
1457 TNF- α therapy ameliorates inflammation [106]. Likewise, RA patients treated with
1458 Etanercept have significant reduction of serum IL-33 levels, which showed significant
1459 correlations with the number of tender joints, VAS scale and C-reactive protein [156].
1460 Therefore, targeting IL-33 may provide very promising therapy to RA and certainly deserves
1461 further investigation in clinical trials.

1462 The use of molecules from natural sources represents a promising land to treat RA (but
1463 not only), by acting as primary therapy or at least as therapeutic adjuvants to reduce the daily
1464 doses of conventional drugs that RA patients receive [152,157,158]. Importantly, there are
1465 molecules from natural sources with fewer side effects than current therapies alongside with
1466 low toxicity, allowing utilization of high concentrations. Curcumin and quercetin highlight
1467 between these molecules. Curcumin is an extremely pleiotropic compound and presents more
1468 than 100 different targets [159]. Both quercetin [160] and curcumin [161] block neutrophils
1469 recruitment through inhibition of cellular signaling responsible for actin polymerization in
1470 association with down-regulation of adhesion molecules [162,163]. Curcumin also possesses
1471 bactericidal activity, which could be useful in cases of infectious arthritis [164,165]. The
1472 potential of curcumin in patients with RA was first reported back in 1980 with reduction of
1473 joint swelling, morning stiffness, and improvement of walking time [166]. A clinical trial with
1474 patients with RA also shows that curcumin reduces reported pain, tenderness and swelling of
1475 joint [167]. Corroborating, Meriva® (a curcumin-based medicine) demonstrates efficacy in
1476 clinical trials with patients with osteoarthritis by reducing reported pain (VAS scale) [168]. In
1477 another clinical trial, treatment with Meriva® reduces stiffness and physical signs (treadmill
1478 test), alongside with IL-1 β , IL-6, and sVCAM-1 production [169].

1479 Another natural product-derived molecule that reaches clinical trials is Maritech®,
1480 which is a fucoidan-based medicine – a molecule that binds to the adhesion molecule L-
1481 selectin inhibiting its function. Treatment with Maritech® reduces reported joint pain,
1482 stiffness and difficulty with physical activities in patients with osteoarthritis in phase I and II
1483 open label study [170]. In an accompanying study in healthy volunteers, treatment with
1484 Maritech® enhances serum antioxidant capacity and reduces IL-6 levels [171]. Associated
1485 with that, treatment with Maritech® did not change cholesterol levels, liver, renal, and

1486 hematopoietic functions during three months treatment; demonstrating that it is a safe drug
1487 [170].

1488 Growing evidence point to agonist of receptors that leads to resolution of
1489 inflammatory process as a promising therapy for inflammatory diseases. Resolution of
1490 inflammation is a highly ordered active process, tightly regulated by many lipid mediators,
1491 enzymatically derived from essential polyunsaturated fatty acids (PUFA), and are termed
1492 specialized pro-resolving lipid mediators (SPMs). Lipoxin A4 (LXA4, an agonist of
1493 ALX/FPR2) and resolvins (Rv, an agonist of ALX/FPR2) are the most studied SPMs [172].
1494 For instance, treatment with RvE1 reduces IL-8-induced neutrophil chemotaxis [173] and
1495 similarly, treatment with LXA₄ reduces neutrophil adherence and transmigration [174].
1496 Isolated SPMs efficacy lies, in part, by inhibition of neutrophils recruitment to inflammatory
1497 foci. In fact, SPMs and PUFA have been tested in models of arthritis [109,175-177] and
1498 clinical trials [178], respectively. Using K/BxN serum transfer model, ALX/FPR2 KO mice
1499 exhibit exacerbated disease severity suggesting an endogenous role of SPMs in limiting
1500 disease development [175]. Administration of LXA₄ inhibits edema and neutrophil influx
1501 accompanied with a reduction of preproET-1 mRNA, KC/CXCL1, LTB₄ and TNF- α levels
1502 [109]. Resolvin D1 (RvD1) is a potent regulator of pain and neutrophils recruitment and
1503 binds to ALX/FPR2 receptor [179]. Treatment with RvD1 reduces pain and tissue damage,
1504 proving to be more potent than either steroid or non-steroidal anti-inflammatory drugs
1505 (NSAIDs). Its mechanism lies in the reduction of NF- κ B and COX-2 expression in the spinal
1506 cord, and, in arthritic joints, inhibition of TNF- α and IL-1 β production [177]. Regarding
1507 polyunsaturated fatty acids (PUFAs), patients with RA, who use omega-3 fatty acids reduce
1508 the use of NSAIDs and cardiovascular risk factors, and increase the period of disease
1509 remission compared to patients who did not use omega-3 fatty acids [180]. Randomized
1510 controlled clinical trials using omega-3 fatty acids in RA reported improvement in several
1511 clinical outcomes including reduced duration of morning stiffness, reduced number of tender
1512 or swollen joints, reduced reported joint pain, increased grip strength and decreased use of
1513 NSAIDs [178]. In mononuclear cells from healthy donors and stimulated with LPS,
1514 supplementation with omega-3 fatty acids results in reduced levels of TNF- α and IL-1 β
1515 [181,182]. A meta-analysis of omega-3 fatty acids treatment and pain demonstrates that
1516 supplementation with omega-3 fatty acids for 3-4 months reduces patient-
1517 reported joint pain intensity, minutes of morning stiffness, the number of painful/tender joints,
1518 and NSAIDs consumption [183].

1519 SPMs present anti-inflammatory activity without immunosuppressive activity, which
1520 may be a beneficial feature in infectious arthritis to control painful symptoms and bacterial
1521 load. There is evidence in different experimental models demonstrating that treatment with Rv
1522 enhances the clearance of *E. coli* and *S. aureus* [184,185]. Aligned with that, treatment with
1523 Rv increases the clearance of these bacteria by stimulating non-phlogistic phagocytosis,
1524 enhancing bactericidal activity of antibiotic, which ultimately results in lowering antibiotic
1525 requirement and contributes to diminishing resistance [185]. Regarding isolated SPM, RvE1
1526 reaches clinical trials as RX-10045[®] for dry eye syndrome, and produces dose-dependent,
1527 statistically significant improvements on the primary endpoints of signs and symptoms of dry
1528 eye (Clinicaltrials.gov identifier: NCT00799552) [186,187]. Thus, these set of data
1529 demonstrates robust clinical trials are evidencing the efficacy of omega-3 fatty acids PUFAs
1530 in RA. Associated with animal data, and that SPMs present therapeutic effectiveness in
1531 picogram ranges [172], supporting and encouraging the use of isolated SPMs in clinical trials
1532 for patients with infectious or non-infectious arthritis.

1533 The interaction between platelets and neutrophils is an essential checkpoint to
1534 neutrophil recruitment [188]. An indispensable component is the selectin ligand PSGL-1,
1535 which signaling results in the redistribution of receptors (such as Mac-1 and CXCR2) that
1536 drives neutrophil migration toward tissues [189]. This interaction seems to be an important
1537 step to the formation of NETs as well [189,190]. Accordingly, in a model of collagen-induced
1538 arthritis, treatment with a soluble receptor of PSGL-1 reduces the clinical severity and TNF- α
1539 production [191], which might be explained by inhibition of NETs formation [189,190]
1540 Interestingly, mice that overexpress P-selectin increase the amount of NETs formation
1541 following the stimulation with platelet-activating factor (PAF) [190]. Another point that
1542 strengthens the platelet-neutrophil cross-talk lies in the activation of neutrophils through
1543 receptor advanced glycation end products (RAGE) activation by high mobility group box 1
1544 (HMGB-1) released from platelets [192]. In neutrophils, this induces autophagy and
1545 preferentially leads to death by NETosis instead of apoptosis [193,194]. In fact, mice that
1546 received platelets lacking HMGB-1 gene [194] or mice lacking RAGE [195] fail to release
1547 NETs. Although in conditions such as cancer, autophagy results in enhanced survival of
1548 neutrophils and NETs formation [195]. Future therapeutic intervention exploiting these
1549 mechanisms might involve shifting the balance toward the resolution of inflammation by
1550 promoting aggregation of NETs.

1551 Over the past years, the multistep process of neutrophils recruitment from blood to
1552 tissue has been elegantly studied [34,35,189,196]. A reasonable future approach should

1553 consist in selectively inhibiting neutrophils recruitment accordingly with the characteristic of
1554 initial noxious stimuli. Neutrophils tend to respond better to ending chemoattractant such as
1555 formyl-peptide (fMLP) and C5a than IL-8 and LTB₄ [35]. This spatial, temporal and
1556 hierarchic cascade of mediators involving neutrophil recruitment toward tissues prompts
1557 therapies aiming inhibition of a starter chemoattractant (initial targets, IL-8 and LTB₄) or
1558 ending chemoattractant (final targets, fMLP and C5a) by using selective antagonist targeting a
1559 specific step. Regarding adhesion molecules, neutrophils adhesion and recruitment within
1560 liver sinusoids in sterile inflammation depend on the integrin Mac-1 and ICAM-1 [34]. In
1561 contrast, when used *E. coli* as stimuli for neutrophil recruitment to the liver surface rather than
1562 a necrotic injury, the adhesion of neutrophils in sinusoids depends on CD44 rather than Mac-
1563 1, revealing different adhesion molecules for neutrophil recruitment in infection versus sterile
1564 inflammation [34]. Furthermore, the observation that neutrophils rapidly migrated away from
1565 high concentrations of CXCR2 ligands implies that the necrotactic stimulus must
1566 hierarchically override CXCR2 signaling. Mice lacking fMLP receptor, but not those lacking
1567 CXCR2, significantly attenuated neutrophils chemotaxis next to necrotic foci, demonstrating
1568 that fMLP receptor guides precise localization of neutrophils into areas of sterile tissue
1569 necrosis, and further corroborates this hierarchic cascade [34]. The last barrier to neutrophils
1570 transmigration to inflammatory foci requires a cross-talk between neutrophils and pericytes
1571 [For review see 197]. Pericytes contribute actively to the precise guidance of neutrophils by
1572 paving the road through modulation of basement membrane proteins and signaling hotspots
1573 gaps that drive neutrophils into inflammatory foci [197]. The communication between ICAM-
1574 1 of pericytes and Mac-1 and LFA-1 of neutrophils, alongside to the enlargement of the gaps
1575 size by pericytes mediate this guidance [197-199]. Associated to that, pericytes recognize
1576 DAMPs and PAMPs [197]; produce chemokines, such as CXCL1, IL-8, and MIF; express
1577 ICAM-1 following TNF- α and IL-1 β stimulation [199]; and enhance neutrophils survival
1578 [198] which further increase this cascade.

1579 Taking into account the differential requirement of adhesion molecules in infectious
1580 and non-infectious recruitment, direct inhibition of selective adhesion molecules is a logic
1581 therapeutic approach. In agreement with that, treatment with anti-TNF- α and corticosteroids
1582 of RA patients down-regulate the expression of adhesion molecules such as VCAM-1, ICAM-
1583 1, and E-selectin [150,151,200,201]. Despite that, inhibition of integrin or selectin in varied
1584 clinical trials fails to improve symptoms in patients with RA [202-204] or psoriasis [205].
1585 However, using alefacept (LFA-3-IgG1 fusion protein, a T cell-specific agent) on synovial
1586 inflammation in patients with psoriatic arthritis improves both clinical and histopathological

1587 [206]. This issue remains poorly understood and even in animal models data are
1588 contradictory, since inhibition of adhesion molecules demonstrates efficacy [207,208], or no
1589 efficacy depending on which adhesion molecule was the target. For instance, anti-LFA-1
1590 presents no effect [207] or aggravates the disease, in the case of mice lacking selectin [209].
1591 In a mouse model of collagen-induced arthritis, treatment with PF-03475952 (an anti-CD44
1592 monoclonal antibody) reduces the incidence of arthritis and clinical index [210]. In a clinical
1593 trial with patients with head and neck cancer, the efficacy of an immuno-conjugate
1594 (bivatuzumab mertansine) consisting of an anti-microtubule agent coupled to an anti-CD44v6
1595 monoclonal antibody was tested. In this study, three patients showed a partial response;
1596 however, the treatment caused severe skin toxicity with a fatal outcome, leading to early
1597 termination of this trial [211]. Therefore, a better screening on the approach of adhesion
1598 molecule as the target is still necessary.

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1600 **9) Concluding remarks**

1601 Enlightenment of the neutrophil recruitment cascade and understanding that the
1602 characteristic of initial noxious stimuli requires different adhesion molecules and
1603 chemoattractants might improve the knowledge by which neutrophils migrate toward the
1604 tissue. If a specific chemoattractant or receptor is not available at the appropriate time and
1605 place, the cascade comes to a halt, and the neutrophil inflammatory reaction collapses. Thus,
1606 targeting neutrophils recruitment through this multistep cascade, and controlling NETs
1607 formation (stimulating or not, or perhaps specific pathway “suicidal” or “nonsuicidal”), might
1608 represent prospering therapeutic approaches for non-infectious arthritis. Regarding infectious
1609 arthritis, therapies that enhance the microbicidal activity of neutrophils or macrophages
1610 alongside with anti-inflammatory benefits (as isolated SPMs and curcumin), seem a good
1611 option for the nearest future in an attempt to reduce antibiotic resistance. Therefore,
1612 controlling the inflammatory response is essential to avoid excessive tissue damage and
1613 consequent loss of function in infectious and non-infectious arthritis. Thus, the neutrophil
1614 seems a promising cellular target to achieve a balanced inflammatory response.

1615

1616 **Conflict of interest**

1617 The authors declare no conflict of interest.

1618

1619 **Acknowledgements**

1620 This work was supported by grants from MCTI/SETI/Fundação Araucária (Ministério
 1621 da Ciência, Tecnologia e Inovação/Secretaria da Ciência, Tecnologia, e Ensino Superior do
 1622 Paraná/ Fundação Araucária), Coordenadoria de Aperfeiçoamento de Pessoal de Nível
 1623 Superior (CAPES), Conselho Nacional de Desenvolvimento Científico e Tecnológico
 1624 (CNPq), Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG), and
 1625 Decit/SCTIE/MS (Departamento de Ciência e Tecnologia da Secretaria de Ciência,
 1626 Tecnologia e Insumos Estratégicos, Ministério da Saúde) intermediated by CNPq and support
 1627 of Fundação Araucária and Secretaria de Saúde do Estado do Paraná (SESA-PR), Brazil.

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1629 Table 1. Examples of potential therapies targeting neutrophil recruitment and activation

1630

Type	Compound	Stage/Model	Reported effect	Reference
Natural product-derived	Fucoidan	mBSA-immunised (mice)	Inhibition of neutrophil recruitment and mechanical hyperalgesia	[212]
		Collagen-induced (mice)	Reduction of adhesion molecules, TNF- α and INF- γ production and leukocyte infiltration	[213]
		Human Fibroblast-like synoviocytes (<i>in vitro</i>)	Attenuation of expression and secretion of metalloproteinase, accompanied by suppression of NF- κ B and reduction in the viability, survival, and invasiveness of IL-1 β -treated of RA Fibroblast-like synoviocytes	[214]
Soluble receptor	ST2	mBSA-immunised (mice)	Inhibition of neutrophil recruitment, reduction of pain, and IL-1 β and TNF- α production	[106,215]
		Collagen-induced (mice)	Attenuation of diseases progression, reduction of TNF- α , IL-6 and IL-12 production, and inhibition of neutrophil recruitment	[216]
	PSGL-1	Ovalbumin-sensitized (mice)	Reduction of clinical severity and TNF- α production	[191]
Antibody	ST2	Collagen-	Attenuation of clinical	[217]

		induced (mice)	severity, joint destruction, reduction in neutrophils recruitment and RANKL mRNA expression	
	C5aR	Collagen-induced (mice)	Reduction of clinical and histological score, inhibition of neutrophil recruitment, TNF- α , IL-6, KC and MCP-1 production	[116]
Agonist of receptor	LXA ₄ and BML-111 (lipoxin agonists)	Zymosan (mice)	Inhibition edema and neutrophil influx, preproET-1 mRNA, KC/CXCL1, LTB4 and TNF- α levels	[109]
	RvD1	CFA-induced (rats)	Reduction of hyperalgesia, TNF- α and IL-1 β production, and joint and paw swelling	[177]
	RvE1 (RX-10045)	Clinical Trials (Dry eye syndrome)	Improvement in dryness, stinging, burning, ocular discomfort, and leukocyte recruitment (CAE staining)	[187]
Antagonism of receptor	LY293111N a (LTB4 antagonist)	Collagen-induced (mice)	Reduction of arthritic index and neutrophil recruitment	[130]
	SCH563705 (CXCR1/2 antagonist)	Anti-collagen antibody-induced (mice)	Reduction of disease activity, paw swelling, pro-inflammatory cytokine production, neutrophil recruitment	[114]
	DF2162 (CXCR1/2 allosteric antagonist)	Zymosan (mice)	Inhibition of neutrophil recruitment and mechanical hyperalgesia	[95]
		mBSA-immunised (mice)	Inhibition of neutrophil recruitment, TNF- α , CXCL1, and CXCL2 production	[112]

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1635 Table 2. Examples of emerging therapies in clinical trials targeting neutrophil recruitment and
 1636 activation
 1637

Type	Compound	Disease	Reported effect	Reference
Natural product-derived	Meriva® (curcumin-based medicine)	Osteoarthritis	Reduction of reported pain (WOMAC scale), stiffness and physical signs (treadmill test); inhibition of IL-1 β , IL-6, and sVCAM-1 production	[169]
		Osteoarthritis	Reduction of reported pain (VAS scale)	[168]
	Curcumin	RA	Reduction of reported pain, tenderness, swelling of joint and C-reactive protein	[167]
		RA	Reduction of joint swelling, morning stiffness, and improvement of walking time	[166].
	Maritech® (fucoidan-based medicine)	Osteoarthritis	Reduction of reported joint pain, stiffness, difficulty with physical activities	[170]
Supplementation	Omega-3 fatty acids	RA	Reduction of reported pain, use of NSAID and disease modifying anti-rheumatic drugs	[218]
			Reduction of reported pain, duration of morning stiffness, rheumatoid factor, joint pain and time to onset of fatigue	[219]
			Reduction of reported pain and use of NSAID	[220].

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Reference

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1643 1 Gourley M, Miller FW. Mechanisms of disease: Environmental factors in the pathogenesis of
1644 rheumatic disease. *Nat Clin Pract Rheumatol* 2007;3:172-180.
- 1645 2 Asquith DL, Miller AM, McInnes IB, Liew FY. Animal models of rheumatoid arthritis. *Eur J*
1646 *Immunol* 2009;39:2040-2044.
- 1647 3 Kolaczowska E, Kubes P. Neutrophil recruitment and function in health and inflammation. *Nat*
1648 *Rev Immunol* 2013;13:159-175.
- 1649 4 Mocsai A. Diverse novel functions of neutrophils in immunity, inflammation, and beyond. *J Exp*
1650 *Med* 2013;210:1283-1299.
- 1651 5 Lim K, Hyun YM, Lambert-Emo K, Capece T, Bae S, Miller R, Topham DJ, Kim M. Neutrophil
1652 trails guide influenza-specific cd8(+) t cells in the airways. *Science* 2015;349:aaa4352.
- 1653 6 Wright HL, Moots RJ, Edwards SW. The multifactorial role of neutrophils in rheumatoid arthritis.
1654 *Nat Rev Rheumatol* 2014;10:593-601.
- 1655 7 Styrt B. Species variation in neutrophil biochemistry and function. *J Leukoc Biol* 1989;46:63-74.
- 1656 8 Athens JW, Haab OP, Raab SO, Mauer AM, Ashenbrucker H, Cartwright GE, Wintrobe MM.
1657 Leukokinetic studies. Iv. The total blood, circulating and marginal granulocyte pools and the
1658 granulocyte turnover rate in normal subjects. *J Clin Invest* 1961;40:989-995.
- 1659 9 Rankin SM. The bone marrow: A site of neutrophil clearance. *J Leukoc Biol* 2010;88:241-251.
- 1660 10 Segal AW. How neutrophils kill microbes. *Annu Rev Immunol* 2005;23:197-223.
- 1661 11 Hampton MB, Kettle AJ, Winterbourn CC. Inside the neutrophil phagosome: Oxidants,
1662 myeloperoxidase, and bacterial killing. *Blood* 1998;92:3007-3017.
- 1663 12 Carvalho TT, Borghi SM, Pinho-Ribeiro FA, Mizokami SS, Cunha TM, Ferreira SH, Cunha FQ,
1664 Casagrande R, Verri WA, Jr. Granulocyte-colony stimulating factor (g-csf)-induced mechanical
1665 hyperalgesia in mice: Role for peripheral tnfa, il-1beta and il-10. *Eur J Pharmacol*
1666 2015;749:62-72.
- 1667 13 Pillay J, den Braber I, Vrisekoop N, Kwast LM, de Boer RJ, Borghans JA, Tesselaar K,
1668 Koenderman L. In vivo labeling with 2h2o reveals a human neutrophil lifespan of 5.4 days.
1669 *Blood* 2010;116:625-627.
- 1670 14 Scapini P, Cassatella MA. Social networking of human neutrophils within the immune system.
1671 *Blood* 2014;124:710-719.
- 1672 15 Tanji-Matsuba K, van Eeden SF, Saito Y, Okazawa M, Klut ME, Hayashi S, Hogg JC.
1673 Functional changes in aging polymorphonuclear leukocytes. *Circulation* 1998;97:91-98.
- 1674 16 Zhang D, Chen G, Manwani D, Mortha A, Xu C, Faith JJ, Burk RD, Kunisaki Y, Jang JE,
1675 Scheiermann C, Merad M, Frenette PS. Neutrophil ageing is regulated by the microbiome.
1676 *Nature* 2015;525:528-532.
- 1677 17 Whyte MK, Meagher LC, MacDermot J, Haslett C. Impairment of function in aging neutrophils is
1678 associated with apoptosis. *J Immunol* 1993;150:5124-5134.
- 1679 18 Lopes F, Coelho FM, Costa VV, Vieira EL, Sousa LP, Silva TA, Vieira LQ, Teixeira MM, Pinho
1680 V. Resolution of neutrophilic inflammation by h2o2 in antigen-induced arthritis. *Arthritis Rheum*
1681 2011;63:2651-2660.

- 1682 19 Buckley CD, Gilroy DW, Serhan CN. Proresolving lipid mediators and mechanisms in the
1683 resolution of acute inflammation. *Immunity* 2014;40:315-327.
- 1684 20 Nemeth T, Mocsai A. The role of neutrophils in autoimmune diseases. *Immunol Lett*
1685 2012;143:9-19.
- 1686 21 Xiaoxiao W, Sibiao Y, Xiaopeng X, Ping Z, Gang C. Neutrophils induce the maturation of
1687 immature dendritic cells: A regulatory role of neutrophils in adaptive immune responses.
1688 *Immunol Invest* 2007;36:337-350.
- 1689 22 Beauvillain C, Cunin P, Doni A, Scotet M, Jaillon S, Loiry ML, Magistrelli G, Masternak K,
1690 Chevallier A, Delneste Y, Jeannin P. Ccr7 is involved in the migration of neutrophils to lymph
1691 nodes. *Blood* 2011;117:1196-1204.
- 1692 23 Matsushima H, Geng S, Lu R, Okamoto T, Yao Y, Mayuzumi N, Kotol PF, Chojnacki BJ,
1693 Miyazaki T, Gallo RL, Takashima A. Neutrophil differentiation into a unique hybrid population
1694 exhibiting dual phenotype and functionality of neutrophils and dendritic cells. *Blood*
1695 2013;121:1677-1689.
- 1696 24 Davey MS, Morgan MP, Liuzzi AR, Tyler CJ, Khan MW, Szakmany T, Hall JE, Moser B, Eberl
1697 M. Microbe-specific unconventional t cells induce human neutrophil differentiation into antigen
1698 cross-presenting cells. *J Immunol* 2014;193:3704-3716.
- 1699 25 Riise RE, Bernson E, Aurelius J, Martner A, Pesce S, Della Chiesa M, Marcenaro E, Bylund J,
1700 Hellstrand K, Moretta L, Moretta A, Thoren FB. Tlr-stimulated neutrophils instruct nk cells to
1701 trigger dendritic cell maturation and promote adaptive t cell responses. *J Immunol*
1702 2015;195:1121-1128.
- 1703 26 Takashima A, Yao Y. Neutrophil plasticity: Acquisition of phenotype and functionality of antigen-
1704 presenting cell. *J Leukoc Biol* 2015;98:489-496.
- 1705 27 Eash KJ, Greenbaum AM, Gopalan PK, Link DC. Cxcr2 and cxcr4 antagonistically regulate
1706 neutrophil trafficking from murine bone marrow. *J Clin Invest* 2010;120:2423-2431.
- 1707 28 Sadik CD, Kim ND, Luster AD. Neutrophils cascading their way to inflammation. *Trends*
1708 *Immunol* 2011;32:452-460.
- 1709 29 Vestweber D. How leukocytes cross the vascular endothelium. *Nat Rev Immunol* 2015;15:692-
1710 704.
- 1711 30 Russo RC, Garcia CC, Teixeira MM, Amaral FA. The cxcl8/il-8 chemokine family and its
1712 receptors in inflammatory diseases. *Expert Rev Clin Immunol* 2014;10:593-619.
- 1713 31 Martin EL, Souza DG, Fagundes CT, Amaral FA, Assenzio B, Puntorieri V, Del Sorbo L, Fanelli
1714 V, Bosco M, Delsedime L, Pinho JF, Lemos VS, Souto FO, Alves-Filho JC, Cunha FQ, Slutsky
1715 AS, Ruckle T, Hirsch E, Teixeira MM, Ranieri VM. Phosphoinositide-3 kinase gamma activity
1716 contributes to sepsis and organ damage by altering neutrophil recruitment. *Am J Respir Crit*
1717 *Care Med* 2010;182:762-773.
- 1718 32 Alves-Filho JC, Sonogo F, Souto FO, Freitas A, Verri WA, Jr., Auxiliadora-Martins M, Basile-
1719 Filho A, McKenzie AN, Xu D, Cunha FQ, Liew FY. Interleukin-33 attenuates sepsis by
1720 enhancing neutrophil influx to the site of infection. *Nat Med* 2010;16:708-712.
- 1721 33 Souto FO, Alves-Filho JC, Turato WM, Auxiliadora-Martins M, Basile-Filho A, Cunha FQ.
1722 Essential role of ccr2 in neutrophil tissue infiltration and multiple organ dysfunction in sepsis. *Am*
1723 *J Respir Crit Care Med* 2011;183:234-242.
- 1724 34 McDonald B, Pittman K, Menezes GB, Hirota SA, Slaba I, Waterhouse CC, Beck PL, Muruve
1725 DA, Kuberski P. Intravascular danger signals guide neutrophils to sites of sterile inflammation.
1726 *Science* 2010;330:362-366.

