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ESTADUAL DE LONDRINA

WALISON AUGUSTO DA SILVA BRITO

**ANÁLISE DO ENVOLVIMENTO DE MARCADORES
SISTÊMICOS DE ESTRESSE OXIDATIVO E DE PROCESSO
INFLAMATÓRIO NO DESENVOLVIMENTO DE MÚLTIPLOS
TIPOS DE CÂNCER DE PELE NÃO-MELANOMA**

Londrina
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Dissertação apresentada ao Programa de Pós-Graduação em Patologia Experimental da Universidade Estadual de Londrina, como requisito parcial para obtenção do título de mestre em Patologia Experimental.

Orientadora: Profa. Dra. Alessandra Lourenço Cecchini Armani

Co-orientadora: Profa. Dra. Poliana Camila Marinello

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WALISON AUGUSTO DA SILVA BRITO

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BANCA EXAMINADORA

Profa. Dra. Alessandra Lourenço Cecchini
Armani
Universidade Estadual de Londrina - UEL

Profa. Dra. Andréa Name Colado Simão
Universidade Estadual de Londrina - UEL

Profa. Dra. Karen Brajão de Oliveira
Universidade Estadual de Londrina - UEL

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*“Aprender é a única coisa que a mente
não se cansa, nunca tem medo e nunca
se arrepende”*

(DA VINCI, Leonardo).

BRITO, Walison Augusto da Silva. **Análise do envolvimento de marcadores sistêmicos de estresse oxidativo e de processo inflamatório no desenvolvimento de múltiplos tipos de câncer de pele não-melanoma.** 2020. 66 f. Dissertação (Mestrado em Patologia Experimental) – Universidade Estadual de Londrina, Londrina, 2020.

RESUMO

O câncer de pele não-melanoma (CPNM) é a neoplasia mais incidente no mundo. O tipo mais comum é o carcinoma basocelular (CBC), seguido pelo carcinoma espinocelular (CEC), que por sua vez, pode originar-se de uma lesão pré-neoplásica denominada queratose actínica (QA). Sabe-se que o estresse oxidativo (EO) tem um papel importante na carcinogênese da pele, que ocorre principalmente devido à radiação ultravioleta, que também é capaz de induzir processo inflamatório, ambos relacionados com a iniciação, promoção e a progressão tumoral. Entretanto, não está claro se marcadores sistêmicos de EO e do processo inflamatório estão associados com a presença de único e múltiplos tipos de CPNM. Assim, este trabalho tem como objetivo investigar marcadores sistêmicos de EO e do processo inflamatório em pacientes com QA e CPNM que poderiam estar associados com a presença de múltiplos tipos de CPNM em um mesmo paciente. Participantes foram categorizados em 6 grupos: Controle (sem câncer, n=75), CBC (n=90), CEC (n=24), QA (n=19), CEC+CBC (n=13) e CEC+CBC+QA (n=36) (CEP-UEL n. processo 1.077.557 e n. 3.146.725). Três comparações foram feitas entre os grupos: 1) Lesões de CPNM de único tipo: controle vs CBC vs CEC; 2) Diferença entre lesão pré-maligna e CEC: controle vs QA vs CEC; 3) Lesões de múltiplos tipos de CPNM: CBC vs CEC vs CEC+CBC vs CEC+CBC+QA. Amostras de sangue foram coletadas, processadas e armazenadas para posteriores análises. Dados qualitativos foram analisados pelo teste de Qui-quadrado e pelo teste z para comparar proporções. Dados quantitativos foram analisados pelo ANOVA (one-way) ou Kruskal-Wallis depois da verificação da normalidade e homogeneidade dos dados. Todos os resultados com $p < 0,1$ na análise univariada foram incluídos no modelo multinomial de regressão logística. $p < 0,05$ foi considerado significativo. Na comparação 1, os pacientes com CBC e CEC apresentaram níveis menores de EO sistêmicos comparados ao controle, observado na redução da razão glutatona oxidada e reduzida ($p=0,012$ e $p=0,025$, respectivamente) e na redução da lipoperoxidação ($p=0,004$ e $p=0,002$, respectivamente). Na comparação 2, somente redução na lipoperoxidação dos pacientes com CEC comparados ao grupo controle ($p=0,037$) foi observada. Nenhum dos parâmetros avaliados diferiu entre a QA e o CEC. Na comparação 3, foi possível observar que a atividade da gama-glutamil transpeptidase (GGT) é diferente nos pacientes com um único tipo de CPNM, e também que os pacientes com ambos os tipos de CPNM (CEC+CBC), apresentaram a maior atividade da enzima. Entretanto, essa diferença é perdida quando se considera a localização da lesão. Em conclusão, pacientes com CBC e CEC apresentam baixos níveis sistêmicos de EO; a atividade da GGT sistêmica é o único parâmetro de EO, dentre os avaliados, que difere entre os dois tipos de CPNM, além de estar associada com o desenvolvimento dos dois tipos de CPNM em um mesmo paciente, entretanto, a localização do tumor interfere nessas diferenças; nenhum dos marcadores de estresse oxidativo e inflamação analisados difere sistemicamente entre quem possui a lesão pré-maligna e maligna;