- 1727 35 Foxman EF, Campbell JJ, Butcher EC. Multistep navigation and the combinatorial control of
1728 leukocyte chemotaxis. *J Cell Biol* 1997;139:1349-1360.
- 1729 36 Vanheule V, Janssens R, Boff D, Kitic N, Berghmans N, Ronsse I, Kungl AJ, Amaral FA,
1730 Teixeira MM, Van Damme J, Proost P, Mortier A. The positively charged cooh-terminal
1731 glycosaminoglycan-binding cxcl9(74-103) peptide inhibits cxcl8-induced neutrophil
1732 extravasation and monosodium urate crystal-induced gout in mice. *J Biol Chem*
1733 2015;290:21292-21304.
- 1734 37 Gerliza T, Hecher B, Jeremic D, Fuchs T, Gschwandtner M, Falsone A, Gesslbauer B, Kungl AJ.
1735 A combinatorial approach to biophysically characterise chemokine-glycan binding affinities for
1736 drug development. *Molecules* 2014;19:10618-10634.
- 1737 38 Woodfin A, Voisin MB, Beyrau M, Colom B, Caille D, Diapouli FM, Nash GB, Chavakis T,
1738 Albelda SM, Rainger GE, Meda P, Imhof BA, Nourshargh S. The junctional adhesion molecule
1739 jam-c regulates polarized transendothelial migration of neutrophils in vivo. *Nat Immunol*
1740 2011;12:761-769.
- 1741 39 Tauzin S, Starnes TW, Becker FB, Lam PY, Huttenlocher A. Redox and src family kinase
1742 signaling control leukocyte wound attraction and neutrophil reverse migration. *J Cell Biol*
1743 2014;207:589-598.
- 1744 40 Ellett F, Elks PM, Robertson AL, Ogryzko NV, Renshaw SA. Defining the phenotype of
1745 neutrophils following reverse migration in zebrafish. *J Leukoc Biol* 2015
- 1746 41 Colom B, Bodkin JV, Beyrau M, Woodfin A, Ody C, Rourke C, Chavakis T, Brohi K, Imhof BA,
1747 Nourshargh S. Leukotriene b4-neutrophil elastase axis drives neutrophil reverse
1748 transendothelial cell migration in vivo. *Immunity* 2015;42:1075-1086.
- 1749 42 Nordenfelt P, Tapper H. Phagosome dynamics during phagocytosis by neutrophils. *J Leukoc*
1750 *Biol* 2011;90:271-284.
- 1751 43 Nauseef WM. Myeloperoxidase in human neutrophil host defence. *Cell Microbiol* 2014;16:1146-
1752 1155.
- 1753 44 Metzler KD, Fuchs TA, Nauseef WM, Reumaux D, Roesler J, Schulze I, Wahn V,
1754 Papayannopoulos V, Zychlinsky A. Myeloperoxidase is required for neutrophil extracellular trap
1755 formation: Implications for innate immunity. *Blood* 2011;117:953-959.
- 1756 45 Brechard S, Plancon S, Tschirhart EJ. New insights into the regulation of neutrophil nadph
1757 oxidase activity in the phagosome: A focus on the role of lipid and ca(2+) signaling. *Antioxid*
1758 *Redox Signal* 2013;18:661-676.
- 1759 46 Fernandes ES, Vong CT, Quek S, Cheong J, Awal S, Gentry C, Aubdool AA, Liang L, Bodkin
1760 JV, Bevan S, Heads R, Brain SD. Superoxide generation and leukocyte accumulation: Key
1761 elements in the mediation of leukotriene b(4)-induced itch by transient receptor potential ankyrin
1762 1 and transient receptor potential vanilloid 1. *FASEB J* 2013;27:1664-1673.
- 1763 47 Serafim KG, Navarro SA, Zarpelon AC, Pinho-Ribeiro FA, Fattori V, Cunha TM, Alves-Filho JC,
1764 Cunha FQ, Casagrande R, Verri WA, Jr. Bosentan, a mixed endothelin receptor antagonist,
1765 inhibits superoxide anion-induced pain and inflammation in mice. *Naunyn Schmiedebergs Arch*
1766 *Pharmacol* 2015
- 1767 48 Yamacita-Borin FY, Zarpelon AC, Pinho-Ribeiro FA, Fattori V, Alves-Filho JC, Cunha FQ,
1768 Cunha TM, Casagrande R, Verri WA, Jr. Superoxide anion-induced pain and inflammation
1769 depends on tnfa/tnfr1 signaling in mice. *Neurosci Lett* 2015;605:53-58.
- 1770 49 Fattori V, Pinho-Ribeiro FA, Borghi SM, Alves-Filho JC, Cunha TM, Cunha FQ, Casagrande R,
1771 Verri WA, Jr. Curcumin inhibits superoxide anion-induced pain-like behavior and leukocyte
1772 recruitment by increasing nrf2 expression and reducing nf-kappab activation. *Inflamm Res* 2015

- 1773 50 Maioli NA, Zarpelon AC, Mizokami SS, Calixto-Campos C, Guazelli CF, Hohmann MS, Pinho-
1774 Ribeiro FA, Carvalho TT, Manchope MF, Ferraz CR, Casagrande R, Verri Jr WA. The
1775 superoxide anion donor, potassium superoxide, induces pain and inflammation in mice through
1776 production of reactive oxygen species and cyclooxygenase-2. *Braz J Med Biol Res* 2015;0:0.
- 1777 51 Wang ZQ, Porreca F, Cuzzocrea S, Galen K, Lightfoot R, Masini E, Muscoli C, Mollace V,
1778 Ndengele M, Ischiropoulos H, Salvemini D. A newly identified role for superoxide in
1779 inflammatory pain. *J Pharmacol Exp Ther* 2004;309:869-878.
- 1780 52 Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, Weinrauch Y,
1781 Zychlinsky A. Neutrophil extracellular traps kill bacteria. *Science* 2004;303:1532-1535.
- 1782 53 Branzk N, Papayannopoulos V. Molecular mechanisms regulating netosis in infection and
1783 disease. *Semin Immunopathol* 2013;35:513-530.
- 1784 54 Yipp BG, Kubes P. Netosis: How vital is it? *Blood* 2013;122:2784-2794.
- 1785 55 Kobayashi SD, Braughton KR, Palazzolo-Ballance AM, Kennedy AD, Sampaio E, Kristosturyan
1786 E, Whitney AR, Sturdevant DE, Dorward DW, Holland SM, Kreiswirth BN, Musser JM, DeLeo
1787 FR. Rapid neutrophil destruction following phagocytosis of staphylococcus aureus. *J Innate
1788 Immun* 2010;2:560-575.
- 1789 56 Fuchs TA, Abed U, Goosmann C, Hurwitz R, Schulze I, Wahn V, Weinrauch Y, Brinkmann V,
1790 Zychlinsky A. Novel cell death program leads to neutrophil extracellular traps. *J Cell Biol*
1791 2007;176:231-241.
- 1792 57 Yipp BG, Petri B, Salina D, Jenne CN, Scott BN, Zbytniuk LD, Pittman K, Asaduzzaman M, Wu
1793 K, Meijndert HC, Malawista SE, de Boisleury Chevance A, Zhang K, Conly J, Kubes P.
1794 Infection-induced netosis is a dynamic process involving neutrophil multitasking in vivo. *Nat Med*
1795 2012;18:1386-1393.
- 1796 58 Pilsczek FH, Salina D, Poon KK, Fahey C, Yipp BG, Sibley CD, Robbins SM, Green FH, Surette
1797 MG, Sugai M, Bowden MG, Hussain M, Zhang K, Kubes P. A novel mechanism of rapid nuclear
1798 neutrophil extracellular trap formation in response to staphylococcus aureus. *J Immunol*
1799 2010;185:7413-7425.
- 1800 59 Schauer C, Janko C, Munoz LE, Zhao Y, Kienhofer D, Frey B, Lell M, Manger B, Rech J,
1801 Naschberger E, Holmdahl R, Krenn V, Harrer T, Jeremic I, Bilyy R, Schett G, Hoffmann M,
1802 Herrmann M. Aggregated neutrophil extracellular traps limit inflammation by degrading
1803 cytokines and chemokines. *Nat Med* 2014;20:511-517.
- 1804 60 Jenne CN, Wong CH, Zemp FJ, McDonald B, Rahman MM, Forsyth PA, McFadden G, Kubes
1805 P. Neutrophils recruited to sites of infection protect from virus challenge by releasing neutrophil
1806 extracellular traps. *Cell Host Microbe* 2013;13:169-180.
- 1807 61 McDonald B, Urrutia R, Yipp BG, Jenne CN, Kubes P. Intravascular neutrophil extracellular
1808 traps capture bacteria from the bloodstream during sepsis. *Cell Host Microbe* 2012;12:324-333.
- 1809 62 Urban CF, Reichard U, Brinkmann V, Zychlinsky A. Neutrophil extracellular traps capture and
1810 kill candida albicans yeast and hyphal forms. *Cell Microbiol* 2006;8:668-676.
- 1811 63 Rollet-Labelle E, Vaillancourt M, Marois L, Newkirk MM, Poubelle PE, Naccache PH. Cross-
1812 linking of iggs bound on circulating neutrophils leads to an activation of endothelial cells:
1813 Possible role of rheumatoid factors in rheumatoid arthritis-associated vascular dysfunction. *J
1814 Inflamm (Lond)* 2013;10:27.
- 1815 64 Jonsson H, Allen P, Peng SL. Inflammatory arthritis requires foxo3a to prevent fas ligand-
1816 induced neutrophil apoptosis. *Nat Med* 2005;11:666-671.

- 1817 65 Quayle JA, Watson F, Bucknall RC, Edwards SW. Neutrophils from the synovial fluid of patients
1818 with rheumatoid arthritis express the high affinity immunoglobulin g receptor, fc gamma ri
1819 (cd64): Role of immune complexes and cytokines in induction of receptor expression.
1820 *Immunology* 1997;91:266-273.
- 1821 66 Kundu S, Ghosh P, Datta S, Ghosh A, Chattopadhyay S, Chatterjee M. Oxidative stress as a
1822 potential biomarker for determining disease activity in patients with rheumatoid arthritis. *Free*
1823 *Radic Res* 2012;46:1482-1489.
- 1824 67 Jarvis JN, Petty HR, Tang Y, Frank MB, Tessier PA, Dozmorov I, Jiang K, Kindzelski A, Chen Y,
1825 Cadwell C, Turner M, Szodoray P, McGhee JL, Centola M. Evidence for chronic, peripheral
1826 activation of neutrophils in polyarticular juvenile rheumatoid arthritis. *Arthritis Res Ther*
1827 2006;8:R154.
- 1828 68 Kowanko IC, Bates EJ, Ferrante A. Mechanisms of human neutrophil-mediated cartilage
1829 damage in vitro: The role of lysosomal enzymes, hydrogen peroxide and hypochlorous acid.
1830 *Immunol Cell Biol* 1989;67 (Pt 5):321-329.
- 1831 69 Panasencko OM, Gorudko IV, Sokolov AV. Hypochlorous acid as a precursor of free radicals in
1832 living systems. *Biochemistry (Mosc)* 2013;78:1466-1489.
- 1833 70 Gardi C, Bauerova K, Stringa B, Kuncirova V, Slovak L, Ponist S, Drafi F, Bezakova L, Tedesco
1834 I, Acquaviva A, Bilotto S, Russo GL. Quercetin reduced inflammation and increased antioxidant
1835 defense in rat adjuvant arthritis. *Arch Biochem Biophys* 2015;583:150-157.
- 1836 71 Jeyadevi R, Sivasudha T, Rameshkumar A, Ananth DA, Aseervatham GS, Kumaresan K,
1837 Kumar LD, Jagadeeswari S, Renganathan R. Enhancement of anti arthritic effect of quercetin
1838 using thioglycolic acid-capped cadmium telluride quantum dots as nanocarrier in adjuvant
1839 induced arthritic wistar rats. *Colloids Surf B Biointerfaces* 2013;112:255-263.
- 1840 72 Mun SH, Kim HS, Kim JW, Ko NY, Kim do K, Lee BY, Kim B, Won HS, Shin HS, Han JW, Lee
1841 HY, Kim YM, Choi WS. Oral administration of curcumin suppresses production of matrix
1842 metalloproteinase (mmp)-1 and mmp-3 to ameliorate collagen-induced arthritis: Inhibition of the
1843 pkcdelta/jnk/c-jun pathway. *J Pharmacol Sci* 2009;111:13-21.
- 1844 73 Moon DO, Kim MO, Choi YH, Park YM, Kim GY. Curcumin attenuates inflammatory response in
1845 il-1beta-induced human synovial fibroblasts and collagen-induced arthritis in mouse model. *Int*
1846 *Immunopharmacol* 2010;10:605-610.
- 1847 74 Pruijn GJ. Citrullination and carbamylation in the pathophysiology of rheumatoid arthritis. *Front*
1848 *Immunol* 2015;6:192.
- 1849 75 Li P, Li M, Lindberg MR, Kennett MJ, Xiong N, Wang Y. Pad4 is essential for antibacterial innate
1850 immunity mediated by neutrophil extracellular traps. *J Exp Med* 2010;207:1853-1862.
- 1851 76 Khandpur R, Carmona-Rivera C, Vivekanandan-Giri A, Gizinski A, Yalavarthi S, Knight JS,
1852 Friday S, Li S, Patel RM, Subramanian V, Thompson P, Chen P, Fox DA, Pennathur S, Kaplan
1853 MJ. Nets are a source of citrullinated autoantigens and stimulate inflammatory responses in
1854 rheumatoid arthritis. *Sci Transl Med* 2013;5:178ra140.
- 1855 77 Romero V, Fert-Bober J, Nigrovic PA, Darrah E, Haque UJ, Lee DM, van Eyk J, Rosen A,
1856 Andrade F. Immune-mediated pore-forming pathways induce cellular hypercitrullination and
1857 generate citrullinated autoantigens in rheumatoid arthritis. *Sci Transl Med* 2013;5:209ra150.
- 1858 78 Scally SW, Petersen J, Law SC, Dudek NL, Nel HJ, Loh KL, Wijeyewickrema LC, Eckle SB, van
1859 Heemst J, Pike RN, McCluskey J, Toes RE, La Gruta NL, Purcell AW, Reid HH, Thomas R,
1860 Rossjohn J. A molecular basis for the association of the hla-drb1 locus, citrullination, and
1861 rheumatoid arthritis. *J Exp Med* 2013;210:2569-2582.

- 1862 79 Garcia-Romo GS, Caielli S, Vega B, Connolly J, Allantaz F, Xu Z, Punaro M, Baisch J, Guiducci
1863 C, Coffman RL, Barrat FJ, Banchereau J, Pascual V. Netting neutrophils are major inducers of
1864 type I IFN production in pediatric systemic lupus erythematosus. *Sci Transl Med* 2011;3:73ra20.
- 1865 80 Villanueva E, Yalavarthi S, Berthier CC, Hodgins JB, Khandpur R, Lin AM, Rubin CJ, Zhao W,
1866 Olsen SH, Klinker M, Shealy D, Denny MF, Plumas J, Chaperot L, Kretzler M, Bruce AT, Kaplan
1867 MJ. Netting neutrophils induce endothelial damage, infiltrate tissues, and expose
1868 immunostimulatory molecules in systemic lupus erythematosus. *J Immunol* 2011;187:538-552.
- 1869 81 Spengler J, Lugonja B, Ytterberg AJ, Zubarev RA, Creese AJ, Pearson MJ, Grant MM, Milward
1870 M, Lundberg K, Buckley CD, Filer A, Raza K, Cooper PR, Chapple IL, Scheel-Toellner D.
1871 Release of active peptidyl arginine deiminases by neutrophils can explain production of
1872 extracellular citrullinated autoantigens in rat synovial fluid. *Arthritis Rheumatol* 2015
- 1873 82 Bianchi M, Niemiec MJ, Siler U, Urban CF, Reichenbach J. Restoration of anti-aspergillus
1874 defense by neutrophil extracellular traps in human chronic granulomatous disease after gene
1875 therapy is calprotectin-dependent. *J Allergy Clin Immunol* 2011;127:1243-1252 e1247.
- 1876 83 Rohm M, Grimm MJ, D'Auria AC, Almyroudis NG, Segal BH, Urban CF. NADPH oxidase
1877 promotes neutrophil extracellular trap formation in pulmonary aspergillosis. *Infect Immun*
1878 2014;82:1766-1777.
- 1879 84 Phelps P, McCarty DJ, Jr. Crystal-induced inflammation in canine joints. II. Importance of
1880 polymorphonuclear leukocytes. *J Exp Med* 1966;124:115-126.
- 1881 85 Chakravarti A, Raquil MA, Tessier P, Poubelle PE. Surface rankin of toll-like receptor 4-
1882 stimulated human neutrophils activates osteoclastic bone resorption. *Blood* 2009;114:1633-
1883 1644.
- 1884 86 Ha J, Choi HS, Lee Y, Kwon HJ, Song YW, Kim HH. CXCL2 induced by
1885 receptor activator of NF- κ B ligand enhances osteoclastogenesis. *J Immunol*
1886 2010;184:4717-4724.
- 1887 87 Edwards SW, Hallett MB. Seeing the wood for the trees: The forgotten role of neutrophils in
1888 rheumatoid arthritis. *Immunol Today* 1997;18:320-324.
- 1889 88 Eijkelenboom A, Burgering BM. FoxO3: Signalling integrators for homeostasis maintenance. *Nat*
1890 *Rev Mol Cell Biol* 2013;14:83-97.
- 1891 89 Lee JC, Espeli M, Anderson CA, Linterman MA, Pocock JM, Williams NJ, Roberts R, Viatte S,
1892 Fu B, Peshu N, Hien TT, Phu NH, Wesley E, Edwards C, Ahmad T, Mansfield JC, Geary R,
1893 Dunstan S, Williams TN, Barton A, Vinuesa CG, Consortium UIG, Parkes M, Lyons PA, Smith
1894 KG. Human SNP links differential outcomes in inflammatory and infectious disease to a FoxO3-
1895 regulated pathway. *Cell* 2013;155:57-69.
- 1896 90 Turrel-Davin F, Tournadre A, Pachot A, Arnaud B, Cazalis MA, Mouglin B, Miossec P. FoxO3a
1897 involved in neutrophil and T cell survival is overexpressed in rheumatoid blood and synovial
1898 tissue. *Ann Rheum Dis* 2010;69:755-760.
- 1899 91 Martin WJ, Walton M, Harper J. Resident macrophages initiating and driving inflammation in a
1900 monosodium urate monohydrate crystal-induced murine peritoneal model of acute gout. *Arthritis*
1901 *Rheum* 2009;60:281-289.
- 1902 92 Chang YH, Garalla EJ. Suppression of urate crystal-induced canine joint inflammation by
1903 heterologous anti-polymorphonuclear leukocyte serum. *Arthritis Rheum* 1968;11:145-150.
- 1904 93 Martinon F, Petrilli V, Mayor A, Tardivel A, Tschopp J. Gout-associated uric acid crystals
1905 activate the NALP3 inflammasome. *Nature* 2006;440:237-241.

- 1906 94 Amaral FA, Costa VV, Tavares LD, Sachs D, Coelho FM, Fagundes CT, Soriani FM, Silveira
1907 TN, Cunha LD, Zamboni DS, Quesniaux V, Peres RS, Cunha TM, Cunha FQ, Ryffel B, Souza
1908 DG, Teixeira MM. Nlrp3 inflammasome-mediated neutrophil recruitment and hypernociception
1909 depend on leukotriene b(4) in a murine model of gout. *Arthritis Rheum* 2012;64:474-484.
- 1910 95 Guerrero AT, Verri WA, Jr., Cunha TM, Silva TA, Schivo IR, Dal-Secco D, Canetti C, Rocha FA,
1911 Parada CA, Cunha FQ, Ferreira SH. Involvement of ltb4 in zymosan-induced joint nociception in
1912 mice: Participation of neutrophils and pge2. *J Leukoc Biol* 2008;83:122-130.
- 1913 96 Ting E, Guerrero AT, Cunha TM, Verri WA, Jr., Taylor SM, Woodruff TM, Cunha FQ, Ferreira
1914 SH. Role of complement c5a in mechanical inflammatory hypernociception: Potential use of c5a
1915 receptor antagonists to control inflammatory pain. *Br J Pharmacol* 2008;153:1043-1053.
- 1916 97 Cunha TM, Verri WA, Jr., Schivo IR, Napimoga MH, Parada CA, Poole S, Teixeira MM, Ferreira
1917 SH, Cunha FQ. Crucial role of neutrophils in the development of mechanical inflammatory
1918 hypernociception. *J Leukoc Biol* 2008;83:824-832.
- 1919 98 Verri WA, Jr., Cunha TM, Magro DA, Guerrero AT, Vieira SM, Carregaro V, Souza GR,
1920 Henriques M, Ferreira SH, Cunha FQ. Targeting endothelin eta and etb receptors inhibits
1921 antigen-induced neutrophil migration and mechanical hypernociception in mice. *Naunyn
1922 Schmiedebergs Arch Pharmacol* 2009;379:271-279.
- 1923 99 Kaandorp CJ, Dinant HJ, van de Laar MA, Moens HJ, Prins AP, Dijkmans BA. Incidence and
1924 sources of native and prosthetic joint infection: A community based prospective survey. *Ann
1925 Rheum Dis* 1997;56:470-475.
- 1926 100 Sharff KA, Richards EP, Townes JM. Clinical management of septic arthritis. *Curr Rheumatol
1927 Rep* 2013;15:332.
- 1928 101 Kaplan SL. Recent lessons for the management of bone and joint infections. *J Infect* 2014;68
1929 Suppl 1:S51-56.
- 1930 102 Verdrengh M, Tarkowski A. Role of neutrophils in experimental septicemia and septic arthritis
1931 induced by staphylococcus aureus. *Infect Immun* 1997;65:2517-2521.
- 1932 103 Ali A, Na M, Svensson MN, Magnusson M, Welin A, Schwarze JC, Mohammad M, Josefsson E,
1933 Pullerits R, Jin T. Il-1 receptor antagonist treatment aggravates staphylococcal septic arthritis
1934 and sepsis in mice. *PLoS One* 2015;10:e0131645.
- 1935 104 Gjertsson I, Hultgren OH, Collins LV, Pettersson S, Tarkowski A. Impact of transcription factors
1936 ap-1 and nf-kappab on the outcome of experimental staphylococcus aureus arthritis and sepsis.
1937 *Microbes Infect* 2001;3:527-534.
- 1938 105 Alves-Filho JC, de Freitas A, Spiller F, Souto FO, Cunha FQ. The role of neutrophils in severe
1939 sepsis. *Shock* 2008;30 Suppl 1:3-9.
- 1940 106 Verri WA, Jr., Souto FO, Vieira SM, Almeida SC, Fukada SY, Xu D, Alves-Filho JC, Cunha TM,
1941 Guerrero AT, Mattos-Guimaraes RB, Oliveira FR, Teixeira MM, Silva JS, McInnes IB, Ferreira
1942 SH, Louzada-Junior P, Liew FY, Cunha FQ. Il-33 induces neutrophil migration in rheumatoid
1943 arthritis and is a target of anti-tnf therapy. *Ann Rheum Dis* 2010;69:1697-1703.
- 1944 107 Verri WA, Jr., Cunha TM, Parada CA, Poole S, Cunha FQ, Ferreira SH. Hypernociceptive role
1945 of cytokines and chemokines: Targets for analgesic drug development? *Pharmacol Ther*
1946 2006;112:116-138.
- 1947 108 Rocha FA, Leite AK, Pompeu MM, Cunha TM, Verri WA, Jr., Soares FM, Castro RR, Cunha
1948 FQ. Protective effect of an extract from ascaris suum in experimental arthritis models. *Infect
1949 Immun* 2008;76:2736-2745.

- 1950 109 Conte FP, Menezes-de-Lima O, Jr., Verri WA, Jr., Cunha FQ, Penido C, Henriques MG. Lipoxin
1951 a(4) attenuates zymosan-induced arthritis by modulating endothelin-1 and its effects. *Br J*
1952 *Pharmacol* 2010;161:911-924.
- 1953 110 Bachelerie F, Ben-Baruch A, Burkhardt AM, Combadiere C, Farber JM, Graham GJ, Horuk R,
1954 Sparre-Ulrich AH, Locati M, Luster AD, Mantovani A, Matsushima K, Murphy PM, Nibbs R,
1955 Nomiyama H, Power CA, Proudfoot AE, Rosenkilde MM, Rot A, Sozzani S, Thelen M, Yoshie
1956 O, Zlotnik A. International union of basic and clinical pharmacology. [corrected]. Lxxxix. Update
1957 on the extended family of chemokine receptors and introducing a new nomenclature for atypical
1958 chemokine receptors. *Pharmacol Rev* 2014;66:1-79.
- 1959 111 Cunha TM, Barsante MM, Guerrero AT, Verri WA, Jr., Ferreira SH, Coelho FM, Bertini R, Di
1960 Giacinto C, Allegretti M, Cunha FQ, Teixeira MM. Treatment with df 2162, a non-competitive
1961 allosteric inhibitor of cxcr1/2, diminishes neutrophil influx and inflammatory hypernociception in
1962 mice. *Br J Pharmacol* 2008;154:460-470.
- 1963 112 Coelho FM, Pinho V, Amaral FA, Sachs D, Costa VV, Rodrigues DH, Vieira AT, Silva TA, Souza
1964 DG, Bertini R, Teixeira AL, Teixeira MM. The chemokine receptors cxcr1/cxcr2 modulate
1965 antigen-induced arthritis by regulating adhesion of neutrophils to the synovial microvasculature.
1966 *Arthritis Rheum* 2008;58:2329-2337.
- 1967 113 Lemos HP, Grespan R, Vieira SM, Cunha TM, Verri WA, Jr., Fernandes KS, Souto FO, McInnes
1968 IB, Ferreira SH, Liew FY, Cunha FQ. Prostaglandin mediates il-23/il-17-induced neutrophil
1969 migration in inflammation by inhibiting il-12 and ifngamma production. *Proc Natl Acad Sci U S A*
1970 2009;106:5954-5959.
- 1971 114 Min SH, Wang Y, Gonsiorek W, Anilkumar G, Kozlowski J, Lundell D, Fine JS, Grant EP.
1972 Pharmacological targeting reveals distinct roles for cxcr2/cxcr1 and ccr2 in a mouse model of
1973 arthritis. *Biochem Biophys Res Commun* 2010;391:1080-1086.
- 1974 115 Sarma JV, Ward PA. New developments in c5a receptor signaling. *Cell Health Cytoskelet*
1975 2012;4:73-82.
- 1976 116 Andersson C, Wenander CS, Usher PA, Hebsgaard JB, Sondergaard BC, Rono B, Mackay C,
1977 Friedrichsen B, Chang C, Tang R, Hornum L. Rapid-onset clinical and mechanistic effects of
1978 anti-c5ar treatment in the mouse collagen-induced arthritis model. *Clin Exp Immunol*
1979 2014;177:219-233.
- 1980 117 Mehta G, Scheinman RI, Holers VM, Banda NK. A new approach for the treatment of arthritis in
1981 mice with a novel conjugate of an anti-c5ar1 antibody and c5 small interfering rna. *J Immunol*
1982 2015;194:5446-5454.
- 1983 118 Sadik CD, Kim ND, Iwakura Y, Luster AD. Neutrophils orchestrate their own recruitment in
1984 murine arthritis through c5ar and fcgammar signaling. *Proc Natl Acad Sci U S A*
1985 2012;109:E3177-3185.
- 1986 119 Banda NK, Hyatt S, Antonioli AH, White JT, Glogowska M, Takahashi K, Merkel TJ, Stahl GL,
1987 Mueller-Ortiz S, Wetsel R, Arend WP, Holers VM. Role of c3a receptors, c5a receptors, and
1988 complement protein c6 deficiency in collagen antibody-induced arthritis in mice. *J Immunol*
1989 2012;188:1469-1478.
- 1990 120 Deree J, Lall R, Melbostad H, Grant M, Hoyt DB, Coimbra R. Neutrophil degranulation and the
1991 effects of phosphodiesterase inhibition. *J Surg Res* 2006;133:22-28.
- 1992 121 Bombini G, Canetti C, Rocha FA, Cunha FQ. Tumour necrosis factor-alpha mediates neutrophil
1993 migration to the knee synovial cavity during immune inflammation. *Eur J Pharmacol*
1994 2004;496:197-204.
- 1995 122 Amaral FA, Bastos LF, Oliveira TH, Dias AC, Oliveira VL, Tavares LD, Costa VV, Galvao I,
1996 Soriani FM, Szymkowski DE, Ryffel B, Souza DG, Teixeira MM. Transmembrane tnfr-alpha is

- 1997 sufficient for articular inflammation and hypernociception in a mouse model of gout. *Eur J Immunol* 2016;46:204-211.
- 1998
- 1999 123 Queiroz-Junior CM, Bessoni RL, Costa VV, Souza DG, Teixeira MM, Silva TA. Preventive and
2000 therapeutic anti-tnf-alpha therapy with pentoxifylline decreases arthritis and the associated
2001 periodontal co-morbidity in mice. *Life Sci* 2013;93:423-428.
- 2002 124 Monach PA, Mathis D, Benoist C. The k/bxn arthritis model. *Curr Protoc Immunol* 2008;Chapter
2003 15:Unit 15 22.
- 2004 125 Chen M, Lam BK, Kanaoka Y, Nigrovic PA, Audoly LP, Austen KF, Lee DM. Neutrophil-derived
2005 leukotriene b4 is required for inflammatory arthritis. *J Exp Med* 2006;203:837-842.
- 2006 126 Wu MY, Lin TH, Chiu YC, Liou HC, Yang RS, Fu WM. Involvement of 15-lipoxygenase in the
2007 inflammatory arthritis. *J Cell Biochem* 2012;113:2279-2289.
- 2008 127 Kim ND, Chou RC, Seung E, Tager AM, Luster AD. A unique requirement for the leukotriene b4
2009 receptor blt1 for neutrophil recruitment in inflammatory arthritis. *J Exp Med* 2006;203:829-835.
- 2010 128 Mathis SP, Jala VR, Lee DM, Haribabu B. Nonredundant roles for leukotriene b4 receptors blt1
2011 and blt2 in inflammatory arthritis. *J Immunol* 2010;185:3049-3056.
- 2012 129 Chou RC, Kim ND, Sadik CD, Seung E, Lan Y, Byrne MH, Haribabu B, Iwakura Y, Luster AD.
2013 Lipid-cytokine-chemokine cascade drives neutrophil recruitment in a murine model of
2014 inflammatory arthritis. *Immunity* 2010;33:266-278.
- 2015 130 Kuwabara K, Yasui K, Jyoyama H, Maruyama T, Fleisch JH, Hori Y. Effects of the second-
2016 generation leukotriene b(4) receptor antagonist, ly293111na, on leukocyte infiltration and
2017 collagen-induced arthritis in mice. *Eur J Pharmacol* 2000;402:275-285.
- 2018 131 Shao WH, Del Prete A, Bock CB, Haribabu B. Targeted disruption of leukotriene b4 receptors
2019 blt1 and blt2: A critical role for blt1 in collagen-induced arthritis in mice. *J Immunol*
2020 2006;176:6254-6261.
- 2021 132 Verri WA, Jr., Cunha TM, Ferreira SH, Wei X, Leung BP, Fraser A, McInnes IB, Liew FY, Cunha
2022 FQ. Il-15 mediates antigen-induced neutrophil migration by triggering il-18 production. *Eur J*
2023 *Immunol* 2007;37:3373-3380.
- 2024 133 Verri WA, Jr., Cunha TM, Parada CA, Wei XQ, Ferreira SH, Liew FY, Cunha FQ. Il-15 mediates
2025 immune inflammatory hypernociception by triggering a sequential release of ifn-gamma,
2026 endothelin, and prostaglandin. *Proc Natl Acad Sci U S A* 2006;103:9721-9725.
- 2027 134 Baslund B, Tvede N, Danneskiold-Samsoe B, Larsson P, Panayi G, Petersen J, Petersen LJ,
2028 Beurskens FJ, Schuurman J, van de Winkel JG, Parren PW, Gracie JA, Jongbloed S, Liew FY,
2029 McInnes IB. Targeting interleukin-15 in patients with rheumatoid arthritis: A proof-of-concept
2030 study. *Arthritis Rheum* 2005;52:2686-2692.
- 2031 135 Blaho VA, Zhang Y, Hughes-Hanks JM, Brown CR. 5-lipoxygenase-deficient mice infected with
2032 *borrelia burgdorferi* develop persistent arthritis. *J Immunol* 2011;186:3076-3084.
- 2033 136 Amlong CA, Nardelli DT, Peterson SH, Warner TF, Callister SM, Schell RF. Anti-interleukin-15
2034 prevents arthritis in borrelia-vaccinated and -infected mice. *Clin Vaccine Immunol* 2006;13:289-
2035 296.
- 2036 137 Jablonska E, Marcinczyk M, Talarek L, Pancewicz S, Hermanowska-Szpakowicz T, Jablonski J.
2037 Il-15 in the culture supernatants of pmn and pbmc and the serum of patients with lyme disease.
2038 *Rocz Akad Med Bialymst* 2003;48:78-81.
- 2039 138 Maruyama K, Akira S. Emerging molecules in the interface between skeletal system and innate
2040 immunity. *Pharmacol Res* 2015;99:223-228.

- 2041 139 Edwards JC, Cambridge G. B-cell targeting in rheumatoid arthritis and other autoimmune
2042 diseases. *Nat Rev Immunol* 2006;6:394-403.
- 2043 140 Smolen JS, Steiner G. Therapeutic strategies for rheumatoid arthritis. *Nat Rev Drug Discov*
2044 2003;2:473-488.
- 2045 141 Lubberts E. The il-23-il-17 axis in inflammatory arthritis. *Nat Rev Rheumatol* 2015;11:415-429.
- 2046 142 Wipke BT, Allen PM. Essential role of neutrophils in the initiation and progression of a murine
2047 model of rheumatoid arthritis. *J Immunol* 2001;167:1601-1608.
- 2048 143 Burmester GR, Feist E, Dorner T. Emerging cell and cytokine targets in rheumatoid arthritis. *Nat*
2049 *Rev Rheumatol* 2014;10:77-88.
- 2050 144 McInnes IB, Liew FY. Cytokine networks--towards new therapies for rheumatoid arthritis. *Nat*
2051 *Clin Pract Rheumatol* 2005;1:31-39.
- 2052 145 Weinmann P, Moura RA, Caetano-Lopes JR, Pereira PA, Canhao H, Queiroz MV, Fonseca JE.
2053 Delayed neutrophil apoptosis in very early rheumatoid arthritis patients is abrogated by
2054 methotrexate therapy. *Clin Exp Rheumatol* 2007;25:885-887.
- 2055 146 Majumdar S, Aggarwal BB. Methotrexate suppresses nf-kappab activation through inhibition of
2056 ikappabalpha phosphorylation and degradation. *J Immunol* 2001;167:2911-2920.
- 2057 147 Sperling RI, Benincaso AI, Anderson RJ, Coblyn JS, Austen KF, Weinblatt ME. Acute and
2058 chronic suppression of leukotriene b4 synthesis ex vivo in neutrophils from patients with
2059 rheumatoid arthritis beginning treatment with methotrexate. *Arthritis Rheum* 1992;35:376-384.
- 2060 148 Wessels JA, Huizinga TW, Guchelaar HJ. Recent insights in the pharmacological actions of
2061 methotrexate in the treatment of rheumatoid arthritis. *Rheumatology (Oxford)* 2008;47:249-255.
- 2062 149 Wright HL, Chikura B, Bucknall RC, Moots RJ, Edwards SW. Changes in expression of
2063 membrane tnf, nf- κ b activation and neutrophil apoptosis during active and resolved
2064 inflammation. *Ann Rheum Dis* 2011;70:537-543.
- 2065 150 Rios-Navarro C, de Pablo C, Collado-Diaz V, Orden S, Blas-Garcia A, Martinez-Cuesta MA,
2066 Esplugues JV, Alvarez A. Differential effects of anti-tnf-alpha and anti-il-12/23 agents on human
2067 leukocyte-endothelial cell interactions. *Eur J Pharmacol* 2015;765:355-365.
- 2068 151 Wittkowski H, Foell D, af Klint E, De Rycke L, De Keyser F, Frosch M, Ulfgren AK, Roth J.
2069 Effects of intra-articular corticosteroids and anti-tnf therapy on neutrophil activation in
2070 rheumatoid arthritis. *Ann Rheum Dis* 2007;66:1020-1025.
- 2071 152 Gelderman KA, Hultqvist M, Olsson LM, Bauer K, Pizzolla A, Olofsson P, Holmdahl R.
2072 Rheumatoid arthritis: The role of reactive oxygen species in disease development and
2073 therapeutic strategies. *Antioxid Redox Signal* 2007;9:1541-1567.
- 2074 153 Heasman SJ, Giles KM, Ward C, Rossi AG, Haslett C, Dransfield I. Glucocorticoid-mediated
2075 regulation of granulocyte apoptosis and macrophage phagocytosis of apoptotic cells:
2076 Implications for the resolution of inflammation. *J Endocrinol* 2003;178:29-36.
- 2077 154 Talabot-Ayer D, McKee T, Gindre P, Bas S, Baeten DL, Gabay C, Palmer G. Distinct serum and
2078 synovial fluid interleukin (il)-33 levels in rheumatoid arthritis, psoriatic arthritis and osteoarthritis.
2079 *Joint Bone Spine* 2012;79:32-37.
- 2080 155 Hong YS, Moon SJ, Joo YB, Jeon CH, Cho ML, Ju JH, Oh HJ, Heo YJ, Park SH, Kim HY, Min
2081 JK. Measurement of interleukin-33 (il-33) and il-33 receptors (sst2 and st2l) in patients with
2082 rheumatoid arthritis. *J Korean Med Sci* 2011;26:1132-1139.

- 2083 156 Kageyama Y, Torikai E, Tsujimura K, Kobayashi M. Involvement of il-33 in the pathogenesis of
2084 rheumatoid arthritis: The effect of etanercept on the serum levels of il-33. *Mod Rheumatol*
2085 2012;22:89-93.
- 2086 157 Natarajan V, Madhan B, Tiku ML. Intra-articular injections of polyphenols protect articular
2087 cartilage from inflammation-induced degradation: Suggesting a potential role in cartilage
2088 therapeutics. *PLoS One* 2015;10:e0127165.
- 2089 158 Khanna D, Sethi G, Ahn KS, Pandey MK, Kunnumakkara AB, Sung B, Aggarwal A, Aggarwal
2090 BB. Natural products as a gold mine for arthritis treatment. *Curr Opin Pharmacol* 2007;7:344-
2091 351.
- 2092 159 Aggarwal BB, Harikumar KB. Potential therapeutic effects of curcumin, the anti-inflammatory
2093 agent, against neurodegenerative, cardiovascular, pulmonary, metabolic, autoimmune and
2094 neoplastic diseases. *Int J Biochem Cell Biol* 2009;41:40-59.
- 2095 160 Souto FO, Zarpelon AC, Staurengo-Ferrari L, Fattori V, Casagrande R, Fonseca MJ, Cunha
2096 TM, Ferreira SH, Cunha FQ, Verri WA, Jr. Quercetin reduces neutrophil recruitment induced by
2097 cxcl8, ltb4, and fmlp: Inhibition of actin polymerization. *J Nat Prod* 2011;74:113-118.
- 2098 161 Larmonier CB, Midura-Kiela MT, Ramalingam R, Laubitz D, Janikashvili N, Larmonier N,
2099 Ghishan FK, Kiela PR. Modulation of neutrophil motility by curcumin: Implications for
2100 inflammatory bowel disease. *Inflamm Bowel Dis* 2011;17:503-515.
- 2101 162 Kim DC, Lee W, Bae JS. Vascular anti-inflammatory effects of curcumin on hmgb1-mediated
2102 responses in vitro. *Inflamm Res* 2011;60:1161-1168.
- 2103 163 Suyenaga ES, Klein-Junior LC, Passos Cdos S, Marin R, Santin JR, Machado ID, Farsky SH,
2104 Henriques AT. Beyond organoleptic characteristics: The pharmacological potential of flavonoids
2105 and their role in leukocyte migration and in l-selectin and beta2-integrin expression during
2106 inflammation. *Phytother Res* 2014;28:1406-1411.
- 2107 164 Luer S, Troller R, Aebi C. Antibacterial and antiinflammatory kinetics of curcumin as a potential
2108 antimucositis agent in cancer patients. *Nutr Cancer* 2012;64:975-981.
- 2109 165 Santos AM, Lopes T, Oleastro M, Gato IV, Floch P, Benejat L, Chaves P, Pereira T, Seixas E,
2110 Machado J, Guerreiro AS. Curcumin inhibits gastric inflammation induced by helicobacter pylori
2111 infection in a mouse model. *Nutrients* 2015;7:306-320.
- 2112 166 Deodhar SD, Sethi R, Srimal RC. Preliminary study on antirheumatic activity of curcumin
2113 (diferuloyl methane). *Indian J Med Res* 1980;71:632-634.
- 2114 167 Chandran B, Goel A. A randomized, pilot study to assess the efficacy and safety of curcumin in
2115 patients with active rheumatoid arthritis. *Phytother Res* 2012;26:1719-1725.
- 2116 168 Di Pierro F, Rapacioli G, Di Maio EA, Appendino G, Franceschi F, Togni S. Comparative
2117 evaluation of the pain-relieving properties of a lecithinized formulation of curcumin (meriva((r))),
2118 nimesulide, and acetaminophen. *J Pain Res* 2013;6:201-205.
- 2119 169 Belcaro G, Cesarone MR, Dugall M, Pellegrini L, Ledda A, Grossi MG, Togni S, Appendino G.
2120 Efficacy and safety of meriva(r), a curcumin-phosphatidylcholine complex, during extended
2121 administration in osteoarthritis patients. *Altern Med Rev* 2010;15:337-344.
- 2122 170 Myers SP, O'Connor J, Fitton JH, Brooks L, Rolfe M, Connellan P, Wohlmuth H, Cheras PA,
2123 Morris C. A combined phase i and ii open label study on the effects of a seaweed extract
2124 nutrient complex on osteoarthritis. *Biologics* 2010;4:33-44.
- 2125 171 Myers SP, O'Connor J, Fitton JH, Brooks L, Rolfe M, Connellan P, Wohlmuth H, Cheras PA,
2126 Morris C. A combined phase i and ii open-label study on the immunomodulatory effects of
2127 seaweed extract nutrient complex. *Biologics* 2011;5:45-60.

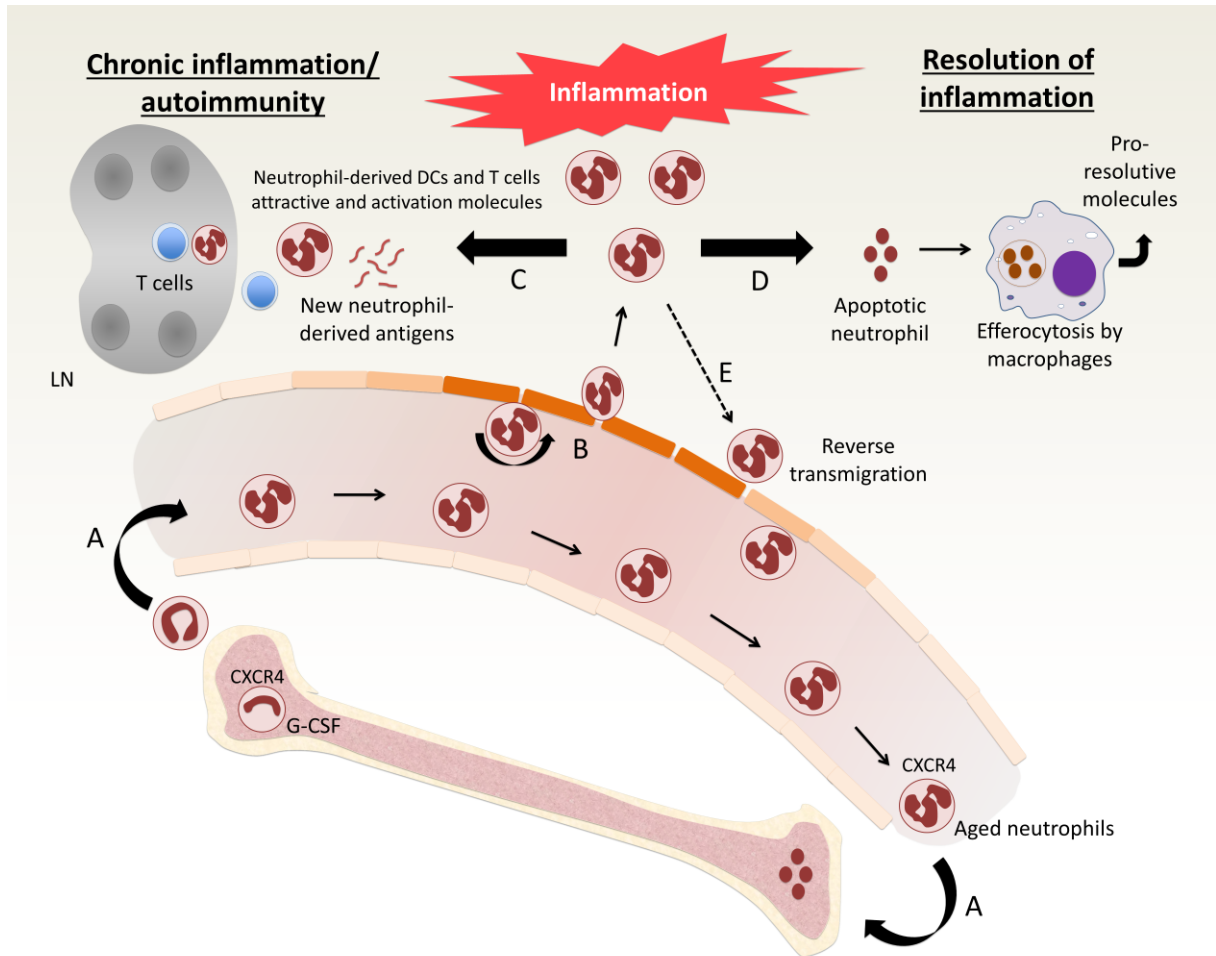
- 2128 172 Serhan CN, Chiang N, Van Dyke TE. Resolving inflammation: Dual anti-inflammatory and pro-
2129 resolution lipid mediators. *Nat Rev Immunol* 2008;8:349-361.
- 2130 173 Oh SF, Dona M, Fredman G, Krishnamoorthy S, Irimia D, Serhan CN. Resolvin e2 formation
2131 and impact in inflammation resolution. *J Immunol* 2012;188:4527-4534.
- 2132 174 Serhan CN, Maddox JF, Petasis NA, Akritopoulou-Zanze I, Papayianni A, Brady HR, Colgan
2133 SP, Madara JL. Design of lipoxin a4 stable analogs that block transmigration and adhesion of
2134 human neutrophils. *Biochemistry* 1995;34:14609-14615.
- 2135 175 Dufton N, Hannon R, Brancaleone V, Dalli J, Patel HB, Gray M, D'Acquisto F, Buckingham JC,
2136 Perretti M, Flower RJ. Anti-inflammatory role of the murine formyl-peptide receptor 2: Ligand-
2137 specific effects on leukocyte responses and experimental inflammation. *J Immunol*
2138 2010;184:2611-2619.
- 2139 176 Chan MM, Moore AR. Resolution of inflammation in murine autoimmune arthritis is disrupted by
2140 cyclooxygenase-2 inhibition and restored by prostaglandin e2-mediated lipoxin a4 production. *J*
2141 *Immunol* 2010;184:6418-6426.
- 2142 177 Lima-Garcia JF, Dutra RC, da Silva K, Motta EM, Campos MM, Calixto JB. The precursor of
2143 resolvin d series and aspirin-triggered resolvin d1 display anti-hyperalgesic properties in
2144 adjuvant-induced arthritis in rats. *Br J Pharmacol* 2011;164:278-293.
- 2145 178 Calder PC. Session 3: Joint nutrition society and irish nutrition and dietetic institute symposium
2146 on 'nutrition and autoimmune disease' pufa, inflammatory processes and rheumatoid arthritis.
2147 *Proc Nutr Soc* 2008;67:409-418.
- 2148 179 Norling LV, Dalli J, Flower RJ, Serhan CN, Perretti M. Resolvin d1 limits polymorphonuclear
2149 leukocyte recruitment to inflammatory loci: Receptor-dependent actions. *Arterioscler Thromb*
2150 *Vasc Biol* 2012;32:1970-1978.
- 2151 180 Cleland LG, Caughey GE, James MJ, Proudman SM. Reduction of cardiovascular risk factors
2152 with longterm fish oil treatment in early rheumatoid arthritis. *J Rheumatol* 2006;33:1973-1979.
- 2153 181 Gallai V, Sarchielli P, Trequattrini A, Franceschini M, Floridi A, Firenze C, Alberti A, Di
2154 Benedetto D, Stragliotto E. Cytokine secretion and eicosanoid production in the peripheral blood
2155 mononuclear cells of ms patients undergoing dietary supplementation with n-3 polyunsaturated
2156 fatty acids. *J Neuroimmunol* 1995;56:143-153.
- 2157 182 Caughey GE, Mantzioris E, Gibson RA, Cleland LG, James MJ. The effect on human tumor
2158 necrosis factor alpha and interleukin 1 beta production of diets enriched in n-3 fatty acids from
2159 vegetable oil or fish oil. *Am J Clin Nutr* 1996;63:116-122.
- 2160 183 Goldberg RJ, Katz J. A meta-analysis of the analgesic effects of omega-3 polyunsaturated fatty
2161 acid supplementation for inflammatory joint pain. *Pain* 2007;129:210-223.
- 2162 184 Seki H, Fukunaga K, Arita M, Arai H, Nakanishi H, Taguchi R, Miyasho T, Takamiya R, Asano
2163 K, Ishizaka A, Takeda J, Levy BD. The anti-inflammatory and proresolving mediator resolvin e1
2164 protects mice from bacterial pneumonia and acute lung injury. *J Immunol* 2010;184:836-843.
- 2165 185 Chiang N, Fredman G, Backhed F, Oh SF, Vickery T, Schmidt BA, Serhan CN. Infection
2166 regulates pro-resolving mediators that lower antibiotic requirements. *Nature* 2012;484:524-528.
- 2167 186 Norling LV, Perretti M. The role of omega-3 derived resolvins in arthritis. *Curr Opin Pharmacol*
2168 2013;13:476-481.
- 2169 187 Lee CH. Resolvins as new fascinating drug candidates for inflammatory diseases. *Arch Pharm*
2170 *Res* 2012;35:3-7.

- 2171 188 Ghasemzadeh M, Hosseini E. Platelet-leukocyte crosstalk: Linking proinflammatory responses
2172 to procoagulant state. *Thromb Res* 2013;131:191-197.
- 2173 189 Sreeramkumar V, Adrover JM, Ballesteros I, Cuartero MI, Rossaint J, Bilbao I, Nacher M,
2174 Pitaval C, Radovanovic I, Fukui Y, McEver RP, Filippi MD, Lizasoain I, Ruiz-Cabello J, Zarbock
2175 A, Moro MA, Hidalgo A. Neutrophils scan for activated platelets to initiate inflammation. *Science*
2176 2014;346:1234-1238.
- 2177 190 Etulain J, Martinod K, Wong SL, Cifuni SM, Schattner M, Wagner DD. P-selectin promotes
2178 neutrophil extracellular trap formation in mice. *Blood* 2015;126:242-246.
- 2179 191 Sumariwalla PF, Malfait AM, Feldmann M. P-selectin glycoprotein ligand 1 therapy ameliorates
2180 established collagen-induced arthritis in dba/1 mice partly through the suppression of tumour
2181 necrosis factor. *Clin Exp Immunol* 2004;136:67-75.
- 2182 192 Burrage PS, Mix KS, Brinckerhoff CE. Matrix metalloproteinases: Role in arthritis. *Front Biosci*
2183 2006;11:529-543.
- 2184 193 Itakura A, McCarty OJ. Pivotal role for the mtor pathway in the formation of neutrophil
2185 extracellular traps via regulation of autophagy. *Am J Physiol Cell Physiol* 2013;305:C348-354.
- 2186 194 Maugeri N, Campana L, Gavina M, Covino C, De Metrio M, Panciroli C, Maiuri L, Maseri A,
2187 D'Angelo A, Bianchi ME, Rovere-Querini P, Manfredi AA. Activated platelets present high
2188 mobility group box 1 to neutrophils, inducing autophagy and promoting the extrusion of
2189 neutrophil extracellular traps. *J Thromb Haemost* 2014;12:2074-2088.
- 2190 195 Boone BA, Orlichenko L, Schapiro NE, Loughran P, Gianfrate GC, Ellis JT, Singhi AD, Kang R,
2191 Tang D, Lotze MT, Zeh HJ. The receptor for advanced glycation end products (rage) enhances
2192 autophagy and neutrophil extracellular traps in pancreatic cancer. *Cancer Gene Ther*
2193 2015;22:326-334.
- 2194 196 Campbell JJ, Foxman EF, Butcher EC. Chemoattractant receptor cross talk as a regulatory
2195 mechanism in leukocyte adhesion and migration. *Eur J Immunol* 1997;27:2571-2578.
- 2196 197 Nourshargh S, Alon R. Leukocyte migration into inflamed tissues. *Immunity* 2014;41:694-707.
- 2197 198 Stark K, Eckart A, Haidari S, Tirniceriu A, Lorenz M, von Bruhl ML, Gartner F, Khandoga AG,
2198 Legate KR, Pless R, Hepper I, Lauber K, Walzog B, Massberg S. Capillary and arteriolar
2199 pericytes attract innate leukocytes exiting through venules and 'instruct' them with pattern-
2200 recognition and motility programs. *Nat Immunol* 2013;14:41-51.
- 2201 199 Proebstl D, Voisin MB, Woodfin A, Whiteford J, D'Acquisto F, Jones GE, Rowe D, Nourshargh
2202 S. Pericytes support neutrophil subendothelial cell crawling and breaching of venular walls in
2203 vivo. *J Exp Med* 2012;209:1219-1234.
- 2204 200 Tak PP, Taylor PC, Breedveld FC, Smeets TJ, Daha MR, Kluin PM, Meinders AE, Maini RN.
2205 Decrease in cellularity and expression of adhesion molecules by anti-tumor necrosis factor
2206 alpha monoclonal antibody treatment in patients with rheumatoid arthritis. *Arthritis Rheum*
2207 1996;39:1077-1081.
- 2208 201 Youssef PP, Triantafyllou S, Parker A, Coleman M, Roberts-Thomson PJ, Ahern MJ, Smith MD.
2209 Effects of pulse methylprednisolone on cell adhesion molecules in the synovial membrane in
2210 rheumatoid arthritis. Reduced e-selectin and intercellular adhesion molecule 1 expression.
2211 *Arthritis Rheum* 1996;39:1970-1979.
- 2212 202 Wilder RL. Integrin alpha v beta 3 as a target for treatment of rheumatoid arthritis and related
2213 rheumatic diseases. *Ann Rheum Dis* 2002;61 Suppl 2:ii96-99.
- 2214 203 Haringman JJ, Oostendorp RL, Tak PP. Targeting cellular adhesion molecules, chemokines and
2215 chemokine receptors in rheumatoid arthritis. *Expert Opin Emerg Drugs* 2005;10:299-310.