os parâmetros de inflamação avaliados não diferem sistemicamente nos pacientes, independente de possuírem um ou mais tipos de CPNM.

Palavras-chave: Gama-glutamil transpeptidase. Peroxidação lipídica. Defesa antioxidante. Carcinoma basocelular. Carcinoma espinocelular.

BRITO, Walison Augusto da Silva. **Analysis of the involvement of systemic markers of oxidative stress and inflammatory process in the development of multiple types of nonmelanoma skin cancer.** 2020. 66 p. Dissertação (Mestrado em Patologia Experimental) – Universidade Estadual de Londrina, Londrina, 2020.

ABSTRACT

Nonmelanoma skin cancer (NMSC) is the most incident neoplasm in the world. The most common type is basal cell carcinoma (BCC), followed by squamous cell carcinoma (SCC), which in turn may originate from a preneoplastic lesion called actinic keratosis (AK). It is known that oxidative stress (OS) plays an important role in skin carcinogenesis, which occurs mainly due to ultraviolet radiation, which is also capable of inducing inflammatory process, both related to tumor initiation, promotion and progression. However, it is not clear if systemic markers of OS and inflammatory process are associated with the presence of single and multiple NMSC. Thus, this work aims to investigate systemic markers of OS and inflammatory process in patients with AK and NMSC that could be associated to the presence of multiple types of NMSC. Participants were categorized in 6 groups: Control (without skin cancer, n=75), BCC (n=90), SCC (n=24), AK (n=19), SCC+BCC (n=13) and SCC+BCC+AK (n=36) (CEP-UEL process n. 1.077.557 and n. 3.146.725). Three comparisons were made between the groups: 1) Single type of NMSC lesions: control vs BCC vs SCC; 2) Difference between preneoplastic lesion and SCC: control vs AK vs SCC; 3) Multiple types of NMSC lesions: BCC vs SCC vs SCC+BCC vs SCC+BCC+AK. Blood samples were collected, processed and stored for further analysis. Qualitative data were analyzed by Chi-square test z test for proportion comparison. Quantitative data were analyzed by ANOVA (one-way) or Kruskal-Wallis after the verification of data normality and homogeneity. All results with $p < 0.1$ in the univariate analysis were included in the multinomial logistic regression model. $p < 0.05$ was considered significant. In comparison 1, patients with BCC and SCC presented lower levels of systemic OS compared to control, observed in the reduction of the oxidized and reduced glutathione ratio ($p = 0.012$ and $p = 0.025$, respectively) and in the reduction of lipoperoxidation ($p = 0.004$ and $p = 0.002$, respectively). In comparison 2, only reduction in lipoperoxidation of patients with CPB compared to the control group ($p = 0.037$) was observed. None of the parameters evaluated differed between QA and SCC. In comparison 3, it was possible to observe that the activity of gamma-glutamyl transpeptidase (GGT) is different in patients with a single type of NMSC, and also that patients with both types of NMSC (SCC+BCC) had the highest enzyme activity. However, this difference is lost when considering the location of the lesion. In conclusion, patients with BCC and SCC have low systemic levels of OS; systemic GGT activity is the only OS parameter among those evaluated, which differs between the two types of NMSC, besides being associated with the development of the two types of NMSC in the same patient, however, the location of the tumor interferes with these differences; none of the markers of oxidative stress and inflammation analyzed differs systemically between those who have the premalignant and malignant lesion; the evaluated inflammation parameters do not differ systemically in patients, regardless of whether they have one or more types of NMSC.