- 2216 204 Maksymowych WP, Blackburn WD, Jr., Tami JA, Shanahan WR, Jr. A randomized, placebo
2217 controlled trial of an antisense oligodeoxynucleotide to intercellular adhesion molecule-1 in the
2218 treatment of severe rheumatoid arthritis. *J Rheumatol* 2002;29:447-453.
- 2219 205 Bhushan M, Bleiker TO, Ballsdon AE, Allen MH, Sopwith M, Robinson MK, Clarke C, Weller RP,
2220 Graham-Brown RA, Keefe M, Barker JN, Griffiths CE. Anti-e-selectin is ineffective in the
2221 treatment of psoriasis: A randomized trial. *Br J Dermatol* 2002;146:824-831.
- 2222 206 Kraan MC, van Kuijk AW, Dinant HJ, Goedkoop AY, Smeets TJ, de Rie MA, Dijkmans BA,
2223 Vaishnav AK, Bos JD, Tak PP. Alefacept treatment in psoriatic arthritis: Reduction of the
2224 effector t cell population in peripheral blood and synovial tissue is associated with improvement
2225 of clinical signs of arthritis. *Arthritis Rheum* 2002;46:2776-2784.
- 2226 207 Issekutz AC, Nakazato S, Issekutz TB. Differential roles of v α -4(cd49d/cd29) and I α -
2227 1(cd11a/cd18) integrins and e- and p-selectin during developing and established active or
2228 adoptively transferred adjuvant arthritis in the rat. *Immunol Cell Biol* 2003;81:397-408.
- 2229 208 Badger AM, Blake S, Kapadia R, Sarkar S, Levin J, Swift BA, Hoffman SJ, Stroup GB, Miller
2230 WH, Gowen M, Lark MW. Disease-modifying activity of sb 273005, an orally active, nonpeptide
2231 α v β 3 (vitronectin receptor) antagonist, in rat adjuvant-induced arthritis. *Arthritis Rheum*
2232 2001;44:128-137.
- 2233 209 Ruth JH, Amin MA, Woods JM, He X, Samuel S, Yi N, Haas CS, Koch AE, Bullard DC.
2234 Accelerated development of arthritis in mice lacking endothelial selectins. *Arthritis Res Ther*
2235 2005;7:R959-970.
- 2236 210 Runnels HA, Weber GL, Min J, Kudlacz EM, Zobel JF, Donovan CB, Thiede MA, Zhang J,
2237 Alpert RB, Salafia MA, Milici AJ, Burdette D, Bell RR, Beebe JS, Xu X. Pf-03475952: A potent
2238 and neutralizing fully human anti-cd44 antibody for therapeutic applications in inflammatory
2239 diseases. *Adv Ther* 2010;27:168-180.
- 2240 211 Riechelmann H, Sauter A, Golze W, Hanft G, Schroen C, Hoermann K, Erhardt T, Gronau S.
2241 Phase i trial with the cd44v6-targeting immunoconjugate bivatuzumab mertansine in head and
2242 neck squamous cell carcinoma. *Oral Oncol* 2008;44:823-829.
- 2243 212 Pinto LG, Cunha TM, Vieira SM, Lemos HP, Verri WA, Jr., Cunha FQ, Ferreira SH. Il-17
2244 mediates articular hypernociception in antigen-induced arthritis in mice. *Pain* 2010;148:247-256.
- 2245 213 Park SB, Chun KR, Kim JK, Suk K, Jung YM, Lee WH. The differential effect of high and low
2246 molecular weight fucoidans on the severity of collagen-induced arthritis in mice. *Phytother Res*
2247 2010;24:1384-1391.
- 2248 214 Shu Z, Shi X, Nie D, Guan B. Low-molecular-weight fucoidan inhibits the viability and
2249 invasiveness and triggers apoptosis in il-1 β -treated human rheumatoid arthritis fibroblast
2250 synoviocytes. *Inflammation* 2015;38:1777-1786.
- 2251 215 Verri WA, Jr., Guerrero AT, Fukada SY, Valerio DA, Cunha TM, Xu D, Ferreira SH, Liew FY,
2252 Cunha FQ. Il-33 mediates antigen-induced cutaneous and articular hypernociception in mice.
2253 *Proc Natl Acad Sci U S A* 2008;105:2723-2728.
- 2254 216 Leung BP, Xu D, Culshaw S, McInnes IB, Liew FY. A novel therapy of murine collagen-induced
2255 arthritis with soluble t1/st2. *J Immunol* 2004;173:145-150.
- 2256 217 Palmer G, Talabot-Ayer D, Lamacchia C, Toy D, Seemayer CA, Viatte S, Finckh A, Smith DE,
2257 Gabay C. Inhibition of interleukin-33 signaling attenuates the severity of experimental arthritis.
2258 *Arthritis Rheum* 2009;60:738-749.
- 2259 218 Geusens P, Wouters C, Nijs J, Jiang Y, Dequeker J. Long-term effect of omega-3 fatty acid
2260 supplementation in active rheumatoid arthritis. A 12-month, double-blind, controlled study.
2261 *Arthritis Rheum* 1994;37:824-829.

- 2262 219 Berbert AA, Kondo CR, Almendra CL, Matsuo T, Dichi I. Supplementation of fish oil and olive oil
2263 in patients with rheumatoid arthritis. *Nutrition* 2005;21:131-136.
- 2264 220 Galarraga B, Ho M, Youssef HM, Hill A, McMahon H, Hall C, Ogston S, Nuki G, Belch JJ. Cod
2265 liver oil (n-3 fatty acids) as an non-steroidal anti-inflammatory drug sparing agent in rheumatoid
2266 arthritis. *Rheumatology (Oxford)* 2008;47:665-669.
- 2267 221 Brinkmann V, Zychlinsky A. Beneficial suicide: Why neutrophils die to make nets. *Nat Rev*
2268 *Microbiol* 2007;5:577-582.
- 2269 222 Gokin AP, Fareed MU, Pan HL, Hans G, Strichartz GR, Davar G. Local injection of endothelin-1
2270 produces pain-like behavior and excitation of nociceptors in rats. *J Neurosci* 2001;21:5358-
2271 5366.
- 2272
- 2273
- 2274
- 2275
- 2276
- 2277
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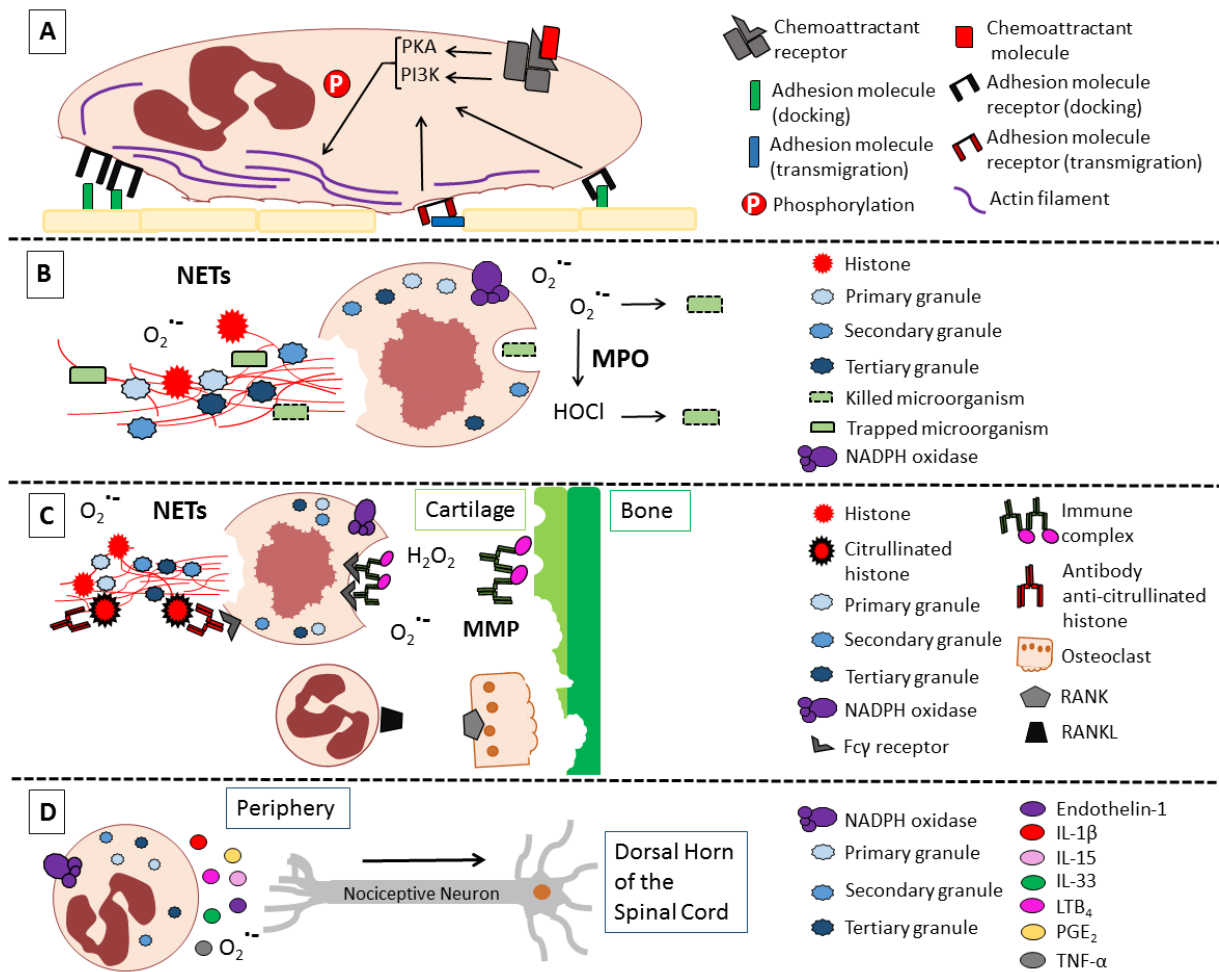
2310 **Figure Legends**
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Figure 1: Schematic view of neutrophil circulation in homeostatic and inflammatory conditions. (A) New formed neutrophils in bone marrow reach blood vessels after the loss of CXCR4 on their surface. Aged circulating neutrophils express back its chemokine receptor that drives their return to bone marrow, where they became apoptotic and phagocytosed by resident macrophages. (B) In inflammatory condition, circulating neutrophils interact with active endothelial cells and reach the site of inflammation through a plethora of chemoattractants. (C) Neutrophils contribute to chronic inflammation and adaptive immunity through the release of new potential autoantigens (proteinase 3 and myeloperoxidase), by guiding dendritic cells and T cells to the site of inflammation or by direct interaction with T cells. (D) On the other hand, efferocytosis of apoptotic neutrophils by resident macrophages contributes to the resolution of inflammation by the release of several pro-resolutive molecules. (E) The reverse transmigration of neutrophil from inflammatory milieu could contribute to the resolution of acute inflammation or could favor the development of inflammation to distant organs. LN: lymph node; G-CSF: granulocyte-colony stimulating factor.

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Figure 2: Neutrophils and arthritis: physiopathological mechanisms in their beneficial and detrimental roles in disease. (A) Schematic representation of the neutrophil recruitment mechanisms. Neutrophils interact with endothelial cells through adhesion molecules and chemokines linked to proteoglycan molecules. This interaction leads to activation of neutrophil intracellular second messengers and activation of actin filaments resulting in transmigration towards the inflammatory foci [28,29,35,129,196]. (B) Schematic representation of the neutrophil microbicidal mechanisms. Upon phagocytosis, the neutrophil preformed killing molecules stored in granules are released into the phagolysosome to encounter the microorganisms. Enzymes such as NADPH oxidase and MPO produce the microbicidal molecules superoxide anion and HOCl, respectively. Neutrophils can also release NETs, which entrap and kill microorganisms [10,11,52,61]. The release of NETs depends on cellular nucleus modifications leading to loss of nucleus integrity [221]. (C) Schematic representation of the neutrophil-dependent tissue lesion mechanisms. Immune complex and pro-inflammatory molecules activate neutrophils that produce ROS and release MMPs responsible for cartilage destruction. Neutrophils can also induce osteoclast differentiation and activation leading to bone erosion. NETs contribute forming citrullinated histones and as an immune consequence, there is the production of antibodies against citrullinated histones that account to the activation of neutrophils and other cells contributing to tissue damage [74,81,85,118,192]. (D) Schematic representation of the neutrophil-dependent pain mechanisms. Recruited neutrophils further produce hyperalgesic molecules capable of activating nociceptive neurons causing pain [51,95-98,222]. MPO: myeloperoxidase; MMP: matrix metalloproteinases; HOCl: hypochlorite; $O_2^{\cdot-}$: superoxide anion.

2360 4 ARTIGO PARA PUBLICAÇÃO (JOURNAL OF NATURAL PRODUCTS)

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2362 O presente trabalho foi realizado no Laboratório de Dor, Inflamação,
2363 Neuropatia e Câncer, da Universidade Estadual de Londrina e segue as normas da
2364 revista Journal of Natural Products. Os resultados parciais estão descritos no artigo
2365 intitulado “Curcumin ameliorates titanium dioxide-induced arthritis by reducing pain,
2366 neutrophils recruitment and proteoglycan degradation”

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2382 **Curcumin ameliorates titanium dioxide-induced arthritis by reducing pain, neutrophils**
2383 **recruitment and proteoglycan degradation**

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ABSTRACT

2406 Arthritic diseases affect millions of people worldwide and reduce substantially the quality of
2407 life. The inflammatory response in the joint induces intense neutrophils recruitment, which
2408 leads to cartilage and bone destructions. A usual outcome of patients with arthritic diseases is
2409 arthroplasty, a procedure known as total or partial replacement of affected articulations. One
2410 of the main biomaterial used to fabricate the prosthesis is titanium dioxide. 10 to 15% of
2411 patients respond against the debris released. In this sense, we aim to investigate the efficacy of
2412 curcumin in a titanium dioxide-induced arthritis. Mice were pretreated with curcumin at 10
2413 and 100 mg/kg 1 h before titanium dioxide injection and after that, daily during 30 days. After
2414 30 days of stimulus, the joint was collected for analysis. By using an electronic version of von
2415 Frey filaments, we investigated the effects of curcumin in pain intensity to a mechanical
2416 stimulus (mechanical threshold) after titanium dioxide injection. We observed that intra-
2417 articular injection of titanium dioxide increased pain and edema. Treatment with curcumin at
2418 100 mg/kg reduced both parameters. Associated to that, curcumin reduced MPO activity, IL-
2419 33 production, and reduced proteoglycan degradation. Therefore, we demonstrated the
2420 efficacy of curcumin in a titanium dioxide-induced arthritis

2421 **Keywords:** arthritis, curcumin, IL-33, implant, pain, titanium dioxide.

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2424 1. Introduction

2425 Acute pain has protective function and acts as warn to the host for maintenance bodily
2426 integrity. Chronic pain, however, is a maladaptive response and represents a substantial and
2427 growing unmet medical need worldwide. The sensitization of nociceptive neurons is a
2428 common denominator of all types of pain leading to hyperalgesia (an increased response to a
2429 stimulus that is normally painful) and/or allodynia (pain due to a stimulus that does not
2430 normally provoke pain)¹. Several mediators contribute to nociceptor sensitization or
2431 activation, such as prostaglandin (PG) E₂, sympathetic amines^{2, 3}, endothelin-1^{4, 5}, superoxide
2432 anion^{6, 7}, and the pro-inflammatory cytokines IL-33⁸, IL-1 β ⁹ and TNF- α ¹⁰. Although
2433 hyperalgesia usually occurs in patients, it does not represent a major clinical problem. On the
2434 other hand, allodynia does, since impairs normal daily activities of the patients and brusquely
2435 reduces the quality of life.

2436 Arthritis is a common denomination to diseases that affect joints. Among these
2437 diseases, highlight rheumatoid arthritis, gout, osteoarthritis and septic arthritis^{11, 12}. A usual
2438 outcome of patients with arthritic diseases is arthroplasty, a procedure known as total or
2439 partial replacement of affected articulations¹³. Patients with implants present extraordinary
2440 benefits and increase of life quality^{14, 15}. Unfortunately, 10 to 15% of patients respond against
2441 the debris released by biomaterials that constitute implants, which ultimately leads to
2442 osteolysis^{16, 17}. One of these biomaterials is titanium dioxide. In a specific case, a patient
2443 without familiar history of arthritic diseases develops implant-related arthritis¹⁸. In fact, after
2444 incubation with titanium dioxide, mononuclear cells derived from patient presented increased
2445 levels of TNF- α production¹⁸. *In vitro* evidence in HepG2 cells suggest that titanium dioxide
2446 activates NF- κ B signaling pathway and has genotoxic effects¹⁹. Corroborating these finding,
2447 injection of titanium dioxide increases neutrophils recruitment and IL-1 β production²⁰. Taking
2448 into account these finding, we have standardized a model of arthritis induced by titanium
2449 dioxide. In this model, intra-articular injection of titanium dioxide induces implant related
2450 arthritis, that leads to pain, edema, increases RANK-RANKL signaling pathway, enhances
2451 pro-inflammatory cytokines production and ultimately leads to joint destruction (Borghi et al.
2452 2016, in preparation)

2453 Although Traditional medicine remains as a good source of therapeutic compounds to
2454 contemporaneous medicine, Western medical establishment views their application with
2455 skepticism. Current treatment of patients with rheumatoid arthritis lies in glucocorticoids,
2456 non-steroidal anti-inflammatory drugs and disease modifying anti-rheumatic drugs.

2457 Monoclonal antibodies and recombinant proteins usually lead to adaptive immunity against
2458 the drugs limiting their use^{21, 22}. In this sense, the use of natural products-derived molecules
2459 represents a promising land to treat chronic pain, by acting as main therapy or at least as
2460 therapeutic adjuvants to reduce the daily doses of conventional drugs that these patients
2461 receive²³⁻²⁵. Curcumin [(E,E)-1,7-bis(4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione]
2462 is a yellow pigment present in the rhizome of the plant *Curcuma longa* considered a
2463 promising compound. One of the greatest benefits of molecules derived from natural sources,
2464 such as curcumin, is that they have been tested “clinically” for thousands of years, since the
2465 traditional medicine practitioners in South Asia use these molecules to treat a variety of
2466 diseases, including arthritis²⁶. Curcumin is a highly pleiotropic molecule with antifungal,
2467 antiviral, antitumor, and antioxidant activities²⁷ with more than 100 different molecular
2468 targets. The potential of curcumin in patients with RA was first reported back in 1980 with
2469 reduction of joint swelling, morning stiffness, and improvement of walking time²⁸.
2470 Corroborating, in a clinical trial with osteoarthritic patients under treatment with Meriva® (a
2471 curcumin-based medicine) reduces reported pain (VAS scale)²⁹. In another clinical trial,
2472 treatment with Meriva® reduces stiffness and physical signs (treadmill test); alongside with
2473 IL-1 β , IL-6, and sVCAM-1 productio³⁰. Recently, a meta-analysis study containing 606
2474 patients with 306 under treatment with curcumin further corroborates these findings and states
2475 that curcumin has potential to be a novel alternative for pain relief³¹. We have previously
2476 demonstrated the analgesic effect of curcumin in acute model of pain induced by superoxide
2477 anion⁷. We observed that treatment with curcumin provides inhibits pain-like behaviors,
2478 reduces pro-inflammatory cytokines production and increases antioxidant capacity. Taking
2479 into account these aforementioned observations, we aim to investigate the potential effect of
2480 curcumin in titanium dioxide-induced arthritis.
2481

2482 2. General experimental procedures

2483 2.1. Animals

2484 Male Swiss mice (25-30 g) from the Universidade Estadual de Londrina, Paraná, Brasil, were
2485 used in this study. Mice were housed in standard clear plastic cages with free access to food
2486 and water and a light/dark cycle of 12:12 h and kept at 21° C. All behavioral tests were
2487 performed between 9 am and 5 pm in a temperature-controlled room. Animal care and
2488 handling procedures followed the International Association for the Study of Pain (IASP)
2489 guidelines. The Ethics Committee of the Universidade Estadual de Londrina approved this
2490 study (Process number 21934.2015.55). All efforts were made to minimize the number of
2491 animals used and their suffering. The experimenters were blinded to the treatments.

2492

2493 2.2. Experimental procedures

2494 Mice were pretreated per oral (p.o.) with 10 or 100 mg/kg of curcumin or with vehicle (20%
2495 Tween 80 in saline) 1 h before single intra-articular (i.a.) injection of 3 mg of titanium dioxide
2496 and after that, mice were treated daily until the end of the experiment. During the specified
2497 time points mechanical threshold and edema were evaluated. 30 days after the stimulus, the
2498 following parameters were determined: myeloperoxidase activity, proteoglycan
2499 quantification, and cytokines production. The dose of titanium dioxide and time points for
2500 sample collection were determined in standardizing experiments in our laboratory (Borghini
2501 2016, in preparation). Titanium dioxide was diluted in 10 µL of sterile saline immediately
2502 before application. Based on the results from mechanical hyperalgesia and edema, the dose of
2503 curcumin of 100 mg/kg was chosen and used in the next experiments.

2504

2505 2.3. Electronic pressure meter test

2506 Electronic pressure meter test was evaluated by an electronic version of von Frey's
2507 filaments as reported previously³². In a quiet room, temperature controlled, mice were placed
2508 in acrylic cages (12 × 10 × 17 cm) with wire grid floors 15-30 min before the start of testing.
2509 The test consisted of evoking a hind paw flexion reflex with a handheld force transducer
2510 (electronic anesthesiometer, IITC Life Science, Woodland Hills, CA) adapted with a 4.15
2511 mm² polypropylene tip. The investigator was trained to apply the tip perpendicularly to the
2512 central area of the plantar hind paw with a gradual increase in pressure. The gradual increase
2513 in pressure was manually performed in blinded experiments. The upper limit pressure was 15
2514 g. The end-point was characterized by the removal of the paw followed by clear flinching

2515 movements. After paw withdrawal, the intensity of the pressure was automatically recorded,
2516 and the final value for the response was obtained by averaging three measurements. The
2517 animals were tested before and after treatments. The flexion-elicited withdrawal threshold is
2518 expressed in grams (g).

2519

2520 2.4 Joint edema

2521 The volume of the joint was measured in mm with a caliper (Mitutoyo, Suzano, SP,
2522 Brazil) before (zero time) the i.a. stimulus with titanium dioxide, and after the administration
2523 of stimulus in the indicated time points. The joint edema is expressed as $\Delta\text{mm}/\text{joint}$.

2524

2525 2.6. Myeloperoxidase (MPO) activity

2526 The neutrophil recruitment and macrophage recruitment to the joint was evaluated by
2527 the MPO activity as previously described⁷. Briefly, mice were terminally anesthetized, and the
2528 joint samples were collected in 400 μL of 50 mM K_2HPO_4 buffer (pH 6.0) containing 0.5%
2529 HTAB and then homogenized in ice-cold Tissue-Tearor (Biospec). After that, homogenates
2530 were centrifuged (16100g, 2 min, 4 °C), and the supernatants were collected. For the MPO
2531 assay, aliquots of 30 μL of supernatant were placed in a 96-well plate and mixed with 200 μL
2532 of 50 mM K_2HPO_4 buffer (pH 6.0), containing 0.0167% ortho-dianisidine dihydrochloride
2533 and 0.05% H_2O_2 . The absorbance was determined after 5 min at 450 nm (Multiskan GO
2534 microplate spectrophotometer, ThermoScientific, Vantaa, Finland). The MPO activity of
2535 samples was compared to a standard curve of neutrophils and presented as MPO activity.

2536

2537 2.7. Proteoglycan quantification assay

2538 Proteoglycan contents were determined as described previously³³. In brief, patella was
2539 carefully collected from each animal and fixed with formaldehyde (4%) overnight using a
2540 shaker. They were then transferred into a solution of formic acid (5%) and incubated for 4 h
2541 using a shaker for decalcification. Each patella was then placed into 100 ml papain digestion
2542 buffer consisting of a papain suspension (5 mg/mL) in calcium and magnesium-free PBS with
2543 5 mM cysteine and 10 mM EDTA, pH 7.4. The samples were sealed and incubated in a
2544 humidified container in a 60°C oven for 16 h. After reaching room temperature, the samples
2545 were centrifuged (1000 g \times 10 min) to collect the condensation droplets. Next, 50 ml of the
2546 supernatants and of serial chondroitin sulfate solutions (standard curve; 50–1000 mg/mL) was
2547 placed into 96-well microtiter plates. The chondroitin sulfate STD solutions were also
2548 incubated with papain digestion buffer. Then, 300 mL of a 1,9-dimethylmethylene blue

2549 (DMMB; 50 mg/L) solution was added to each well, and the absorbance at 525 nm was
2550 measured immediately in a plate reader. The GAG content was calculated from the standard
2551 curve. The DMMB solution was prepared by dissolving 50 mg DMMB in 5 ml ethanol and
2552 diluting to a volume of 1000 ml with 0.2% (w/v) sodium formate buffer, pH 3.5.

2553

2554 2.8. Cytokine measurement

2555 Joint samples were collected 30 days after the injection of titanium dioxide, and
2556 homogenized in 500 μ L of ice-cold buffer containing protease inhibitors, and centrifuged
2557 (3000 rpm \times 10 min \times 4 $^{\circ}$ C), and the supernatants used to measure IL-33 levels by an
2558 enzyme-linked immunosorbent assay (ELISA) using eBioscience kits. As a control, the
2559 concentration of this cytokine was determined in animals injected with saline. The results are
2560 expressed as picograms (pg) of cytokine/100 mg of tissue.

2561

2562 2.9. Statistical analysis

2563 Results are presented as means \pm SEM of measurements made on six mice in each
2564 group per experiment and are representative of two separate experiments. Two-way repeated
2565 measures analysis of variance (ANOVA) followed by Tukey's *post hoc* was used to compare
2566 all groups and doses at all times when responses were measured at different times after the
2567 stimulus injection. Differences between responses were evaluated by one-way ANOVA
2568 followed by Tukey's *post hoc* for data of single time point. Statistical differences were
2569 considered significant when $P < 0.05$.

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2574 3. Results

2575

2576 3.1. Curcumin at 100 mg/kg reduces pain and joint edema

2577 We first sought to determine the dose of curcumin capable of reducing pain. Mice
2578 were pretreated with curcumin (p.o) at 10 and 100 mg/kg 1 h before single intra-articular (i.a.)
2579 injection of 3 mg of titanium dioxide and after that, mice were treated daily until the end of
2580 the experiment. 30 days after stimulus joint was collected for analysis. We observed that
2581 titanium dioxide was able to reduce mechanical threshold (Fig 1A) and increased joint edema
2582 (Fig 1B). On the other hand, treatment with curcumin at 100 mg/kg reduced pain by
2583 increasing mechanical threshold (Fig 1A) and reduced joint edema (Fig 1B) in the evaluated
2584 time points.

2585

2586 3.2. Curcumin at 100 mg/kg reduces proteoglycan degradation

2587 Next, we look for investigating whether curcumin could reduce proteoglycan
2588 degradation. Mice were pretreated with curcumin (p.o) at 10 and 100 mg/kg 1 h before single
2589 intra-articular (i.a.) injection of 3 mg of titanium dioxide and after that, mice were treated
2590 daily until the end of the experiment. 30 days after stimulus joint was collected for analysis.
2591 What we found was that titanium dioxide increased proteoglycan degradation and treatment
2592 with curcumin at 100 mg/kg avoided proteoglycan degradation (Fig 2).

2593

2594 3.3. Curcumin at 100 mg/kg reduces myeloperoxidase activity

2595 The next step was to investigate whether curcumin could reduce neutrophils
2596 recruitment. Mice were pretreated with curcumin (p.o) at 100 mg/kg 1 h before single intra-
2597 articular (i.a.) injection of 3 mg of titanium dioxide and after that, mice were treated daily
2598 until the end of the experiment. 30 days after stimulus joint was collected for analysis. We
2599 observed that titanium dioxide increased neutrophils recruitment estimated by increased
2600 myeloperoxidase activity and treatment with curcumin at 100 mg/kg reduced it (Fig 3).

2601

2602 3.4. Curcumin at 100 mg/kg reduces IL-33 production

2603 To evaluate the effect of curcumin on cytokine production, we sought to determinate
2604 IL-33 production. Mice were pretreated with curcumin (p.o) at 100 mg/kg 1 h before single
2605 intra-articular (i.a.) injection of 3 mg of titanium dioxide and after that, mice were treated
2606 daily until the end of the experiment. 30 days after stimulus joint was collected for analysis.
2607 We observed that titanium dioxide increased the levels of IL-33 and treatment with curcumin
2608 at 100 mg/kg reduced IL-33 production (Fig 4).

2609

2610 4. Discussion

2611

2612 We have previously demonstrate that injection of titanium dioxide into joint induces
2613 arthritis and that treatment with quercetin ameliorates the impairment provided by titanium
2614 dioxide (Borghi 2016, in preparation). Regarding curcumin, we demonstrate that in acute
2615 model of pain, curcumin inhibits superoxide anion induced-pain-like behaviors by reducing
2616 NF- κ B activity and as consequence pro-inflammatory cytokines and increasing Nrf2
2617 expression and as consequence antioxidant defense system and IL-10 production⁷. In the
2618 present work, we further corroborated the analgesic effect of curcumin by demonstrating its
2619 efficacy in a chronic model, such as arthritis. Herein, we demonstrate that curcumin at 100
2620 mg/kg reduced pain, edema, neutrophils recruitment and proteoglycan degradation in titanium
2621 dioxide-induced arthritis.

2622

2623 Chronic pain is a public health problem and has a negative impact on the quality life of
2624 people affected. Arthritic diseases are accompanied with chronic pain which is a consistent
2625 factor to suicidal thoughts and behaviors³⁴ and has a straight relationship with depression³⁵
2626 since occurs alongside with allodynia which impairs normal daily activities. Therefore, seek
2627 for strategies that aim reduce pain should always remain in focus. A usual outcome of patients
2628 with arthritic diseases is arthroplasty, a procedure known as total or partial replacement of
2629 affected articulations¹³. Unfortunately, 10-15% of patients that wear prosthesis present
2630 osteolysis due prosthetics debris release^{16, 17}. Reports of osteolysis date 1970 and have been
2631 associated with all biomaterials, including titanium dioxide^{16, 17}. Generation of giant cells and
2632 “frustrated phagocytosis” in response to microparticles, or endosomal destabilization as a
2633 result of phagocytosis of nanoparticles, are two of the major pathways through which wear
2634 debris activate the NLRP3 inflammasome and lead to the release of the mature form of IL-
2635 1 β ^{16, 20, 36, 37}. Ultimately, this innate immune response leads to activation of RANK-RANKL
2636 signaling pathway in osteoclasts and promotes bone erosion^{16, 37}.

2636

2637 IL-33 is a cytokine from IL-1 family and the only agonist of ST2³⁸. We observed that
2638 titanium dioxide increased IL-33 levels and curcumin reduced it. In the absence of
2639 inflammation, IL-33 possess nuclear localization, which means that can act as an alarmin,
2640 and, therefore, is rapidly released in stressful conditions³⁸. We have demonstrated that
2641 injection of IL-33 promotes mechanical hyperalgesia and increases pro-inflammatory
2642 cytokines such as IL-1 β and TNF- α ⁸. Corroborating, we also demonstrated that IL-33 is
2643 rapidly produced (30 minutes after carrageenan injection) and remains up to 5 hours, in
neutrophils dependent manner, since treatment with fucoidan reduces IL-33 production³⁹.

2644 Importantly, 2 hours after carrageenan injection, mice lacking ST2 presents reduced levels of
2645 TNF- α suggesting that IL-33 production is an upstream event to TNF- α ³⁹ in the cascade of
2646 hyperalgesic cytokines³. Additionally to acute effects, an important point to keep in mind is
2647 that the IL-33/ST2 signaling pathway has a pro-nociceptive role in chronic diseases such as
2648 rheumatoid arthritis⁴⁰⁻⁴² and neuropathy⁴³. In a model of neuropathic pain, we demonstrated
2649 that IL-33 production remains elevated during 21 days and that oligodendrocytes are the major
2650 source of IL-33⁴³. Apart from its hyperalgesic effect *per se*, we demonstrated that neutrophils
2651 from peripheral blood of patients with rheumatoid arthritis treated only with methotrexate
2652 present higher expression of ST2 compared to healthy donors or patients under treatment with
2653 methotrexate plus infliximab. After TNF- α stimulus, neutrophils from healthy donors
2654 increased the expression of ST2. This finding added new evidence in the mechanisms of anti-
2655 TNF therapy by demonstrating that TNF- α primes neutrophils to respond to IL-33
2656 chemotaxis⁴¹. Regarding bone loss, elevated levels of IL-33 reduces bone mineral density,
2657 increases leukocytes recruitment, and increases the level of pro-inflammatory cytokines such
2658 as IL-6 and INF- γ . This ultimately leads to bone erosion⁴⁴. Corroborating, IL-33 increases the
2659 expression of RANKL in osteoclasts⁴⁵. Importantly, mice lacking IL-33 did not change body
2660 weight and biomechanical strength⁴⁴. In this sense, inhibition of IL-33/ST2 signaling is a very
2661 promisor therapy since interacts with multiple pathways. To our knowledge, this is the first
2662 evidence of curcumin inhibiting IL-33 production. Curcumin inhibits IKK activity, and
2663 therefore avoids p65-p50 NF- κ B heterodimer activation and consequent translocation to the
2664 nucleus²⁷. This is a possible explanation by which curcumin inhibited IL-33 levels herein
2665 observed.

2666 Neutrophils play an important role in several chronic diseases, especially in arthritic
2667 diseases, which they are the major cells in the inflammatory infiltrate²². Herein, injection
2668 titanium dioxide enhanced MPO activity and curcumin was able to reduce. Neutrophils' vast
2669 microbicidal array contributes to the destruction of cartilage and bone directly²² or indirectly
2670 by activation osteoclasts through RANK-RANKL signaling pathway⁴⁶. The discovery that
2671 neutrophils release NETs added new weapons in this vast arsenal⁴⁷ and opened new avenues
2672 for the treatment of these diseases. The release of NETs triggers a robust autoimmune
2673 response and collaborates to the genesis of diseases such as systemic lupus erythematosus,
2674 rheumatoid arthritis, and gout²². In rheumatoid arthritis, NETosis seems the primary source of
2675 citrullinated antigens⁴⁸ which are generated by peptidyl arginine deiminase 4⁴⁹ and contributes
2676 to aggravate the disease^{50, 51}. This mechanism seems important in implant-related arthritis
2677 since titanium dioxide triggers histone citrullination and NETs release⁵². Apart from NETs,

2678 neutrophils are the major source of pro-inflammatory cytokines such as IL-33, IL-1 β , and
2679 TNF- α in arthritic diseases^{22, 41}. These cytokines have central role in the pathophysiology of
2680 arthritic diseases and pain states. Another striking feature is that titanium dioxide inhibits
2681 neutrophils apoptosis and increases IL-8 and IL-6 production⁵³. Associated to that, titanium
2682 dioxide enhances neutrophils phagocytosis in Syk (spleen tyrosine kinase)-dependent
2683 manner⁵⁴ and as consequence activates NADPH oxidase which leads to augmentation of ROS
2684 production⁵⁵. These mechanisms provide deleterious consequences in the inflammatory milieu
2685 and perpetuate a loop that leads to joint destruction, since NADPH oxidase, ROS⁵⁶, and
2686 MPO⁵⁷ are required for neutrophils undergo NETosis. Curcumin inhibits neutrophils actin
2687 polymerization⁵⁸ and reduces adhesion molecules expression⁵⁹, which results in inhibition of
2688 recruitment of these cells into inflammatory foci herein observed. Therefore, inhibition of
2689 neutrophils recruitment is a crucial step to reduce cartilage and bone destructions and –
2690 alongside with NF- κ B inhibition – may explain why we observed a reduction in IL-33
2691 production aligned with a reduction in proteoglycan degradation.

2692 Curcumin is a highly pleiotropic molecule and its mechanism of action of curcumin
2693 depends on a synergism involving multiples signaling pathways e cells. In the present work,
2694 we demonstrated the analgesic effect of curcumin in implant-related arthritis model.
2695 Therefore, treatment with curcumin at 100 mg/kg reduced neutrophils recruitment into
2696 inflammatory foci. As a consequence of that, we observed reduced levels of IL-33,
2697 proteoglycan degradation, and finally reduced pain and edema induced by titanium dioxide
2698 injection.

2699

2700 **Conflict of interest**

2701 The authors declare no conflict of interest.

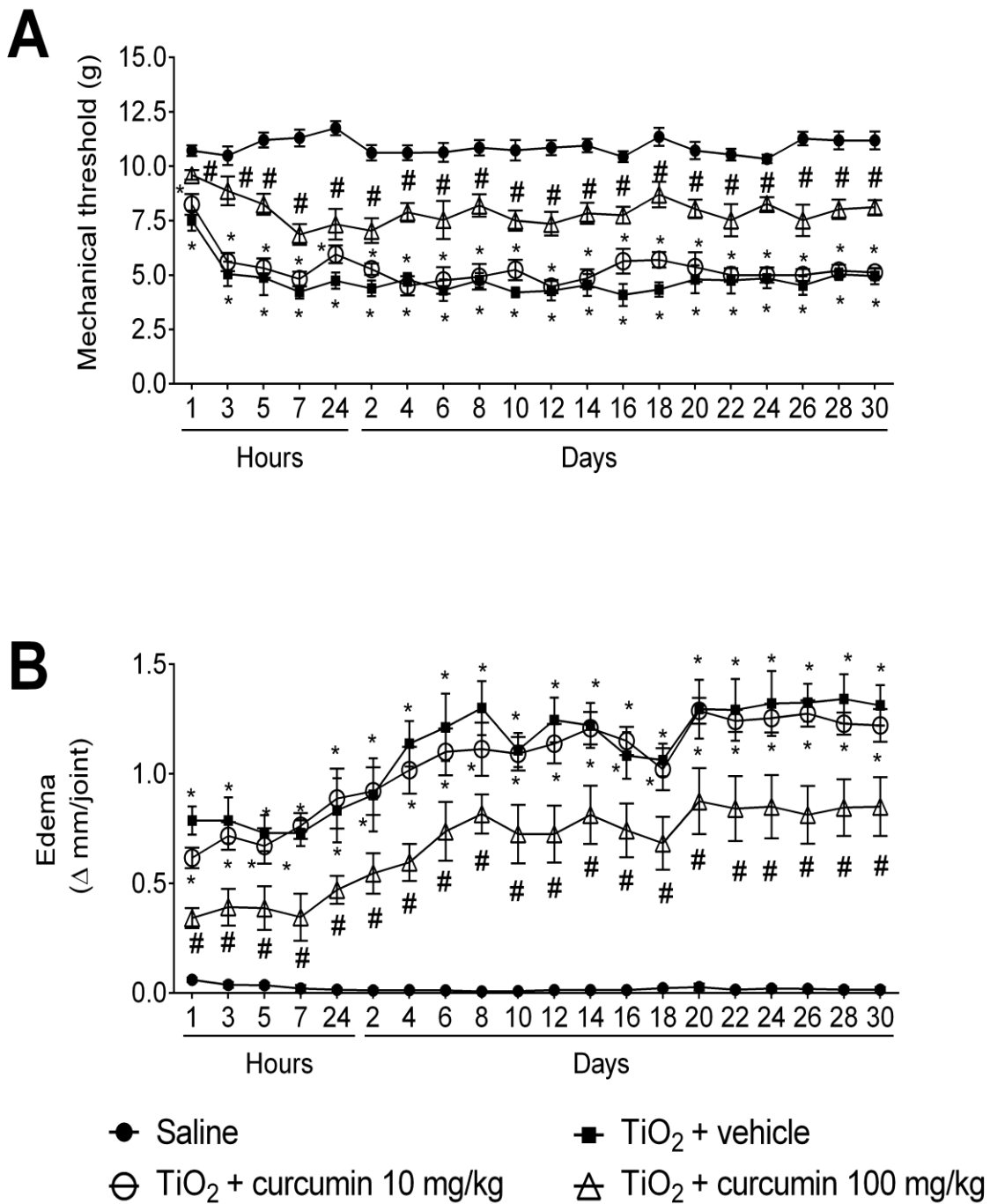
2702 **Acknowledgments**

2703 Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), São Paulo
2704 Research Foundation (FAPESP) under grant agreements number 2011/19670-0 (Thematic
2705 project) and 2013/08216-2 (Center for Research in Inflammatory Disease-CRID),
2706 Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Ministério da
2707 Ciência Tecnologia e Inovação (MCTI), Secretaria da Ciência, Tecnologia e Ensino Superior
2708 (SETI), Fundação Araucária and Parana State Government grants supported this study
2709 (Brazil).

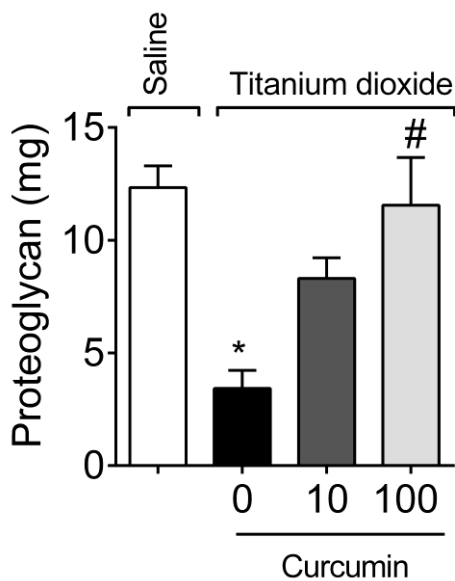
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2712 Figure captions
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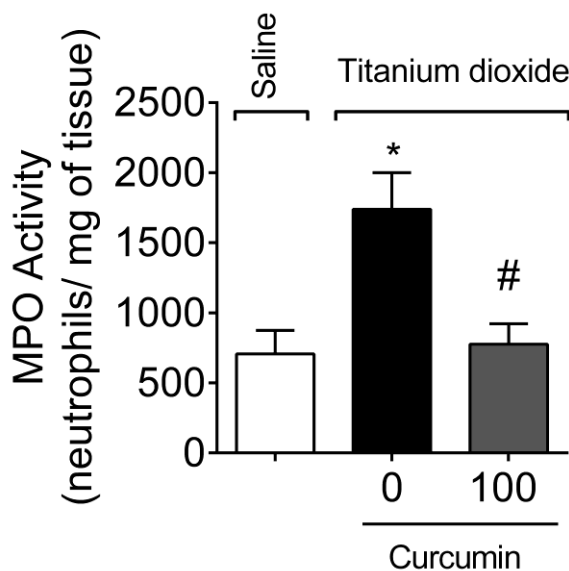
2714 Figure 1. Curcumin at 100 mg/kg reduces pain and joint edema.
2715 Mice were pretreated with curcumin (p.o) at 10 and 100 mg/kg 1 h before single intra-
2716 articular (i.a.) injection of 3 mg of titanium dioxide and after that, mice were treated daily
2717 until the end of the experiment. Mechanical threshold (A) and edema (B) were measured at
2718 the specified time points. Results are expressed as mean \pm SEM (n = 6 per group per
2719 experiment, representative of two separate experiments). *p<0.05 vs. saline group #p<0.05 vs.
2720 titanium dioxide group (two-way repeated measures ANOVA followed Tukey's *post hoc*)
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Figure 2. Curcumin at 100 mg/kg reduces proteoglycan degradation

Mice were pretreated with curcumin (p.o) at 10 and 100 mg/kg 1 h before single intra-articular (i.a.) injection of 3 mg of titanium dioxide and after that, mice were treated daily until the end of the experiment. 30 days after stimulus joint was collected for proteoglycan quantification. Results are expressed as mean \pm SEM (n = 6 per group per experiment, representative of two separate experiments). *p<0.05 vs. saline group #p<0.05 vs. titanium dioxide group (one-way ANOVA followed Tukey's *post hoc*)

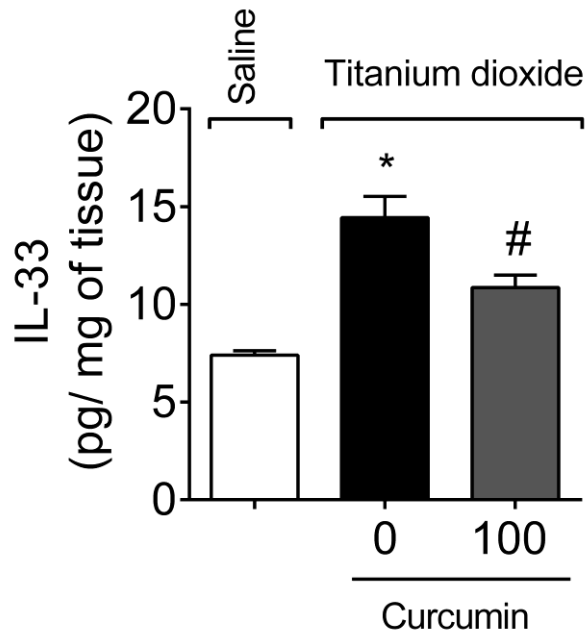


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Figure 3. Curcumin at 100 mg/kg reduces myeloperoxidase activity

Mice were pretreated with curcumin (p.o) 100 mg/kg 1 h before single intra-articular (i.a.) injection of 3 mg of titanium dioxide and after that, mice were treated daily until the end of the experiment. 30 days after stimulus joint was collected for MPO activity. Results are expressed as mean \pm SEM (n = 6 per group per experiment, representative of two separate experiments). *p<0.05 vs. saline group #p<0.05 vs. titanium dioxide group (one-way ANOVA followed Tukey's *post hoc*)

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Figure 4. Curcumin at 100 mg/kg reduces IL-33 production
Mice were pretreated with curcumin (p.o) at 100 mg/kg 1 h before single intra-articular (i.a.) injection of 3 mg of titanium dioxide and after that, mice were treated daily until the end of the experiment. 30 days after stimulus joint was collected for dosage of IL-33 by ELISA. Results are expressed as mean \pm SEM (n = 6 per group per experiment, representative of two separate experiments). *p<0.05 vs. saline group #p<0.05 vs. titanium dioxide group (one-way ANOVA followed Tukey's *post hoc*)

2763 **References**

- 2764 1. Verri, W. A., Jr.; Cunha, T. M.; Parada, C. A.; Poole, S.; Cunha, F. Q.; Ferreira, S. H.,
2765 Hypernociceptive role of cytokines and chemokines: targets for analgesic drug development?
2766 *Pharmacol Ther* **2006**, *112*, 116-38.
- 2767 2. Nakamura, M.; Ferreira, S. H., A peripheral sympathetic component in inflammatory hyperalgesia.
2768 *Eur J Pharmacol* **1987**, *135*, 145-53.
- 2769 3. Cunha, T. M.; Verri, W. A., Jr.; Silva, J. S.; Poole, S.; Cunha, F. Q.; Ferreira, S. H., A cascade of
2770 cytokines mediates mechanical inflammatory hypernociception in mice. *Proc Natl Acad Sci U S A*
2771 **2005**, *102*, 1755-60.
- 2772 4. Donate, P. B.; Cunha, T. M.; Verri, W. A., Jr.; Junta, C. M.; Lima, F. O.; Vieira, S. M.; Peres, R.
2773 S.; Bombonato-Prado, K. F.; Louzada, P., Jr.; Ferreira, S. H.; Donadi, E. A.; Passos, G. A.;
2774 Cunha, F. Q., Bosentan, an endothelin receptor antagonist, ameliorates collagen-induced
2775 arthritis: the role of TNF-alpha in the induction of endothelin system genes. *Inflamm Res* **2012**,
2776 *61*, 337-48.
- 2777 5. Gokin, A. P.; Fareed, M. U.; Pan, H. L.; Hans, G.; Strichartz, G. R.; Davar, G., Local injection of
2778 endothelin-1 produces pain-like behavior and excitation of nociceptors in rats. *J Neurosci* **2001**,
2779 *21*, 5358-66.
- 2780 6. Wang, Z. Q.; Porreca, F.; Cuzzocrea, S.; Galen, K.; Lightfoot, R.; Masini, E.; Muscoli, C.; Mollace,
2781 V.; Ndengele, M.; Ischiropoulos, H.; Salvemini, D., A newly identified role for superoxide in
2782 inflammatory pain. *J Pharmacol Exp Ther* **2004**, *309*, 869-78.
- 2783 7. Fattori, V.; Pinho-Ribeiro, F. A.; Borghi, S. M.; Alves-Filho, J. C.; Cunha, T. M.; Cunha, F. Q.;
2784 Casagrande, R.; Verri, W. A., Jr., Curcumin inhibits superoxide anion-induced pain-like behavior
2785 and leukocyte recruitment by increasing Nrf2 expression and reducing NF-kappaB activation.
2786 *Inflamm Res* **2015**, *64*, 993-1003.
- 2787 8. Verri, W. A., Jr.; Guerrero, A. T.; Fukada, S. Y.; Valerio, D. A.; Cunha, T. M.; Xu, D.; Ferreira, S.
2788 H.; Liew, F. Y.; Cunha, F. Q., IL-33 mediates antigen-induced cutaneous and articular
2789 hypernociception in mice. *Proc Natl Acad Sci U S A* **2008**, *105*, 2723-8.
- 2790 9. Binshok, A. M.; Wang, H.; Zimmermann, K.; Amaya, F.; Vardeh, D.; Shi, L.; Brenner, G. J.; Ji, R.
2791 R.; Bean, B. P.; Woolf, C. J.; Samad, T. A., Nociceptors are interleukin-1beta sensors. *J Neurosci*
2792 **2008**, *28*, 14062-73.
- 2793 10. Jin, X.; Gereau, R. W. t., Acute p38-mediated modulation of tetrodotoxin-resistant sodium
2794 channels in mouse sensory neurons by tumor necrosis factor-alpha. *J Neurosci* **2006**, *26*, 246-55.
- 2795 11. Smolen, J. S.; Aletaha, D.; Redlich, K., The pathogenesis of rheumatoid arthritis: new insights
2796 from old clinical data? *Nat Rev Rheumatol* **2012**, *8*, 235-43.
- 2797 12. Martin, W. J.; Harper, J. L., Innate inflammation and resolution in acute gout. *Immunol Cell Biol*
2798 **2010**, *88*, 15-9.
- 2799 13. Lee, K.; Goodman, S. B., Current state and future of joint replacements in the hip and knee.
2800 *Expert Rev Med Devices* **2008**, *5*, 383-93.
- 2801 14. da Silva, R. R.; Santos, A. A.; de Sampaio Carvalho Junior, J.; Matos, M. A., Quality of life after
2802 total knee arthroplasty: systematic review. *Rev Bras Ortop* **2014**, *49*, 520-7.
- 2803 15. Mariconda, M.; Galasso, O.; Costa, G. G.; Recano, P.; Cerbasi, S., Quality of life and functionality
2804 after total hip arthroplasty: a long-term follow-up study. *BMC Musculoskelet Disord* **2011**, *12*, 222.

- 2805 16. Goodman, S. B., Wear particles, periprosthetic osteolysis and the immune system. *Biomaterials*
2806 **2007**, *28*, 5044-8.
- 2807 17. Harris, W. H., Wear and periprosthetic osteolysis: the problem. *Clin Orthop Relat Res* **2001**, 66-
2808 70.
- 2809 18. Dorner, T.; Haas, J.; Loddenkemper, C.; von Baehr, V.; Salama, A., Implant-related inflammatory
2810 arthritis. *Nat Clin Pract Rheumatol* **2006**, *2*, 53-6; quiz 57.
- 2811 19. Prasad, R. Y.; Simmons, S. O.; Killius, M. G.; Zucker, R. M.; Kligerman, A. D.; Blackman, C. F.;
2812 Fry, R. C.; Demarini, D. M., Cellular interactions and biological responses to titanium dioxide
2813 nanoparticles in HepG2 and BEAS-2B cells: role of cell culture media. *Environ Mol Mutagen*
2814 **2014**, *55*, 336-42.
- 2815 20. St Pierre, C. A.; Chan, M.; Iwakura, Y.; Ayers, D. C.; Kurt-Jones, E. A.; Finberg, R. W.,
2816 Periprosthetic osteolysis: characterizing the innate immune response to titanium wear-particles. *J*
2817 *Orthop Res* **2010**, *28*, 1418-24.
- 2818 21. McInnes, I. B.; Liew, F. Y., Cytokine networks--towards new therapies for rheumatoid arthritis. *Nat*
2819 *Clin Pract Rheumatol* **2005**, *1*, 31-9.
- 2820 22. Fattori, V.; Amaral, F. A.; Verri, W. A., Jr., Neutrophils and arthritis: Role in disease and
2821 pharmacological perspectives. *Pharmacol Res* **2016**.
- 2822 23. Natarajan, V.; Madhan, B.; Tiku, M. L., Intra-Articular Injections of Polyphenols Protect Articular
2823 Cartilage from Inflammation-Induced Degradation: Suggesting a Potential Role in Cartilage
2824 Therapeutics. *PLoS One* **2015**, *10*, e0127165.
- 2825 24. Gelderman, K. A.; Hultqvist, M.; Olsson, L. M.; Bauer, K.; Pizzolla, A.; Olofsson, P.; Holmdahl, R.,
2826 Rheumatoid arthritis: the role of reactive oxygen species in disease development and therapeutic
2827 strategies. *Antioxid Redox Signal* **2007**, *9*, 1541-67.
- 2828 25. Khanna, D.; Sethi, G.; Ahn, K. S.; Pandey, M. K.; Kunnumakkara, A. B.; Sung, B.; Aggarwal, A.;
2829 Aggarwal, B. B., Natural products as a gold mine for arthritis treatment. *Curr Opin Pharmacol*
2830 **2007**, *7*, 344-51.
- 2831 26. Aggarwal, B. B.; Sung, B., Pharmacological basis for the role of curcumin in chronic diseases: an
2832 age-old spice with modern targets. *Trends Pharmacol Sci* **2009**, *30*, 85-94.
- 2833 27. Zhou, H.; Beevers, C. S.; Huang, S., The targets of curcumin. *Curr Drug Targets* **2011**, *12*, 332-
2834 47.
- 2835 28. Deodhar, S. D.; Sethi, R.; Srimal, R. C., Preliminary study on antirheumatic activity of curcumin
2836 (diferuloyl methane). *Indian J Med Res* **1980**, *71*, 632-4.
- 2837 29. Di Pierro, F.; Rapacioli, G.; Di Maio, E. A.; Appendino, G.; Franceschi, F.; Togni, S., Comparative
2838 evaluation of the pain-relieving properties of a lecithinized formulation of curcumin (Meriva((R))),
2839 nimesulide, and acetaminophen. *J Pain Res* **2013**, *6*, 201-5.
- 2840 30. Belcaro, G.; Cesarone, M. R.; Dugall, M.; Pellegrini, L.; Ledda, A.; Grossi, M. G.; Togni, S.;
2841 Appendino, G., Efficacy and safety of Meriva(R), a curcumin-phosphatidylcholine complex, during
2842 extended administration in osteoarthritis patients. *Altern Med Rev* **2010**, *15*, 337-44.
- 2843 31. Sahebkar, A.; Henrotin, Y., Analgesic Efficacy and Safety of Curcuminoids in Clinical Practice: A
2844 Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Pain Med* **2015**.
- 2845 32. Guerrero, A. T.; Verri, W. A., Jr.; Cunha, T. M.; Silva, T. A.; Schivo, I. R.; Dal-Secco, D.; Canetti,
2846 C.; Rocha, F. A.; Parada, C. A.; Cunha, F. Q.; Ferreira, S. H., Involvement of LTB4 in zymosan-
2847 induced joint nociception in mice: participation of neutrophils and PGE2. *J Leukoc Biol* **2008**, *83*,
2848 122-30.