Keywords: Gamma-glutamyl transpeptidase. Lipid peroxidation. Antioxidant defenses. Basal cell carcinoma Squamous cell carcinoma.

LISTA DE ABREVIATURAS E SIGLAS

| | |
|----------------|---|
| ANOVA | Análise de variância |
| AOPP | Produtos avançados da oxidação de proteínas (Advanced Oxidation Protein Products) |
| CBC | Carcinoma de células basais ou basocelular |
| CEC | Carcinoma de células escamosas ou espinocelular |
| CPNM | Câncer de pele não-melanoma |
| EO | Estresse oxidativo |
| ERO | Espécies reativas de oxigênio |
| GGT | Gama-glutamil transpeptidase |
| GSH | Glutathiona reduzida |
| GSSG | Glutathiona oxidada |
| H&E | Hematoxilina e Eosina |
| HPV | Vírus do papiloma humano (Human papiloma virus) |
| IL-10 | Interleucina-10 |
| INCA | Instituto Nacional do Câncer |
| MDA | Malondialdeído |
| QA | Queratose actínica |
| QL | Quimiluminescência |
| SOD | Superóxido dismutase |
| TGF- β 1 | Fator transformador do crescimento beta 1 (Transforming growth factor beta 1) |
| TNF- α | Fator de necrose tumoral alfa (Tumor necrosis factor alpha) |
| UV | Ultravioleta |

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1 INTRODUÇÃO

1.1 CÂNCER DE PELE NÃO-MELANOMA

As neoplasias cutâneas podem se desenvolver a partir de células epidérmicas, de melanócitos, de células mesenquimais ou ainda de células hematolinfoides. Dentre as neoplasias cutâneas a mais comum no mundo é a de células epidérmicas. Segundo a Organização Mundial da Saúde, a incidência de cânceres de pele, não melanoma e melanoma, tem aumentado muito nas últimas décadas e isso pode ser relacionado com o aumento da população em envelhecimento, uma vez que este tipo de câncer está intimamente relacionado com a exposição solar ao longo da vida (GLOBAL BURDEN OF DISEASE CANCER COLLABORATION, 2019; WHO, 2019).

No Brasil, o câncer de pele não-melanoma (CPNM) é o mais frequente e corresponde a 30% de todos os tumores malignos registrados no país segundo o Instituto Nacional do Câncer (INCA). Dentre os tumores de pele, o CPNM é o de maior incidência, apresenta baixa mortalidade e altos percentuais de cura se diagnosticado precocemente. O INCA estima para o biênio de 2020-2021, aproximadamente 176 mil novos casos, sendo 83 mil e 93 mil casos abrangendo homens e mulheres, respectivamente (INCA, 2020). O CPNM engloba o carcinoma de células basais (CBC) e o carcinoma de células escamosas (CEC), mesmo possuindo algumas similaridades, esses dois tipos de CPNM possuem diferentes taxas de incidência e diferenças etiológicas (LUCAS et al., 2006). Ambas neoplasias possuem risco de mortalidade e de metástase baixos em comparação com o melanoma, porém, como a cura se dá por remoção cirúrgica, esse tratamento pode causar grande impacto psicológico e social, uma vez que a lesão geralmente está localizada em regiões visíveis como cabeça, pescoço e face, podendo gerar dor e desfiguração (BAILEY et al., 2019; ROTHENBERG; ELLISEN, 2012; WHO, 2019).