- 2849 33. Vieira, S. M.; Cunha, T. M.; Franca, R. F.; Pinto, L. G.; Talbot, J.; Turato, W. M.; Lemos, H. P.;
2850 Lima, J. B.; Verri, W. A., Jr.; Almeida, S. C.; Ferreira, S. H.; Louzada-Junior, P.; Zamboni, D. S.;
2851 Cunha, F. Q., Joint NOD2/RIPK2 signaling regulates IL-17 axis and contributes to the
2852 development of experimental arthritis. *J Immunol* **2012**, *188*, 5116-22.
- 2853 34. Calati, R.; Laglaoui Bakhiyi, C.; Artero, S.; Ilgen, M.; Courtet, P., The impact of physical pain on
2854 suicidal thoughts and behaviors: Meta-analyses. *J Psychiatr Res* **2015**, *71*, 16-32.
- 2855 35. Fishbain, D. A.; Cutler, R.; Rosomoff, H. L.; Rosomoff, R. S., Chronic pain-associated depression:
2856 antecedent or consequence of chronic pain? A review. *Clin J Pain* **1997**, *13*, 116-37.
- 2857 36. Looney, R. J.; Schwarz, E. M.; Boyd, A.; O'Keefe, R. J., Periprosthetic osteolysis: an
2858 immunologist's update. *Curr Opin Rheumatol* **2006**, *18*, 80-7.
- 2859 37. O'Neill, L. A., Immunology. How frustration leads to inflammation. *Science* **2008**, *320*, 619-20.
- 2860 38. Arshad, M. I.; Piquet-Pellorce, C.; Samson, M., IL-33 and HMGB1 alarmins: sensors of cellular
2861 death and their involvement in liver pathology. *Liver Int* **2012**, *32*, 1200-10.
- 2862 39. Zarpelon, A. C.; Cunha, T. M.; Alves-Filho, J. C.; Pinto, L. G.; Ferreira, S. H.; McInnes, I. B.; Xu,
2863 D.; Liew, F. Y.; Cunha, F. Q.; Verri, W. A., Jr., IL-33/ST2 signalling contributes to carrageenin-
2864 induced innate inflammation and inflammatory pain: role of cytokines, endothelin-1 and
2865 prostaglandin E2. *Br J Pharmacol* **2013**, *169*, 90-101.
- 2866 40. Tang, S.; Huang, H.; Hu, F.; Zhou, W.; Guo, J.; Jiang, H.; Mu, R.; Li, Z., Increased IL-33 in
2867 synovial fluid and paired serum is associated with disease activity and autoantibodies in
2868 rheumatoid arthritis. *Clin Dev Immunol* **2013**, *2013*, 985301.
- 2869 41. Verri, W. A., Jr.; Souto, F. O.; Vieira, S. M.; Almeida, S. C.; Fukada, S. Y.; Xu, D.; Alves-Filho, J.
2870 C.; Cunha, T. M.; Guerrero, A. T.; Mattos-Guimaraes, R. B.; Oliveira, F. R.; Teixeira, M. M.; Silva,
2871 J. S.; McInnes, I. B.; Ferreira, S. H.; Louzada-Junior, P.; Liew, F. Y.; Cunha, F. Q., IL-33 induces
2872 neutrophil migration in rheumatoid arthritis and is a target of anti-TNF therapy. *Ann Rheum Dis*
2873 **2010**, *69*, 1697-703.
- 2874 42. Hong, Y. S.; Moon, S. J.; Joo, Y. B.; Jeon, C. H.; Cho, M. L.; Ju, J. H.; Oh, H. J.; Heo, Y. J.; Park,
2875 S. H.; Kim, H. Y.; Min, J. K., Measurement of interleukin-33 (IL-33) and IL-33 receptors (sST2 and
2876 ST2L) in patients with rheumatoid arthritis. *J Korean Med Sci* **2011**, *26*, 1132-9.
- 2877 43. Zarpelon, A. C.; Rodrigues, F. C.; Lopes, A. H.; Souza, G. R.; Carvalho, T. T.; Pinto, L. G.; Xu,
2878 D.; Ferreira, S. H.; Alves-Filho, J. C.; McInnes, I. B.; Ryffel, B.; Quesniaux, V. F.; Reverchon, F.;
2879 Mortaud, S.; Menuet, A.; Liew, F. Y.; Cunha, F. Q.; Cunha, T. M.; Verri, W. A., Jr., Spinal cord
2880 oligodendrocyte-derived alarmin IL-33 mediates neuropathic pain. *FASEB J* **2016**, *30*, 54-65.
- 2881 44. Okragly, A. J.; Hamang, M. J.; Pena, E. A.; Baker, H. E.; Bullock, H. A.; Lucchesi, J.; Martin, A.
2882 P.; Ma, Y. L.; Benschop, R. J., Elevated levels of Interleukin (IL)-33 induce bone pathology but
2883 absence of IL-33 does not negatively impact normal bone homeostasis. *Cytokine* **2016**, *79*, 66-
2884 73.
- 2885 45. Mine, Y.; Makihira, S.; Yamaguchi, Y.; Tanaka, H.; Nikawa, H., Involvement of ERK and p38
2886 MAPK pathways on Interleukin-33-induced RANKL expression in osteoblastic cells. *Cell Biol Int*
2887 **2014**, *38*, 655-62.
- 2888 46. Chakravarti, A.; Raquil, M. A.; Tessier, P.; Poubelle, P. E., Surface RANKL of Toll-like receptor 4-
2889 stimulated human neutrophils activates osteoclastic bone resorption. *Blood* **2009**, *114*, 1633-44.
- 2890 47. Brinkmann, V.; Reichard, U.; Goosmann, C.; Fauler, B.; Uhlemann, Y.; Weiss, D. S.; Weinrauch,
2891 Y.; Zychlinsky, A., Neutrophil extracellular traps kill bacteria. *Science* **2004**, *303*, 1532-5.

- 2893 48. Khandpur, R.; Carmona-Rivera, C.; Vivekanandan-Giri, A.; Gizinski, A.; Yalavarthi, S.; Knight, J.
2894 S.; Friday, S.; Li, S.; Patel, R. M.; Subramanian, V.; Thompson, P.; Chen, P.; Fox, D. A.;
2895 Pennathur, S.; Kaplan, M. J., NETs are a source of citrullinated autoantigens and stimulate
2896 inflammatory responses in rheumatoid arthritis. *Sci Transl Med* **2013**, *5*, 178ra40.
- 2897 49. Spengler, J.; Lugonja, B.; Ytterberg, A. J.; Zubarev, R. A.; Creese, A. J.; Pearson, M. J.; Grant,
2898 M. M.; Milward, M.; Lundberg, K.; Buckley, C. D.; Filer, A.; Raza, K.; Cooper, P. R.; Chapple, I. L.;
2899 Scheel-Toellner, D., Release of active peptidyl arginine deiminases by neutrophils can explain
2900 production of extracellular citrullinated autoantigens in RA synovial fluid. *Arthritis Rheumatol*
2901 **2015**.
- 2902 50. Romero, V.; Fert-Bober, J.; Nigrovic, P. A.; Darrah, E.; Haque, U. J.; Lee, D. M.; van Eyk, J.;
2903 Rosen, A.; Andrade, F., Immune-mediated pore-forming pathways induce cellular
2904 hypercitrullination and generate citrullinated autoantigens in rheumatoid arthritis. *Sci Transl Med*
2905 **2013**, *5*, 209ra150.
- 2906 51. Scally, S. W.; Petersen, J.; Law, S. C.; Dudek, N. L.; Nel, H. J.; Loh, K. L.; Wijeyewickrema, L. C.;
2907 Eckle, S. B.; van Heemst, J.; Pike, R. N.; McCluskey, J.; Toes, R. E.; La Gruta, N. L.; Purcell, A.
2908 W.; Reid, H. H.; Thomas, R.; Rossjohn, J., A molecular basis for the association of the HLA-
2909 DRB1 locus, citrullination, and rheumatoid arthritis. *J Exp Med* **2013**, *210*, 2569-82.
- 2910 52. Vitkov, L.; Krautgartner, W. D.; Obermayer, A.; Stoiber, W.; Hannig, M.; Klappacher, M.; Hartl, D.,
2911 The initial inflammatory response to bioactive implants is characterized by NETosis. *PLoS One*
2912 **2015**, *10*, e0121359.
- 2913 53. Goncalves, D. M.; Chiasson, S.; Girard, D., Activation of human neutrophils by titanium dioxide
2914 (TiO₂) nanoparticles. *Toxicol In Vitro* **2010**, *24*, 1002-8.
- 2915 54. Babin, K.; Goncalves, D. M.; Girard, D., Nanoparticles enhance the ability of human neutrophils to
2916 exert phagocytosis by a Syk-dependent mechanism. *Biochim Biophys Acta* **2015**, *1850*, 2276-82.
- 2917 55. Masoud, R.; Bizouarn, T.; Trepout, S.; Wien, F.; Baciou, L.; Marco, S.; Houee Levin, C., Titanium
2918 Dioxide Nanoparticles Increase Superoxide Anion Production by Acting on NADPH Oxidase.
2919 *PLoS One* **2015**, *10*, e0144829.
- 2920 56. Rohm, M.; Grimm, M. J.; D'Auria, A. C.; Almyroudou, N. G.; Segal, B. H.; Urban, C. F., NADPH
2921 oxidase promotes neutrophil extracellular trap formation in pulmonary aspergillosis. *Infect Immun*
2922 **2014**, *82*, 1766-77.
- 2923 57. Metzler, K. D.; Fuchs, T. A.; Nauseef, W. M.; Reumaux, D.; Roesler, J.; Schulze, I.; Wahn, V.;
2924 Papayannopoulos, V.; Zychlinsky, A., Myeloperoxidase is required for neutrophil extracellular trap
2925 formation: implications for innate immunity. *Blood* **2011**, *117*, 953-9.
- 2926 58. Larmonier, C. B.; Midura-Kiela, M. T.; Ramalingam, R.; Laubitz, D.; Janikashvili, N.; Larmonier,
2927 N.; Ghishan, F. K.; Kiela, P. R., Modulation of neutrophil motility by curcumin: implications for
2928 inflammatory bowel disease. *Inflamm Bowel Dis* **2011**, *17*, 503-15.
- 2929 59. Kim, D. C.; Lee, W.; Bae, J. S., Vascular anti-inflammatory effects of curcumin on HMGB1-
2930 mediated responses in vitro. *Inflamm Res* **2011**, *60*, 1161-8.
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2936 5 ARTIGO 2 PUBLICADO (INFLAMMATION RESEARCH)

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2938 O presente trabalho foi realizado no Laboratório de Dor, Inflamação,
2939 Neuropatia e Câncer, da Universidade Estadual de Londrina e publicado na revista
2940 Inflammation Research. Os resultados estão descritos no artigo intitulado “Curcumin
2941 inhibits superoxide anion-induced pain-like behavior and leukocyte recruitment by
2942 increasing Nrf2 expression and reducing NF-κB activation” (doi: 10.1007/s00011-
2943 015-0885-y).

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2962 **Curcumin inhibits superoxide anion-induced pain-like behavior and leukocyte**
2963 **recruitment by increasing Nrf2 expression and reducing NF- κ B activation**

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2985 **ABSTRACT**

2986 **Objective:** Evaluated the activity of curcumin in superoxide anion-induced pain-like behavior
2987 and leukocyte recruitment in mice.

2988 **Treatment:** Curcumin 10 mg/kg subcutaneously 1h before stimulus.

2989 **Methods:** KO₂ was used as superoxide anion donor. Overt pain-like behaviors were
2990 determined by the number of abdominal writhings, paw flinches and time spent licking the
2991 paw. Mechanical and thermal hyperalgesia were determined using an electronic
2992 anesthesiometer and hot plate, respectively. Cytokine concentration and NF-κB activity were
2993 determined by ELISA, antioxidant effect by nitrobluetretrazolium assay and ABTS radical
2994 scavenging ability. Myeloperoxidase activity was measured by colorimetric assay. The Nrf2,
2995 heme oxygenase-1 (HO-1) and gp91^{phox} mRNA expression was determined by quantitative
2996 PCR. Data were analysed by ANOVA followed by Tukey's *post hoc* and considered
2997 significant when P<0.05

2998 **Results:** Curcumin inhibited superoxide anion-induced overt pain-like behaviors as well as
2999 mechanical and thermal hyperalgesia. Curcumin also inhibited superoxide anion-induced
3000 leukocyte recruitment in the peritoneal cavity and in the paw skin inhibited myeloperoxidase
3001 activity, oxidative stress, IL-1β and TNF-α production and NF-κB activation as well as
3002 enhanced IL-10 production, and HO-1 and Nrf2 mRNA expression.

3003 **Conclusion:** Curcumin inhibits superoxide anion-induced inflammatory pain-like behaviors
3004 and leukocyte recruitment by targeting inflammatory molecules and oxidative stress; and
3005 inducing antioxidant and anti-inflammatory pathways.

3006 **Keywords:** curcumin, IL-10, NF-κB, Nrf2, pain, superoxide anion.

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3009 1. Introduction

3010

3011 Pain is a symptom present in several diseases affecting nearly one in every five
3012 patients and is the main cause of search for medical attendance. Therefore, it is considered an
3013 international health problem [1]. Nociceptive neuron sensitization is a process that facilitates
3014 depolarization. The sensitization of nociceptive neurons is a common denominator of all types
3015 of inflammatory pain leading to hyperalgesia (an increased response to a stimulus that is
3016 normally painful) and/or allodynia (pain due to a stimulus that does not normally provoke
3017 pain) [2]. Several mediators contribute to nociceptor sensitization or activation through
3018 membrane receptors, such as prostaglandin (PG) E₂, sympathetic amines, endothelin-1, and
3019 the pro-inflammatory cytokines IL-1 β and TNF- α [3-5]. A cascade of pro-hyperalgesic
3020 cytokines precedes the release of prostaglandins and sympathetic amines [2]. In mice, this
3021 cascade initiates with the release of the alarmin IL-33 [6] that induces the production of TNF-
3022 α and keratinocyte-derived chemokine (KC/CXCL1). TNF- α and KC/CXCL1 trigger the
3023 production IL-1 β that in turn stimulates the production of PGE₂ [5]. These cytokines further
3024 contribute to inflammatory hyperalgesia by promoting neutrophil recruitment toward the
3025 inflammatory site and inducing the production of PGE₂ and reactive oxygen species by
3026 migratory and tissue resident cells. Altogether culminating in sensitization of primary
3027 nociceptive neurons [7].

3028 In addition to a hierarchy of cytokine release, there is also a temporal profile of
3029 inflammatory events. The inflammatory hyperalgesia depends on the sensitization of the
3030 nociceptors by hyperalgesic mediators. This nociceptor sensitization is not an immediate
3031 event; therefore, the peak of hyperalgesic mediator production occurs before the peak of
3032 hyperalgesia [2]. Similarly, neutrophil recruitment peaks approximately 6 h after cytokine
3033 injection. In fact, the peak of chemotactic cytokine production occurs before maximal
3034 neutrophil recruitment [8].

3035 Reactive oxygen species (ROS) also contribute to inflammatory hyperalgesia,
3036 especially superoxide anion radical (O₂^{•-}) [9, 10]. For instance, the anti-nociceptive effect of
3037 M40403 (SOD mimetic) depends on diminishing O₂^{•-} effects peripherally and centrally.
3038 Peripheral mechanisms triggered by O₂^{•-} are related to the release of pro-hyperalgesic
3039 cytokines, and central mechanisms are dependent on the reaction of O₂^{•-} with nitric oxide
3040 generating peroxynitrite. In turn, peroxynitrite nitrates endogenous MnSOD inactivating this
3041 enzyme and maintaining O₂^{•-} levels [10]. These processes promote maintenance of

3042 inflammatory response and persistence of inflammatory pain. Furthermore, $O_2^{\bullet-}$ can promote
3043 mechanical hyperalgesia alone, e.g. without nerve injury and independently to react with
3044 nitric oxide as demonstrated in a model using antimycin A (mitochondrial complex III
3045 inhibitor) as $O_2^{\bullet-}$ generator. These data corroborate the notion that $O_2^{\bullet-}$ is an important
3046 hyperalgesic component as well as being a therapeutic target [11].

3047 Non-steroidal anti-inflammatory drugs are widely used to treat inflammatory disorders
3048 and pain relief in clinical practice because of their fast onset of action and excellent curative
3049 effects. However, non-steroidal anti-inflammatory drugs induce side effects including
3050 ulceration in the gastrointestinal system [12] and platelet dysfunction [13]. In this sense, the
3051 research on novel anti-inflammatory drugs persists. Vegetables, fruits and drinks (e.g. wine
3052 and tea) of the human diet contain polyphenols. Daily doses of these compounds enhance life
3053 quality and decrease the risk of many inflammatory diseases, such as cardiovascular diseases
3054 [14]. Curcumin [(E,E)-1,7-bis(4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione] is a
3055 yellow pigment present in the rhizome of the plant *Curcuma longa* considered a promising
3056 compound. Apart from its daily use in the kitchens as a condiment and spice, turmeric
3057 containing curcumin has been widely applied by the traditional medicine practitioners in
3058 South Asia for thousands of years to treat dysentery, ulcers, wounds, cough, fever and arthritis
3059 [15]. Furthermore, alimentary supplementation with curcumin seems to prevent
3060 neurodegenerative diseases such as Alzheimer's [16]. Curcumin is a highly pleiotropic
3061 molecule with antifungal [17], antiviral [18] antitumor [19] and antioxidant activities [20].
3062 Importantly, curcumin presents low toxicity at dose regimens of up to 8 g daily [21].
3063 Curcumin inhibits a plethora of molecular targets such as cyclooxygenase, lipoxygenase [22],
3064 matrix metalloproteinase 9 [23] and NF- κ B [24]. In addition, curcumin downregulates the
3065 expression of adhesion molecules, and consequently reduces leukocyte recruitment during
3066 inflammation [25, 26] and contributes to up regulation of transcription factor Nrf2 [27, 28].
3067 Curcumin exhibits poor bioavailability, but this issue has been substantially improved by
3068 various formulation strategies such as Meriva® (Indena SpA, Milan, Italy) documented in
3069 terms of comparative pharmacokinetics [29] and clinical efficacy as promising analgesic, anti-
3070 inflammatory and antioxidant medicine to several conditions [30-32].

3071 Taking into account $O_2^{\bullet-}$ is an important mediator to inflammatory nociceptor
3072 sensitization; the analgesic effect and mechanisms of action of curcumin in KO_2 (an $O_2^{\bullet-}$
3073 donor)-induced inflammation were investigated in the present study.

3074

3075 2. Material and Methods

3076 2.1. Animals

3077 Male Swiss mice (25-30 g) from the Universidade Estadual de Londrina, Paraná,
3078 Brasil, were used in this study. Mice were housed in standard clear plastic cages with free
3079 access to food and water and a light/dark cycle of 12:12 h and kept at 21° C. All behavioral
3080 testing were performed between 9 am and 5 pm in a temperature-controlled room. Animal
3081 care and handling procedures followed the International Association for the Study of Pain
3082 (IASP) guidelines. The Ethics Committee of the Universidade Estadual de Londrina approved
3083 this study (Process number 71.2012.68). All efforts were made to minimize the number of
3084 animals used and their suffering. The experimenters were blinded to the treatments.

3085

3086 2.2. Drugs

3087 The following materials were obtained from the sources indicated: Curcumin (Santa
3088 Cruz Biotechnology, USA), KO₂ (Alfa Aesar, MA, USA), and ABTS [2,2'-azino-bis(3-
3089 ethylbenzothiazoline-6-sulfonate)] (Sigma-Aldrich, St. Louis, MO, USA), NBT (nitroblue
3090 tetrazolium) (Amresco, Solon, OH, USA).

3091

3092 2.3. Experimental procedures

3093 Mice were pretreated subcutaneously (s.c.) with 3, 10, or 30 mg/kg of curcumin or
3094 with vehicle (2% DMSO in saline) 1 h before intraplantar (i.pl.) or intraperitoneal (i.p.)
3095 injection of 30 µg or 1 mg of KO₂, respectively. All tests were performed injecting the
3096 stimulus in the paw except by writhing response. At indicated time points the following
3097 parameters were determined: writhing response, paw flinching and time spent licking the paw
3098 (Fig. 1), mechanical and thermal hyperalgesia (Fig. 2), myeloperoxidase activity in paw skin
3099 samples and peritoneal leukocyte recruitment (Fig. 3), cytokine (TNF-α, IL-1β and IL-10)
3100 production and NF-κB activation (Fig. 4), ABTS assay, O₂^{•-} quantification and gp91^{phox}
3101 mRNA expression (Fig. 5), and Nrf2 and HO-1 mRNA expression (Fig. 6). The doses of KO₂
3102 and time points for sample collection were determined in standardizing experiments in our
3103 laboratory, previous study of our laboratory and described in detail in the following sections
3104 [33]. KO₂ was diluted in saline immediately before application. Based on the results from
3105 behavioral tests, the dose of curcumin of 10 mg/kg was chosen and used in selected
3106 experiments.

3107

3108 2.4. Overt pain-like behavioral tests

3109 Abdominal writhing was induced by i.p. injection of 1 mg of KO₂ [33]. Immediately
3110 after stimulus injection, each mouse was placed individually in a large glass cylinder, and the
3111 intensity of nociceptive behavior was quantified by counting the total number of writhings
3112 occurring between 0 and 20 min after stimulus injection. The writhing response consists of a
3113 contraction of the abdominal muscle together with a stretching of hind limbs. The intensity of
3114 the writhing response was expressed as the cumulative number of abdominal contortions over
3115 20 min.

3116 The number of paw flinches and the time spent licking the paw were determined
3117 between 0 and 30 min after i.pl. injection of 30 µg of KO₂. Each mouse was placed in a large
3118 glass cylinder immediately after stimulus injection. The intensity of nociceptive behavior was
3119 quantified by counting the total number of paw flinches and the time (seconds) spent licking
3120 ipsilateral paw [33].

3121

3122 2.5. Mechanical hyperalgesia test

3123 Mechanical hyperalgesia was evaluated by electronic version of von Frey's filaments
3124 as reported previously [34]. In a quiet room, temperature controlled, mice were placed in
3125 acrylic cages (12 × 10 × 17 cm) with wire grid floors 15-30 min before the start of testing.
3126 The test consisted of evoking a hind paw flexion reflex with a handheld force transducer
3127 (electronic anesthesiometer, IITC Life Science, Woodland Hills, CA) adapted with a 0.5 mm²
3128 polypropylene tip. The investigator was trained to apply the tip perpendicularly to the central
3129 area of the plantar hind paw with a gradual increase in pressure. The gradual increase in
3130 pressure was manually performed in blinded experiments. The upper limit pressure was 15 g.
3131 The end-point was characterized by the removal of the paw followed by clear flinching
3132 movements. After paw withdrawal, the intensity of the pressure was automatically recorded,
3133 and the final value for the response was obtained by averaging three measurements. The
3134 animals were tested before and after treatments. The results are expressed by delta (Δ)
3135 withdrawal threshold (in grams) calculated by subtracting the mean measurements 0.5, 1, 3, 5
3136 and 7 h after stimulus from the zero-time mean measurements [33].

3137

3138 2.6 Thermal hyperalgesia test

3139 Heat thermal hyperalgesia was performed using a hot plate at 55°C ± 1°C. The test
3140 was performed at same intervals on 2.5 [33]. The end-point was characterized by the removal
3141 of the paw followed by clear paw flinching or licking movements. The upper time was 20

3142 seconds to avoid possible injury. The results are expressed by delta (Δ) withdraw latency (in
3143 seconds) calculated by subtracting the mean of zero-time measurements from the mean of
3144 measurements obtained 0.5, 1, 3, 5 and 7 h after stimulus [35].

3145

3146 2.7. Myeloperoxidase (MPO) assay

3147 Neutrophil migration to the hind paw skin tissue of mice was evaluated using an MPO
3148 kinetic-colorimetric assay as described previously [33]. Samples of paw skin tissue were
3149 collected 7 h after stimulus in ice-cold 50 mM K_2HPO_4 buffer (pH 6.0) containing 0.5%
3150 hexadecyltrimethylammonium bromide (HTAB) and kept at $-80^\circ C$ until use. Samples were
3151 homogenized, centrifuged ($16,100 g \times 4 \text{ min}$), and the resulting supernatant was assayed for
3152 MPO activity spectrophotometrically at 450 nm (Multiskan GO Microplate
3153 Spectrophotometer, Thermo Scientific, Vantaa, Finland), with three readings in 1 min. The
3154 MPO activity of samples was compared to a standard curve of neutrophils. Briefly, 10 μL of
3155 sample was mixed with 200 μL of 50 mM phosphate buffer, pH 6.0, containing 0.167 mg/mL
3156 *o*-dianisidine dihydrochloride and 0.015% hydrogen peroxide. The results are presented as
3157 MPO activity (number of neutrophils $\times 10^4$ per mg of tissue).

3158

3159 2.8. Leukocyte recruitment in the peritoneal cavity

3160 Leukocyte recruitment to the peritoneal cavity was evaluated 7 h after i.p. injection of
3161 KO_2 (30 μg /cavity) [33]. Peritoneal cavities were washed with 200 μL of phosphate-buffered
3162 saline (PBS). Total leukocyte counts were performed in a Neubauer chamber after dilution in
3163 Turk's solution (2% acetic acid). Differential cell counts were performed using the Fast
3164 Panoptic Kit for histological analysis (Laborclin, Pinhais, PR, Brasil), and the values are
3165 expressed as the number of cells ($\times 10^6$) per cavity. Total and differential cell counts were
3166 performed under a light microscope (400 \times magnification, Olympus Optical Co., Hamburg,
3167 Germany).

3168

3169 2.9. ABTS assay

3170 The ability of samples to resist oxidative damage was determined by their ability to
3171 scavenge the ABTS radical. The test was performed as described previously [36]. Samples
3172 were collected 3 h after i.pl. stimulus and homogenized immediately in ice-cold KCl buffer
3173 (500 μL , 1.15% weight/volume). The homogenates were centrifuged ($200 g \times 10 \text{ min} \times 4^\circ C$),
3174 and the supernatants were used in the assay. Diluted ABTS solution (200 μL) was mixed with
3175 10 μL of sample in each well. After 6 min of incubation ($25^\circ C$), the absorbance was

3176 measured at 730 nm (Multiskan GO, Thermo Scientific). The results were equated against a
3177 standard Trolox curve (0.02 – 20 μmol).

3178

3179 2.10. $\text{O}_2^{\bullet-}$ production

3180 The quantitation of $\text{O}_2^{\bullet-}$ production in tissue homogenates was performed using the
3181 NBT assay as described previously [36]. Skin samples were collected 3 h after the stimulus.
3182 Briefly, 50 μL of the homogenate was incubated with 100 μl of NBT (1 mg/mL) in 96-well
3183 plates at 37 °C for 1 h. The supernatant was carefully removed and the reduced formazan
3184 solubilized by adding 120 μL of 2 M KOH and 140 μL of DMSO. The NBT reduction was
3185 measured at 600 nm using a microplate spectrophotometer reader (Multiskan GO, Thermo
3186 Scientific). The tissue weight was used for data normalization, thus the results are expressed
3187 as NBT reduction (OD/mg of tissue).

3188

3189 2.11. Cytokine measurement

3190 Skin samples were collected 3 hours after the injection of KO_2 , and homogenized in
3191 500 μL of ice-cold buffer containing protease inhibitors, and centrifuged (3000 rpm \times 10 min
3192 \times 4 °C), and the supernatants used to measure IL-1 β , TNF- α and IL-10 levels by an enzyme-
3193 linked immunosorbent assay (ELISA) using eBioscience kits. As a control, the concentrations
3194 of these cytokines were determined in animals injected with saline. The results are expressed
3195 as picograms (pg) of cytokine/100 mg of tissue.

3196

3197 2.12. NF- κB activity

3198 Skin samples were collected 3 h after stimulus and homogenized in ice-cold lysis
3199 buffer (Cell Signaling). The homogenates were centrifuged (200 g \times 10 min \times 4 °C), and the
3200 supernatants were used to assess the levels of total and phosphorylated NF- κB p65 subunit by
3201 ELISA using PathScan® kits (Cell Signaling) at 450 nm (Multiskan GO Thermo Scientific)
3202 according to the manufacturer's directions. The results are expressed as NF- κB activity (total-
3203 p65/phospho-p65 ratio).

3204

3205 2.13. Reverse transcription and quantitative polymerase chain reaction (RT-qPCR).

3206 RT-qPCR was performed as previously described [36]. Skin samples were collected 3
3207 h after stimulus and homogenized in trizol reagent, and total RNA was isolated according to
3208 manufacturer's directions. The purity of total RNA was measured with a spectrophotometer
3209 and the wavelength absorption ratio (260/280 nm) was between 1.8 and 2.0 for all

3210 preparations. Reverse transcription of total RNA to cDNA, and qPCR were carried out using
3211 GoTaq® 2-Step RT-qPCR System (Promega) and specific primers (Applied Biosystems®).
3212 The relative gene expression was measured using the comparative $2^{-(\Delta\Delta Cq)}$ method. The
3213 primers used were: Nrf2, sense: 5'-TCACACGAGATGAGCTTAGGGCAA-3', antisense: 5'-
3214 TACAGTTCTGGGCGGCGACTTTAT-3'; gp91^{phox}, sense: 5'-
3215 AGCTATGAGGTGGTGATGTTAGTGG-3', antisense: 5'-
3216 CACAATATTTGTACCAGACAGACTTGAG-3'; Heme Oxygenase-1 (HO-1), sense: 5'-
3217 CCCAAAACCTGGCCTGTAAAA-3', antisense: 5'-CGTGGTCAGTCAACATGGAT-3'; β -
3218 actin, sense: 5'-AGCTGCGTTTTACACCCTTT-3', antisense: 5'-
3219 AAGCCATGCCAATGTTGTCT-3'. The expression of β -actin mRNA was used as a
3220 reference gene to normalize data.

3221

3222 2.14. Statistical analysis

3223 Results are presented as means \pm SEM of measurements made on six mice in each
3224 group per experiment and are representative of two separate experiments. Two-way repeated
3225 measures analysis of variance (ANOVA) followed by Tukey's *post hoc* was used to compare
3226 all groups and doses at all times when responses were measured at different times after the
3227 stimulus injection. Differences between responses were evaluated by one-way ANOVA
3228 followed by Tukey's *post hoc* for data of single time point. Statistical differences were
3229 considered significant when $P < 0.05$.

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3238 3. Results

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3240 3.1. Curcumin reduces KO₂-induced overt pain-like behaviors

3241 Mice received curcumin (3, 10, 30 mg/kg, s.c.) or vehicle (2% DMSO in saline)
3242 treatment 1 h before injection of KO₂ (1 mg, i.p.). The total number of writhing was
3243 determined between 0-20 min. Curcumin at the doses of 10 and 30 mg/kg reduced
3244 significantly the number of writhings (Fig. 1a), without differences between these two doses.
3245 Curcumin at 3 mg/kg was unable to show significantly inhibitory effect (Fig. 1a). Thus, the
3246 dose of 10 mg/kg of curcumin was chosen for the next experiments (Fig. 1b, 1c, 2-6).
3247 Curcumin also inhibited KO₂-induced paw flinches (Fig. 1b) and time spent licking the paw
3248 (Fig. 1c) between 0-30 min.

3249

3250 3.2. Curcumin reduces KO₂-induced mechanical and thermal hyperalgesia

3251 Mice received curcumin (10 mg/kg, s.c.) or vehicle treatment 1 h before KO₂ injection
3252 (30 µg/paw) followed by evaluation of mechanical and thermal hyperalgesia. Curcumin
3253 decreased mechanical hyperalgesia at all time points compared to vehicle-treated animals
3254 (Fig. 2a). Pretreatment with curcumin also reduced thermal hyperalgesia (Fig. 2b).

3255

3256 3.3. Curcumin reduces KO₂-induced myeloperoxidase (MPO) activity

3257 Mice received curcumin (10 mg/kg, s.c.) or vehicle treatment 1 h before KO₂ injection
3258 (30 µg/paw). KO₂ induced a significant increase of MPO activity in the paw skin 7 h after its
3259 injection. Curcumin pretreatment reduced KO₂-induced increase of MPO activity (Fig. 3a).
3260 Furthermore, the leukocyte recruitment in the peritoneal cavity was evaluated 7 h after KO₂
3261 i.p. injection (30 µg/cavity). KO₂ increased the total number of leukocytes (Fig 3b),
3262 mononuclear cells (Fig 3c) and neutrophils (Fig 3d) whilst curcumin pretreatment reduced
3263 KO₂-induced recruitment of these cells.

3264

3265 3.4. Effect of curcumin on KO₂-induced cytokine production and NF-κB activation

3266 KO₂ induced significant increase of TNF-α (Fig. 4a) and IL-1β (Fig. 4b) levels in the
3267 paw skin at 3 h after its injection. Curcumin reduced KO₂-induced production of the
3268 hyperalgesic cytokines TNF-α (Fig. 4a) and IL-1β (Fig. 4b). On the other hand, KO₂ induces a
3269 significant increase in IL-10 levels, an anti-hyperalgesic cytokine. Curcumin further increased
3270 the KO₂-induced IL-10 production in the paw skin (Fig. 4c). The total NF-κB p65
3271 concentration remained constant, and the phosphorylated NF-κB p65 increased upon KO₂

3272 stimulus resulting in decreased total NF- κ B p65/phosphorylated NF- κ B p65 ratio. In
3273 agreement with the curcumin inhibition of pro-hyperalgesic cytokine production, it also
3274 inhibited KO₂-induced NF- κ B activation as observed by an increase of the total NF- κ B
3275 p65/phosphorylated NF- κ B p65 ratio (Fig. 4d).

3276

3277 3.5. Curcumin reduces KO₂-induced oxidative stress

3278 O₂^{•-} donor depleted antioxidant capacity and increased O₂^{•-} production in the plantar
3279 tissue (Fig. 5A and 5B). On the other hand, curcumin impaired these deleterious effects,
3280 restoring the antioxidant capacity and inhibiting the increases in O₂^{•-} production (Fig. 5a and
3281 5b). Corroborating these results, we demonstrated pretreatment with curcumin reduced the
3282 mRNA expression of gp91^{phox}, NADPH oxidase subunit (Fig. 5C).

3283

3284 3.6. Curcumin increases mRNA expression for Nrf2 and heme oxygenase-1 (HO-1)

3285 KO₂ reduced while did not affect the mRNA expression for Nrf2 (Fig. 6a) and HO-1
3286 (Fig. 6b) at 3 h, respectively. In turn, curcumin treatment enhanced Nrf2 mRNA expression
3287 (Fig. 6a) by 1.5 fold and HO-1 mRNA expression by 9.5 fold (Fig. 6b).

3288 4. Discussion

3289 Pain is one of the most common clinical signs of inflammation. Therapeutic
3290 approaches to this condition should target the mediators involved in its development, which
3291 include neuropeptides, eicosanoids, cytokines and reactive oxygen species (ROS). These
3292 mediators are closely related since cytokines including TNF- α and IL-1 β induce the
3293 production of eicosanoids and O₂^{•-} while the latter induces the production of cytokines by
3294 activating the NF- κ B pathway [9]. O₂^{•-} is important to the development of thermal
3295 hyperalgesia associated with acute and chronic inflammation [10, 37]; and promotes
3296 mechanical hyperalgesia [11]. Accordingly, treatment with Tempol (SOD mimetic) inhibited
3297 the hyperalgesia and inflammation in carrageenan-induced pain [38]. Hence, we demonstrated
3298 that KO₂ is a useful model to trigger a great variety of O₂^{•-} dependent nociceptive responses
3299 such as abdominal writhings, paw flinches and licking, and mechanical and thermal
3300 hyperalgesia [33]. Altogether, strategies that control O₂^{•-} overproduction and/or its deleterious
3301 effects, including oxidative stress and inflammation, seem promising therapeutic approaches
3302 for pain relief.

3303 The mechanism of action of curcumin depends on a synergism involving weak
3304 targeting of varied proteins within related signaling networks [39]. In inflammatory pain
3305 context, curcumin inhibited overt pain-like behavior in formalin test [40], which has two
3306 components. The early phase (0-5 min) dependent on activation of nociceptors through
3307 TRPA1 receptors, which are important receptors in inflammatory pain and their increased
3308 activity, occurs in O₂^{•-} production dependent manner [41]. The late phase (15-30 min)
3309 depends on inflammatory cytokines production [42, 43]. In the present study, we observed
3310 that curcumin inhibited overt pain-like behaviors, and this may be due to the initial limitation
3311 of direct action O₂^{•-} on nociceptors and capacity of desensitizing TRPA1 receptors [44].
3312 Importantly, curcumin inhibited both paw flinching and licking induced by O₂^{•-}. At least in
3313 the formalin test, paw flinching is considered a peripheral and spinal response and paw licking
3314 presents the structural mechanisms of flinching plus supraspinal nociceptive structures [45,
3315 46]. Therefore, it is likely O₂^{•-} activates peripheral, spinal and supraspinal responses, which
3316 are amenable to curcumin.

3317 In addition to the direct effect on nociceptors, O₂^{•-} activates signaling pathways that
3318 culminate in the production of inflammatory mediators by resident cells promoting neutrophil
3319 recruitment to local of stimulus. In fact, O₂^{•-} contributes to neutrophil recruitment in
3320 inflammatory conditions [47]. Recruited neutrophils produce O₂^{•-}, IL-1 β , TNF- α and

3321 prostaglandin E₂, and thus contribute to the maintenance of inflammatory hyperalgesia [7].
3322 The MPO is an enzyme found in azurophilic granules of neutrophils and macrophages, and
3323 therefore, is an indirect marker of these cells [48]. MPO produces the microbicidal molecule
3324 hypochlorite, a strong oxidant, upon reaction with H₂O₂ and Cl⁻. Curcumin inhibits the
3325 expression of adhesion molecules [25, 26], resulting in the reduction of cell counts in the
3326 inflammatory foci. In agreement, we observed that curcumin inhibited MPO activity in the
3327 paw skin, and total leukocytes, mononuclear cells and neutrophils in the peritoneal cavity.

3328 The transcription factor NF-κB is a master regulator of both innate and adaptive
3329 immune responses. The phosphorylation of IκB (NF-κB inhibitor) allows NF-κB dimer to
3330 translocate from the cytoplasm to the nucleus where it regulates the transcription of a large
3331 number of genes, including pro-inflammatory cytokines and chemokines [49]. M40403 (SOD
3332 mimetic) decreased LPS-induced activation of NF-κB, and this effect was associated with
3333 diminished degradation of cytoplasmic IκB [50], suggesting that O₂^{•-} participates in redox-
3334 regulated NF-κB activation. Curcumin attenuates the development of asthma by inhibiting
3335 NF-κB activation [51]. Corroborating these data, we demonstrated that curcumin decreased
3336 O₂^{•-}-induced NF-κB activation *in vivo* as observed by prevention of KO₂-induced decrease of
3337 the total NF-κB p65/phosphorylated NF-κB p65 ratio. Consequently, KO₂-induced IL-1β and
3338 TNF-α production was reduced by curcumin. These pro-inflammatory cytokines undergo
3339 rapid release and have a pivotal role in the development of hyperalgesia due the recruitment
3340 and activation of immune cells. O₂^{•-} also modulates the activity of TRPV1, an important
3341 component of thermal hyperalgesia [52] and curcumin has shown the capacity to antagonize
3342 TRPV1 [53]. In agreement with that, curcumin decreased KO₂-induced mechanical and
3343 thermal hyperalgesia.

3344 NADPH oxidases are a family of enzyme complexes present in both phagocytic and
3345 non-phagocytic cells, which catalyze the transfer of electrons from NADPH to molecular
3346 oxygen, thereby generating O₂^{•-} and H₂O₂. O₂^{•-} contributes to nociceptor sensitization and as
3347 well as other ROS act as signaling molecule to trigger IL-1β production through activation of
3348 inflammasomes [54]. Therefore, modulating oxidative stress is a conceivable approach to
3349 diminish pain and inflammation [55, 56]. Curcumin presents prominent antioxidant activity
3350 and contributes to increasing GSH, catalase and SOD levels [57]. In fact, curcumin inhibited
3351 thermal hyperalgesia by controlling oxidative stress and normalizing peripheral and spinal
3352 antioxidant enzyme system, which contributed to reducing pro-inflammatory cytokines
3353 production in complete Freund's adjuvant (CFA) inflammation [58]. In addition, curcumin

3354 scavenges DPPH[•] and DMPD^{•+} radicals and improve ferric reducing antioxidant power
3355 (FRAP) *in vitro* [59]. The present data show that curcumin improves the antioxidant capacity
3356 *in vivo* by inhibiting O₂^{•-} production and maintaining the antioxidant defenses observed by re-
3357 establishment of ABTS radical scavenger activity.

3358 Nrf2 is a potent transcriptional factor constantly ubiquitinated through Keap1 and
3359 degraded in the proteasome under regular conditions. In response to oxidative and
3360 electrophilic stress, Keap1 is inactivated, and Nrf2 translocates to nucleus and binds at a
3361 promoter sequence known as the antioxidant-responsive element (ARE) that is found in many
3362 cytoprotective genes. In general, target genes of Nrf2 are involved in glutathione synthesis
3363 and elimination of ROS, providing a protective function against oxidative stress [60, 61].
3364 Evidence demonstrates that curcumin increases Nrf2 activity [27, 28], which lines up well
3365 with a 1.5-fold increase in Nrf2 mRNA expression found in our study. We found O₂^{•-}
3366 decreased mRNA expression of Nrf2, which might contribute to the impairment of an
3367 efficient endogenous antioxidant response. In line with this hypothesis, we found that
3368 curcumin potentiated the O₂^{•-}-induced increased mRNA expression of heme oxygenase-1
3369 (HO-1), a target of Nrf2 transcriptional activity [62]. The crosstalk mechanism between NF-
3370 κB and Nrf2 is puzzling and is still under elucidation. After O₂^{•-} producing stimulus such as
3371 PMA, the p65 subunit competes with Nrf2 for the coactivator CREB-binding protein. As a
3372 consequence, there is hypoacetylation blocking chromatin condensation, which suppresses
3373 Nrf2/ARE gene expression and dampens Nrf2 pathway activation [63]. This is a possible
3374 explanation for the KO₂-induced decrease of Nrf2 mRNA expression in the present study.

3375 HO-1 is a ubiquitous inducible cellular stress enzyme and one of the target genes of
3376 Nrf2. The HO-1 major metabolic function is to catalyze the reaction of heme that leads to the
3377 formation of carbon monoxide, free iron and biliverdin [62]. This reaction allows biliverdin
3378 conversion into the strong antioxidant bilirubin, and ROS participate in the reaction that
3379 converts back bilirubin into biliverdin [64]. This cycle grants the neutralization of ROS and
3380 provides an antioxidant function to HO-1 [64]. Activation of cGMP/ PKG/ ATP-sensitive
3381 potassium channels lead to antinociceptive state [55, 65]. Co-treatment with an HO-1 inducer
3382 increased the antinociceptive effects of opioids and cannabinoids through activation of cGMP/
3383 PKG/ ATP-sensitive potassium channels pathway in a model of CFA-induced pain [66]. On
3384 the other hand, the administration of HO-1 inhibitor increases mechanical hyperalgesia [67].
3385 Several antioxidant molecules can increase HO-1 expression, and curcumin is one of them
3386 [27, 28, 68], a feature that we verified in our model with a 9.5-fold increase of expression.

3387 Interleukin-10 (IL-10) is an anti-inflammatory and anti-hyperalgesic cytokine [2]. IL-
3388 10 deficiency enhances acute exercise-induced muscle hyperalgesia concomitantly with
3389 exacerbated oxidative stress [69]. On the other hand, increased nuclear levels of Nrf2 were
3390 accompanied by high levels of IL-10 in the plasma and the renal proximal tubules of
3391 treadmill-exercised rats [70]. In another model, hypochlorous acid-induced oxidative stress
3392 activated Nrf2 pathway and enhanced mRNA expression of IL-10 [71]. IL-10 was necessary
3393 to increase the expression of HO-1 in LPS-induced endotoxemia [72], suggesting the
3394 existence of an IL-10/HO-1 axis [72], which also may explain the prominent increase of HO-1
3395 mRNA expression found in our study.

3396 In conclusion, we found that curcumin reduced nociceptive responses induced by
3397 KO₂-induced O₂[•] through mechanisms including NF-κB inhibition and induction of Nrf2
3398 mRNA expression, which contribute to inhibit *de novo* O₂[•] generation, IL-1β and TNF-α
3399 production, and enhance the levels of HO-1 and IL-10. This study also contributes to support
3400 that O₂[•] is a fundamental component of inflammatory hyperalgesia, further advances in the
3401 knowledge of the curcumin mechanisms of action, and supports the use of curcumin as an
3402 analgesic.

3403

3404 **Conflict of interest**

3405 The authors declare no conflict of interest.

3406

3407 **Acknowledgments**

3408 Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), São Paulo
3409 Research Foundation (FAPESP) under grant agreements number 2011/19670-0 (Thematic
3410 project) and 2013/08216-2 (Center for Research in Inflammatory Disease-CRID),
3411 Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Ministério da
3412 Ciência Tecnologia e Inovação (MCTI), Secretaria da Ciência, Tecnologia e Ensino Superior
3413 (SETI), Fundação Araucária and Parana State Government grants supported this study
3414 (Brazil).

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3419 **References**

- 3420 1. Goldberg DS, McGee SJ. Pain as a global public health priority. *BMC Public Health* 2011;
3421 11:770.
- 3422 2. Verri WA, Jr., Cunha TM, Parada CA, Poole S, Cunha FQ, Ferreira SH. Hypernociceptive role of
3423 cytokines and chemokines: targets for analgesic drug development? *Pharmacol Ther* 2006;
3424 112:116-38.
- 3425 3. Jin X, Gereau RWt. Acute p38-mediated modulation of tetrodotoxin-resistant sodium channels in
3426 mouse sensory neurons by tumor necrosis factor-alpha. *J Neurosci* 2006; 26:246-55.
- 3427 4. Binshtok AM, Wang H, Zimmermann K, Amaya F, Vardeh D, Shi L, et al. Nociceptors are
3428 interleukin-1beta sensors. *J Neurosci* 2008; 28:14062-73.
- 3429 5. Cunha TM, Verri WA, Jr., Silva JS, Poole S, Cunha FQ, Ferreira SH. A cascade of cytokines
3430 mediates mechanical inflammatory hypernociception in mice. *Proc Natl Acad Sci U S A* 2005;
3431 102:1755-60.
- 3432 6. Zarpelon AC, Cunha TM, Alves-Filho JC, Pinto LG, Ferreira SH, McInnes IB, et al. IL-33/ST2
3433 signalling contributes to carrageenin-induced innate inflammation and inflammatory pain: role of
3434 cytokines, endothelin-1 and prostaglandin E2. *Br J Pharmacol* 2013; 169:90-101.
- 3435 7. Cunha TM, Verri WA, Jr., Schivo IR, Napimoga MH, Parada CA, Poole S, et al. Crucial role of
3436 neutrophils in the development of mechanical inflammatory hypernociception. *J Leukoc Biol*
3437 2008; 83:824-32.
- 3438 8. McDonald B, Pittman K, Menezes GB, Hirota SA, Slaba I, Waterhouse CC, et al. Intravascular
3439 danger signals guide neutrophils to sites of sterile inflammation. *Science* 2010; 330:362-6.
- 3440 9. Cuzzocrea S, Pisano B, Dugo L, Ianaro A, Ndengele M, Salvemini D. Superoxide-related
3441 signaling cascade mediates nuclear factor-kappaB activation in acute inflammation. *Antioxid*
3442 *Redox Signal* 2004; 6:699-704.
- 3443 10. Wang ZQ, Porreca F, Cuzzocrea S, Galen K, Lightfoot R, Masini E, et al. A newly identified role
3444 for superoxide in inflammatory pain. *J Pharmacol Exp Ther* 2004; 309:869-78.
- 3445 11. Kim HY, Chung JM, Chung K. Increased production of mitochondrial superoxide in the spinal
3446 cord induces pain behaviors in mice: the effect of mitochondrial electron transport complex
3447 inhibitors. *Neurosci Lett* 2008; 447:87-91.
- 3448 12. Hawkey CJ. Nonsteroidal anti-inflammatory drug gastropathy. *Gastroenterology* 2000; 119:521-
3449 35.
- 3450 13. Aygun D, Kaplan S, Odaci E, Onger ME, Altunkaynak ME. Toxicity of non-steroidal anti-
3451 inflammatory drugs: a review of melatonin and diclofenac sodium association. *Histol Histopathol*
3452 2012; 27:417-36.
- 3453 14. McCullough ML, Peterson JJ, Patel R, Jacques PF, Shah R, Dwyer JT. Flavonoid intake and
3454 cardiovascular disease mortality in a prospective cohort of US adults. *Am J Clin Nutr* 2012;
3455 95:454-64.
- 3456 15. Aggarwal BB, Sung B. Pharmacological basis for the role of curcumin in chronic diseases: an
3457 age-old spice with modern targets. *Trends Pharmacol Sci* 2009; 30:85-94.

- 3458 16. Calabrese V, Butterfield DA, Stella AM. Nutritional antioxidants and the heme oxygenase
3459 pathway of stress tolerance: novel targets for neuroprotection in Alzheimer's disease. *Ital J*
3460 *Biochem* 2003; 52:177-81.
- 3461 17. Apisariyakul A, Vanittanakom N, Buddhasukh D. Antifungal activity of turmeric oil extracted
3462 from *Curcuma longa* (Zingiberaceae). *J Ethnopharmacol* 1995; 49:163-9.
- 3463 18. Kim HJ, Yoo HS, Kim JC, Park CS, Choi MS, Kim M, et al. Antiviral effect of *Curcuma longa*
3464 Linn extract against hepatitis B virus replication. *J Ethnopharmacol* 2009; 124:189-96.
- 3465 19. Meng QX, Roubin RH, Hanrahan JR. Ethnopharmacological and bioactivity guided investigation
3466 of five TCM anticancer herbs. *J Ethnopharmacol* 2013; 148:229-38.
- 3467 20. Selvam R, Subramanian L, Gayathri R, Angayarkanni N. The anti-oxidant activity of turmeric
3468 (*Curcuma longa*). *J Ethnopharmacol* 1995; 47:59-67.
- 3469 21. Shen LR, Parnell LD, Ordovas JM, Lai CQ. Curcumin and aging. *Biofactors* 2013; 39:133-40.
- 3470 22. Hong J, Bose M, Ju J, Ryu JH, Chen X, Sang S, et al. Modulation of arachidonic acid metabolism
3471 by curcumin and related beta-diketone derivatives: effects on cytosolic phospholipase A(2),
3472 cyclooxygenases and 5-lipoxygenase. *Carcinogenesis* 2004; 25:1671-9.
- 3473 23. Menon LG, Kuttan R, Kuttan G. Anti-metastatic activity of curcumin and catechin. *Cancer Lett*
3474 1999; 141:159-65.
- 3475 24. Aggarwal S, Ichikawa H, Takada Y, Sandur SK, Shishodia S, Aggarwal BB. Curcumin
3476 (diferuloylmethane) down-regulates expression of cell proliferation and antiapoptotic and
3477 metastatic gene products through suppression of IkappaBalpha kinase and Akt activation. *Mol*
3478 *Pharmacol* 2006; 69:195-206.
- 3479 25. Gupta B, Ghosh B. *Curcuma longa* inhibits TNF-alpha induced expression of adhesion molecules
3480 on human umbilical vein endothelial cells. *Int J Immunopharmacol* 1999; 21:745-57.
- 3481 26. Kumar A, Dhawan S, Hardegen NJ, Aggarwal BB. Curcumin (Diferuloylmethane) inhibition of
3482 tumor necrosis factor (TNF)-mediated adhesion of monocytes to endothelial cells by suppression
3483 of cell surface expression of adhesion molecules and of nuclear factor-kappaB activation.
3484 *Biochem Pharmacol* 1998; 55:775-83.
- 3485 27. Gao S, Duan X, Wang X, Dong D, Liu D, Li X, et al. Curcumin attenuates arsenic-induced
3486 hepatic injuries and oxidative stress in experimental mice through activation of Nrf2 pathway,
3487 promotion of arsenic methylation and urinary excretion. *Food Chem Toxicol* 2013; 59:739-47.
- 3488 28. Heeba GH, Mahmoud ME, El Hanafy AA. Anti-inflammatory potential of curcumin and
3489 quercetin in rats: Role of oxidative stress, heme oxygenase-1 and TNF-alpha. *Toxicol Ind Health*
3490 2012.
- 3491 29. Cuomo J, Appendino G, Dern AS, Schneider E, McKinnon TP, Brown MJ, et al. Comparative
3492 absorption of a standardized curcuminoid mixture and its lecithin formulation. *J Nat Prod* 2011;
3493 74:664-9.
- 3494 30. Belcaro G, Cesarone MR, Dugall M, Pellegrini L, Ledda A, Grossi MG, et al. Efficacy and safety
3495 of Meriva(R), a curcumin-phosphatidylcholine complex, during extended administration in
3496 osteoarthritis patients. *Altern Med Rev* 2010; 15:337-44.

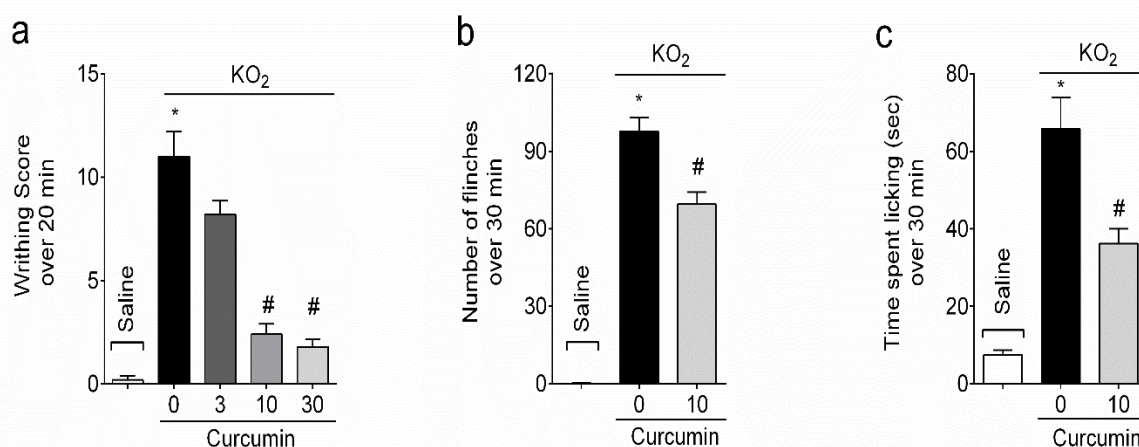
- 3497 31. Di Pierro F, Rapacioli G, Di Maio EA, Appendino G, Franceschi F, Togni S. Comparative
3498 evaluation of the pain-relieving properties of a lecithinized formulation of curcumin
3499 (Meriva(R)), nimesulide, and acetaminophen. *J Pain Res* 2013; 6:201-5.
- 3500 32. Drobnic F, Riera J, Appendino G, Togni S, Franceschi F, Valle X, et al. Reduction of delayed
3501 onset muscle soreness by a novel curcumin delivery system (Meriva(R)): a randomised, placebo-
3502 controlled trial. *J Int Soc Sports Nutr* 2014; 11:31.
- 3503 33. Maioli NA, Zarpelon AC, Mizokami SS, Calixto-Campos C, Guazelli CF, Hohmann MS, et al.
3504 The superoxide anion donor, potassium superoxide, induces pain and inflammation in mice
3505 through production of reactive oxygen species and cyclooxygenase-2. *Braz J Med Biol Res* 2015;
3506 0:0.
- 3507 34. Cunha TM, Verri WA, Jr., Vivancos GG, Moreira IF, Reis S, Parada CA, et al. An electronic
3508 pressure-meter nociception paw test for mice. *Braz J Med Biol Res* 2004; 37:401-7.
- 3509 35. Lavich TR, Cordeiro RS, Silva PM, Martins MA. A novel hot-plate test sensitive to hyperalgesic
3510 stimuli and non-opioid analgesics. *Braz J Med Biol Res* 2005; 38:445-51.
- 3511 36. Hohmann MS, Cardoso RD, Pinho-Ribeiro FA, Crespigio J, Cunha TM, Alves-Filho JC, et al. 5-
3512 lipoygenase deficiency reduces acetaminophen-induced hepatotoxicity and lethality. *Biomed*
3513 *Res Int* 2013; 2013:627046.
- 3514 37. Ndengele MM, Cuzzocrea S, Esposito E, Mazzon E, Di Paola R, Matuschak GM, et al.
3515 Cyclooxygenases 1 and 2 contribute to peroxynitrite-mediated inflammatory pain
3516 hypersensitivity. *FASEB J* 2008; 22:3154-64.
- 3517 38. Khattab MM. TEMPOL, a membrane-permeable radical scavenger, attenuates peroxynitrite- and
3518 superoxide anion-enhanced carrageenan-induced paw edema and hyperalgesia: a key role for
3519 superoxide anion. *Eur J Pharmacol* 2006; 548:167-73.
- 3520 39. Zhou H, Beevers CS, Huang S. The targets of curcumin. *Curr Drug Targets* 2011; 12:332-47.
- 3521 40. Mittal N, Joshi R, Hota D, Chakrabarti A. Evaluation of antihyperalgesic effect of curcumin on
3522 formalin-induced orofacial pain in rat. *Phytother Res* 2009; 23:507-12.
- 3523 41. Fernandes ES, Vong CT, Quek S, Cheong J, Awal S, Gentry C, et al. Superoxide generation and
3524 leukocyte accumulation: key elements in the mediation of leukotriene B(4)-induced itch by
3525 transient receptor potential ankyrin 1 and transient receptor potential vanilloid 1. *FASEB J* 2013;
3526 27:1664-73.
- 3527 42. Chichorro JG, Lorenzetti BB, Zampronio AR. Involvement of bradykinin, cytokines, sympathetic
3528 amines and prostaglandins in formalin-induced orofacial nociception in rats. *Br J Pharmacol*
3529 2004; 141:1175-84.
- 3530 43. McNamara CR, Mandel-Brehm J, Bautista DM, Siemens J, Deranian KL, Zhao M, et al. TRPA1
3531 mediates formalin-induced pain. *Proc Natl Acad Sci U S A* 2007; 104:13525-30.
- 3532 44. Leamy AW, Shukla P, McAlexander MA, Carr MJ, Ghatta S. Curcumin ((E,E)-1,7-bis(4-
3533 hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione) activates and desensitizes the nociceptor
3534 ion channel TRPA1. *Neurosci Lett* 2011; 503:157-62.
- 3535 45. Donahue RR, LaGraize SC, Fuchs PN. Electrolytic lesion of the anterior cingulate cortex
3536 decreases inflammatory, but not neuropathic nociceptive behavior in rats. *Brain Res* 2001;
3537 897:131-8.

- 3538 46. Porro CA, Cavazzuti M, Lui F, Giuliani D, Pellegrini M, Baraldi P. Independent time courses of
3539 supraspinal nociceptive activity and spinally mediated behavior during tonic pain. *Pain* 2003;
3540 104:291-301.
- 3541 47. Finley A, Chen Z, Esposito E, Cuzzocrea S, Sabbadini R, Salvemini D. Sphingosine 1-phosphate
3542 mediates hyperalgesia via a neutrophil-dependent mechanism. *PLoS One* 2013; 8:e55255.
- 3543 48. Lazarevic-Pasti T, Leskovac A, Vasic V. Myeloperoxidase Inhibitors as Potential Drugs. *Curr*
3544 *Drug Metab* 2015; 16:168-90.
- 3545 49. Ghosh S, Hayden MS. New regulators of NF-kappaB in inflammation. *Nat Rev Immunol* 2008;
3546 8:837-48.
- 3547 50. Ndengele MM, Muscoli C, Wang ZQ, Doyle TM, Matuschak GM, Salvemini D. Superoxide
3548 potentiates NF-kappaB activation and modulates endotoxin-induced cytokine production in
3549 alveolar macrophages. *Shock* 2005; 23:186-93.
- 3550 51. Oh SW, Cha JY, Jung JE, Chang BC, Kwon HJ, Lee BR, et al. Curcumin attenuates allergic
3551 airway inflammation and hyper-responsiveness in mice through NF-kappaB inhibition. *J*
3552 *Ethnopharmacol* 2011; 136:414-21.
- 3553 52. Ibi M, Matsuno K, Shiba D, Katsuyama M, Iwata K, Kakehi T, et al. Reactive oxygen species
3554 derived from NOX1/NADPH oxidase enhance inflammatory pain. *J Neurosci* 2008; 28:9486-94.
- 3555 53. Yeon KY, Kim SA, Kim YH, Lee MK, Ahn DK, Kim HJ, et al. Curcumin produces an
3556 antihyperalgesic effect via antagonism of TRPV1. *J Dent Res* 2010; 89:170-4.
- 3557 54. Nakahira K, Haspel JA, Rathinam VA, Lee SJ, Dolinay T, Lam HC, et al. Autophagy proteins
3558 regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by
3559 the NALP3 inflammasome. *Nat Immunol* 2011; 12:222-30.
- 3560 55. Staurengo-Ferrari L, Mizokami SS, Fattori V, Silva JJ, Zanichelli PG, Georgetti SR, et al. The
3561 ruthenium nitric oxide donor, [Ru(HEDTA)NO], inhibits acute nociception in mice by
3562 modulating oxidative stress, cytokine production and activating the cGMP/PKG/ATP-sensitive
3563 potassium channel signaling pathway. *Naunyn Schmiedebergs Arch Pharmacol* 2014.
- 3564 56. Valerio DA, Georgetti SR, Magro DA, Casagrande R, Cunha TM, Vicentini FT, et al. Quercetin
3565 reduces inflammatory pain: inhibition of oxidative stress and cytokine production. *J Nat Prod*
3566 2009; 72:1975-9.
- 3567 57. Al-Omar FA, Nagi MN, Abdulgadir MM, Al Joni KS, Al-Majed AA. Immediate and delayed
3568 treatments with curcumin prevents forebrain ischemia-induced neuronal damage and oxidative
3569 insult in the rat hippocampus. *Neurochem Res* 2006; 31:611-8.
- 3570 58. Singh AK, Vinayak M. Curcumin attenuates CFA induced thermal hyperalgesia by modulation of
3571 antioxidant enzymes and down regulation of TNF-alpha, IL-1beta and IL-6. *Neurochem Res*
3572 2015; 40:463-72.
- 3573 59. Ak T, Gulcin I. Antioxidant and radical scavenging properties of curcumin. *Chem Biol Interact*
3574 2008; 174:27-37.
- 3575 60. Ishii T, Itoh K, Takahashi S, Sato H, Yanagawa T, Katoh Y, et al. Transcription factor Nrf2
3576 coordinately regulates a group of oxidative stress-inducible genes in macrophages. *J Biol Chem*
3577 2000; 275:16023-9.

- 3578 61. Manna K, Khan A, Kr Das D, Bandhu Kesh S, Das U, Ghosh S, et al. Protective effect of coconut
3579 water concentrate and its active component shikimic acid against hydroperoxide mediated
3580 oxidative stress through suppression of NF-kappaB and activation of Nrf2 pathway. *J*
3581 *Ethnopharmacol* 2014; 155:132-46.
- 3582 62. Pae HO, Son Y, Kim NH, Jeong HJ, Chang KC, Chung HT. Role of heme oxygenase in
3583 preserving vascular bioactive NO. *Nitric Oxide* 2010; 23:251-7.
- 3584 63. Liu GH, Qu J, Shen X. NF-kappaB/p65 antagonizes Nrf2-ARE pathway by depriving CBP from
3585 Nrf2 and facilitating recruitment of HDAC3 to MafK. *Biochim Biophys Acta* 2008; 1783:713-27.
- 3586 64. Kapitulnik J. Bilirubin: an endogenous product of heme degradation with both cytotoxic and
3587 cytoprotective properties. *Mol Pharmacol* 2004; 66:773-9.
- 3588 65. Zarpelon AC, Souza GR, Cunha TM, Schivo IR, Marchesi M, Casagrande R, et al. The nitroxyl
3589 donor, Angeli's salt, inhibits inflammatory hyperalgesia in rats. *Neuropharmacology* 2013; 71:1-
3590 9.
- 3591 66. Carcole M, Castany S, Leanez S, Pol O. Treatment with a heme oxygenase 1 inducer enhances
3592 the antinociceptive effects of micro-opioid, delta-opioid, and cannabinoid 2 receptors during
3593 inflammatory pain. *J Pharmacol Exp Ther* 2014; 351:224-32.
- 3594 67. Steiner AA, Branco LG, Cunha FQ, Ferreira SH. Role of the haeme oxygenase/carbon monoxide
3595 pathway in mechanical nociceptor hypersensitivity. *Br J Pharmacol* 2001; 132:1673-82.
- 3596 68. Bao W, Li K, Rong S, Yao P, Hao L, Ying C, et al. Curcumin alleviates ethanol-induced
3597 hepatocytes oxidative damage involving heme oxygenase-1 induction. *J Ethnopharmacol* 2010;
3598 128:549-53.
- 3599 69. Borghi SM, Pinho-Ribeiro FA, Zarpelon AC, Cunha TM, Alves-Filho JC, Ferreira SH, et al.
3600 Interleukin-10 limits intense acute swimming-induced muscle mechanical hyperalgesia in mice.
3601 *Exp Physiol* 2015.
- 3602 70. Asghar M, George L, Lokhandwala MF. Exercise decreases oxidative stress and inflammation
3603 and restores renal dopamine D1 receptor function in old rats. *Am J Physiol Renal Physiol* 2007;
3604 293:F914-9.
- 3605 71. Woods CG, Fu J, Xue P, Hou Y, Pluta LJ, Yang L, et al. Dose-dependent transitions in Nrf2-
3606 mediated adaptive response and related stress responses to hypochlorous acid in mouse
3607 macrophages. *Toxicol Appl Pharmacol* 2009; 238:27-36.
- 3608 72. Lee TS, Chau LY. Heme oxygenase-1 mediates the anti-inflammatory effect of interleukin-10 in
3609 mice. *Nat Med* 2002; 8:240-6.

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3621 **Figures**
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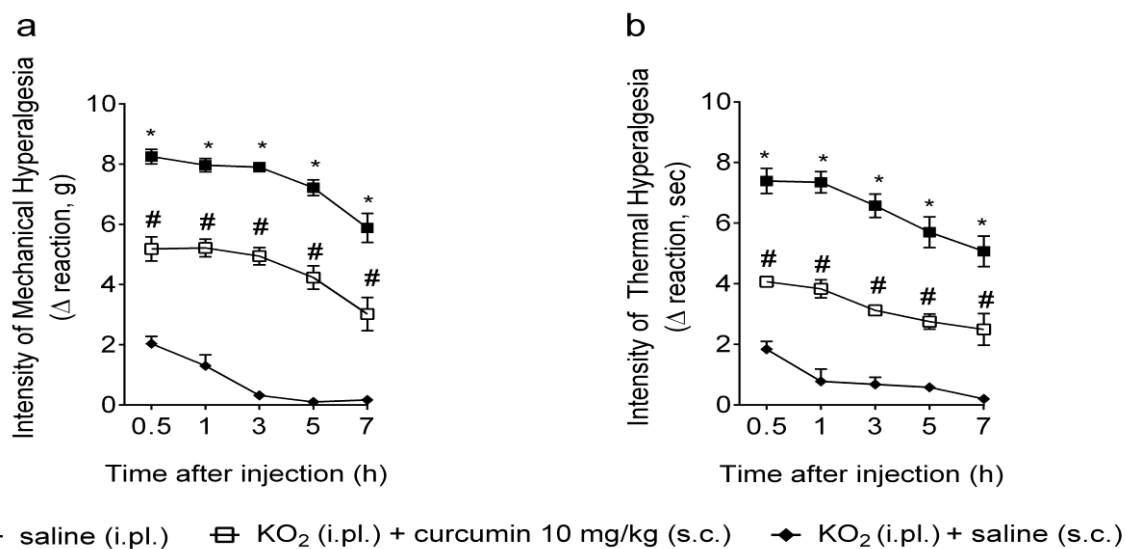
3623

3624 **Fig. 1 Curcumin inhibits KO₂-induced overt pain-like behaviors**

3625 Mice were treated at doses 3, 10 and 30 mg/kg (s.c.) 1 h before i.pl. injection of 30 μ g KO₂ or i.p.
3626 injection of 1 mg KO₂. Zero mg/kg of curcumin stands for vehicle group. Total number of writting (a)
3627 was evaluated 0-20 min after i.p. injection of KO₂. The number of flinches (b) and time spent licking
3628 (c) the paw were evaluated 0- 30 min after i.pl. injection of KO₂. Results are expressed as mean \pm
3629 SEM (n=6 per group per experiment, representative of two separate experiments). *p<0.05 vs. saline
3630 group #p<0.05 vs. KO₂ group. (one-way ANOVA followed Tukey's *post hoc*).

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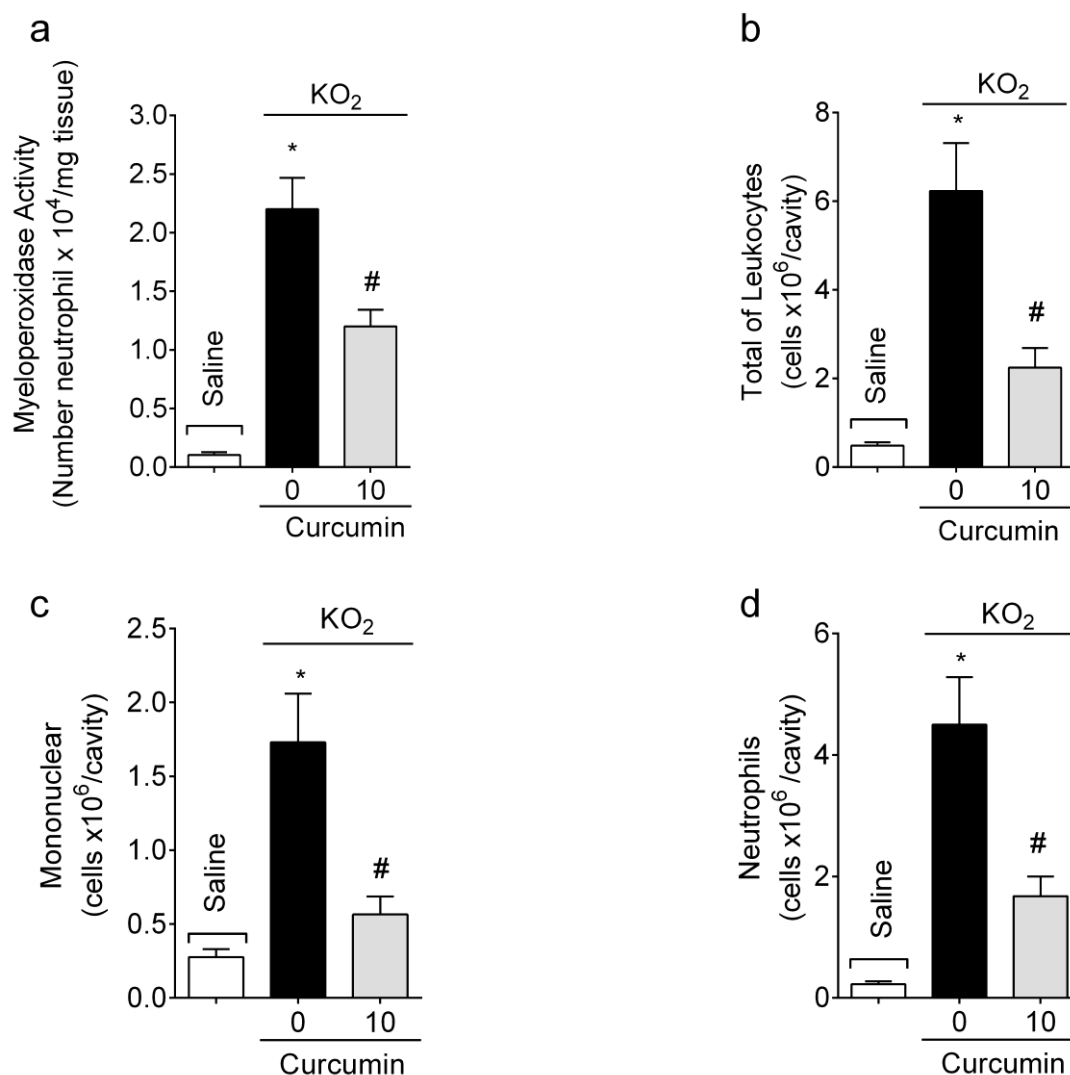
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3634 **Fig. 2 Curcumin inhibits KO₂-induced mechanical and thermal hyperalgesia**

3635 Mice were treated with curcumin 10 mg/kg (s.c.) 1 h before i.pl. injection of 30 μ g KO₂. The test was
3636 performed at 0.5, 1, 3, 5 and 7 h after stimulus for both mechanical (a) and thermal (b) hyperalgesia.
3637 Results are expressed as mean \pm SEM (n=6 per group per experiment, representative of two separate
3638 experiments). *p<0.05 vs. saline group #p<0.05 vs. KO₂ group (two-way repeated measures ANOVA
3639 followed Tukey's *post hoc*).



3640

3641 **Fig. 3 Curcumin inhibits KO₂-induced leukocyte recruitment and myeloperoxidase activity**

3642 Mice were pretreated with curcumin 10 mg/kg (s.c.) 1 h before i.pl. or i.p. injection of 30 μ g KO₂.

3643 Zero mg/kg of curcumin stands for vehicle group. Samples were collected 7 h after KO₂ injection.

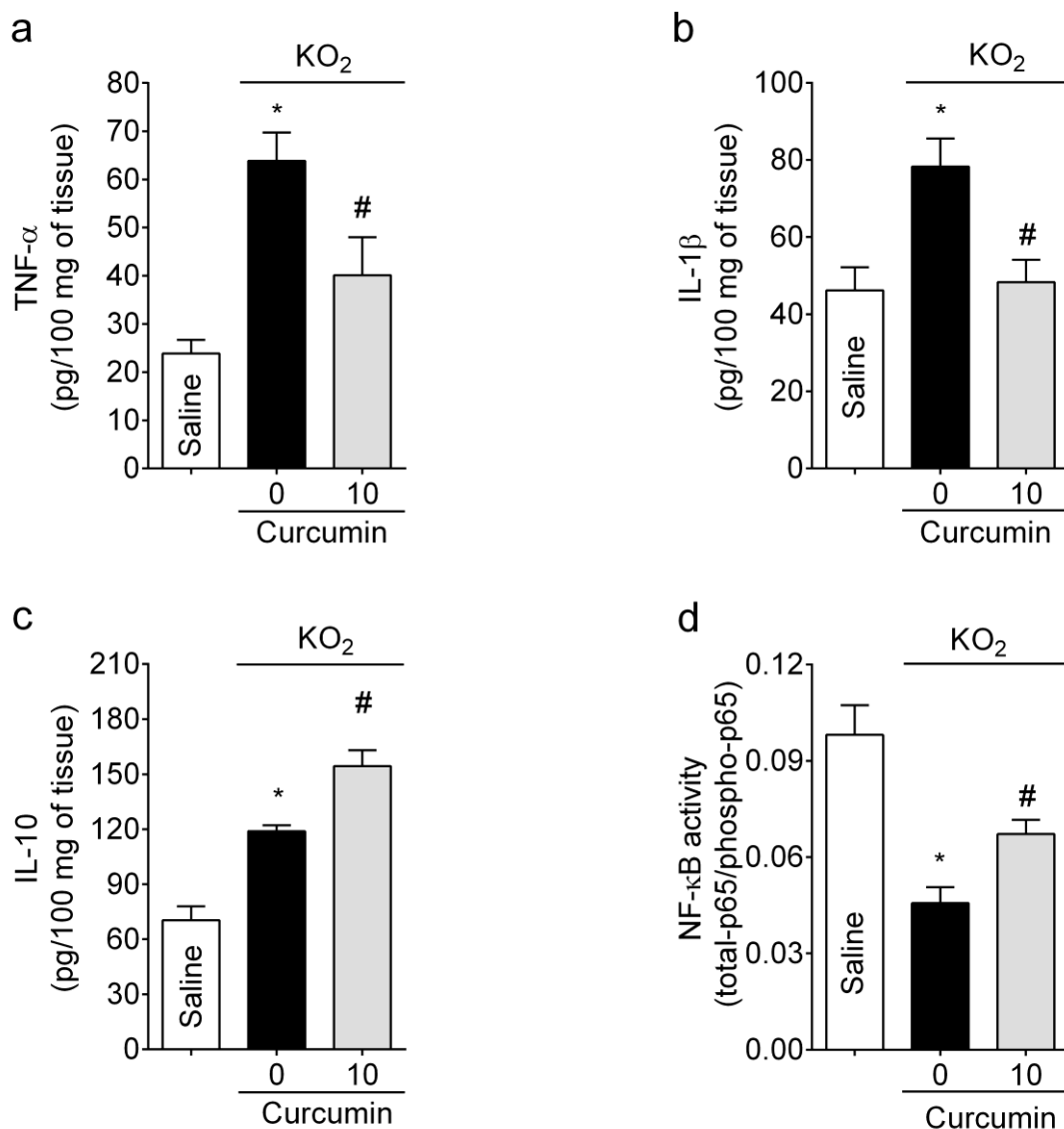
3644 Myeloperoxidase activity was determined in paw skin samples (a). The total number of leukocyte (b),

3645 mononuclear cells (c) and neutrophils (d) were determined in peritoneal washes. Results are expressed

3646 as mean \pm SEM (n=6 per group per experiment, representative of two separate experiments). *p<0.05

3647 vs. saline group #p<0.05 vs. KO₂ group (one-way ANOVA followed Tukey's *post hoc*).

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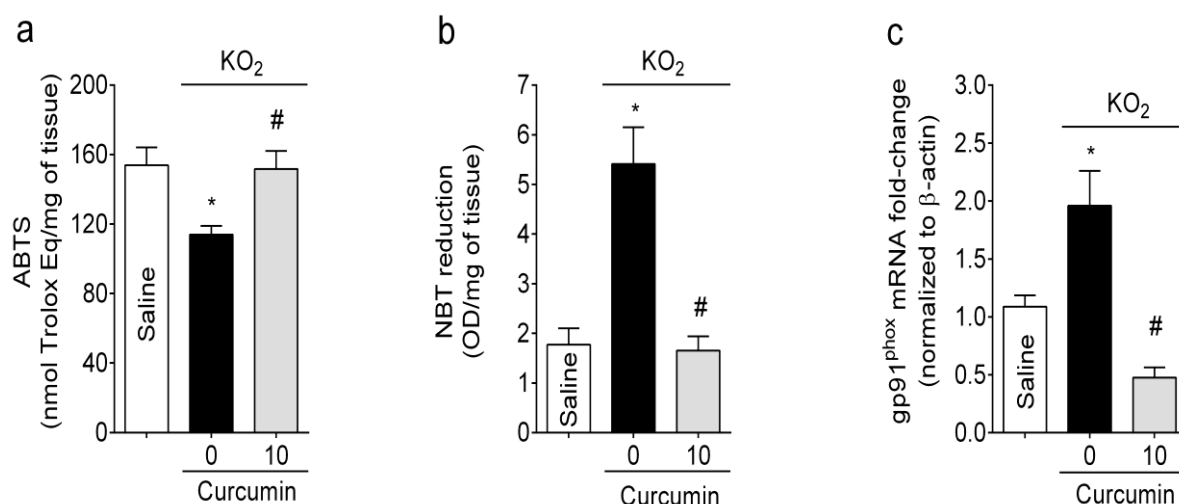


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3650 **Fig. 4 Curcumin inhibits KO₂-induced cytokines production and NF- κ B activation**

3651 Mice were treated with curcumin 10 mg/kg (s.c.) 1 h before i.pl. injection of 30 μ g KO₂. Zero mg/kg
 3652 of curcumin stands for vehicle group. Samples were collected 3 h after stimulus injection to determine
 3653 TNF- α (a), IL-1 β (b), IL-10 (c) levels and NF- κ B activity (d). Results are expressed as mean \pm SEM
 3654 (n=6 per group per experiment, representative of two separate experiments). *p<0.05 vs. saline group
 3655 #p<0.05 vs. KO₂ group (one-way ANOVA followed Tukey's *post hoc*).
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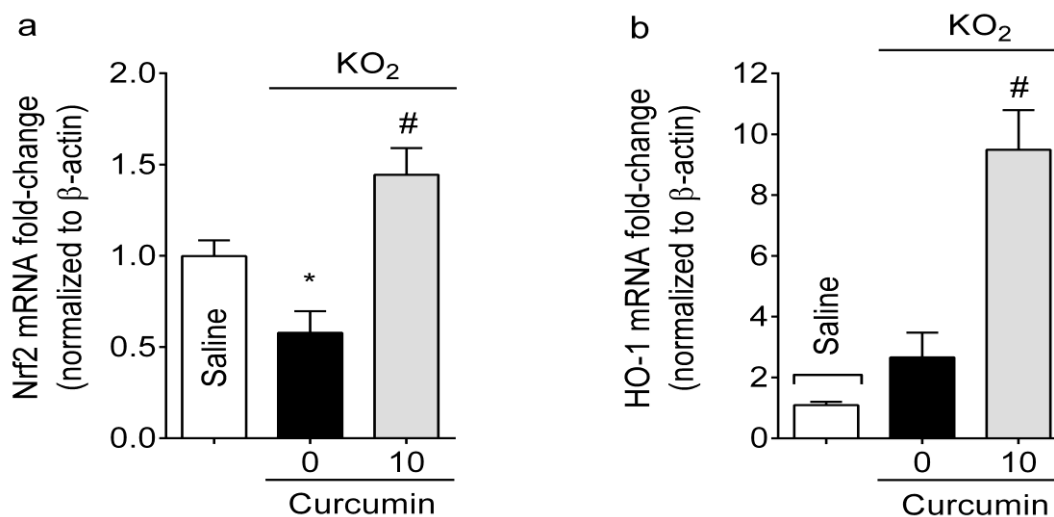


3658

3659 **Fig. 5 Curcumin inhibits KO₂-induced oxidative stress**

3660 Mice were treated with curcumin 10 mg/kg (s.c.) 1 h before i.pl. injection of 30 μg KO₂. Zero mg/kg
 3661 of curcumin stands for vehicle group. Sample were collected 3 h after stimulus to determine total
 3662 antioxidant capacity (ABTS assay) (a), O₂^{•-} production (NBT assay) (b) and gp91^{phox} mRNA
 3663 expression (c). β-actin was used as a reference gene to normalize data mRNA expression. Results are
 3664 expressed as mean ± SEM (n=6 per group per experiment, representative of two separate
 3665 experiments). *p<0.05 vs. saline group #p<0.05 vs. KO₂ group (one-way ANOVA followed Tukey's
 3666 *post hoc*).

3667



3668

3669 **Fig. 6 Curcumin increases KO₂-induced Nrf2 and HO-1 mRNA expression**

3670 Mice were treated with curcumin 10 mg/kg (s.c.) 1 h before i.pl. injection of 30 μg KO₂. Zero mg/kg
 3671 of curcumin stands for vehicle group. Sample were collected 3 h after stimulus to determine Nrf2 (a)
 3672 and HO-1 (b) mRNA expression. β-actin was used as a reference gene to normalize data mRNA
 3673 expression. Results are expressed as mean ± SEM (n=6 per group per experiment, representative of
 3674 two separate experiments). *p<0.05 vs. saline group #p<0.05 vs. KO₂ group (ANOVA followed
 3675 Tukey's *post hoc*).

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3678 **6 CONCLUSÃO**

3679

3680 O modelo de artrite induzido por dióxido de titânio e o modelo agudo de
3681 inflamação e dor induzido pelo ânion superóxido foram padronizados pelo nosso
3682 laboratório e se mostram como alternativas viáveis para os modelos pré-existestes
3683 de dor e inflamação. A curcumina, é uma molécula com potencial analgésico
3684 promissor, tendo em vista que atua em múltiplas vias de sinalização possuindo mais
3685 de 100 diferentes alvos moleculares, entre eles: inibição do NF-κB e citocinas
3686 *downstream*, inibição da sinalização entre RANK-RANKL, aumento da atividade do
3687 Nrf2 e HO-1, inibição do recrutamento de neutrófilos etc; além de possuir baixa
3688 toxicidade. Sua efiácia é confirmada com o desenvolvimento de diversas moléculas
3689 análogas e principalmente de um medicamento tendo a curcumina como princípio
3690 ativo (Meriva[®]), o qual já utilizado em diversos ensaios clínicos com pacientes com
3691 câncer de mama, osteoartrite, psoríase, dor muscular de início tardio (DOMS) entre
3692 outras.

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