Os principais fatores de risco para o desenvolvimento do CPNM são a pigmentação da pele e a exposição à radiação ultravioleta (UV). A radiação UV possui diversos efeitos sobre a pele, como indução de eritema, mutação e imunossupressão, promoção de síntese de vitamina D e bronzeamento, além de possuir efeito carcinogênico, relacionando-se com o desenvolvimento de cerca de 90% dos casos de CPNM (FAJUYIGBE; YOUNG, 2016; KOH et al., 1996). A

1 pigmentação da pele confere fotoproteção, pois a melanina é capaz de absorver uma
 2 banda larga de radiação UV, além de ter propriedades antioxidantes, protegendo a
 3 pele do estresse oxidativo induzido pela radiação (BRENNER; HEARING, 2008). Por
 4 essa razão, a concentração de melanina pode ser inversamente relacionada com a
 5 incidência de cânceres de pele, que acomete mais facilmente indivíduos de pele clara
 6 (Classificação de Fitzpatrick I-III) (Figura 1) do que aqueles de pele escura
 7 (Classificação de Fitzpatrick IV-VI) (Figura 1). Os indivíduos que possuem pele escura
 8 tem maior presença de eumelanina, que é um tipo de melanina cuja propriedade
 9 fotoprotetora é superior ao outro tipo, a feomelanina, que está presente em maior
 10 quantidade nos indivíduos de pele mais clara (BRENNER; HEARING, 2008).



11

12 **Figura 1.** Classificação de Fitzpatrick dos fotótipos de pele. Adaptado de Metro Jornal (2019).
 13 Fonte: [https://www.metrojornal.com.br/estilo-vida/2019/01/12/verao-atencao-pele-cuidados-](https://www.metrojornal.com.br/estilo-vida/2019/01/12/verao-atencao-pele-cuidados-simples.html)
 14 [simples.html](https://www.metrojornal.com.br/estilo-vida/2019/01/12/verao-atencao-pele-cuidados-simples.html)

15 Além da exposição à radiação UV e a pigmentação da pele, outros
 16 fatores de risco também favorecem o desenvolvimento de CPNM, dentre eles a
 17 imunossupressão, associada ou não a transplantes de órgãos, fatores genéticos e
 18 infecção cutânea por algumas cepas do vírus do papiloma humano (HPV) são os mais
 19 importantes (BELBASIS et al., 2016; HOFBAUER; BAVINCK; EUVRARD, 2010;
 20 TESSARI; GIROLOMONI, 2012). O tabagismo é um hábito que está associado com o
 21 desenvolvimento de diferentes tipos de câncer, principalmente CEC em mucosa oral,
 22 além de indivíduos tabagistas com CPNM terem uma maior propensão a desenvolver
 23 outros tipos de câncer extracutâneos, tais como mama, cólon, esôfago, rins, pulmão,
 24 tireoide, entre outros (ROLLISON et al., 2012; SILVERBERG; RATNER, 2015).

1 Estudos apontam que pacientes com histórico prévio de neoplasias
2 cutâneas apresentam maior chance de desenvolver novas lesões, sejam essas do
3 mesmo tipo da precedente ou de tipos diferentes (BELBASIS et al., 2016; DUARTE et
4 al., 2018; PANDEYA; OLSEN; WHITEMAN, 2017). Levando em conta que geralmente
5 apenas a lesão primária é registrada, dados epidemiológicos sobre a existência de
6 múltiplos tipos de CPNM são escassos, assim como existe grande dificuldade em
7 compreender como esse processo ocorre (BELBASIS et al., 2016; DUARTE et al.,
8 2018; PANDEYA; OLSEN; WHITEMAN, 2017). Entretanto, o melhor entendimento do
9 desenvolvimento dessas lesões, como a participação do EO e do processo
10 inflamatório, por exemplo, pode auxiliar na redução das taxas morbidade e
11 mortalidade decorrentes desses tipos de câncer.

12

13 1.2 CARCINOMA DE CÉLULAS BASAIS OU BASOCELULAR (CBC)

14 O CBC é o câncer mais frequentemente encontrado em humanos de
15 pele clara, correspondendo a 75% dos casos de CPNM (BRANDT; MOORE, 2019).
16 Apresenta crescimento lento e as metástases são raras, entretanto, sua taxa de
17 morbidade é relativamente alta devida a sua capacidade de invasão tecidual, podendo
18 lesionar tecidos adjacentes a pele (MARZUKA; BOOK, 2015). A exposição intensa e
19 prolongada aos raios UV do sol está relacionada com o desenvolvimento da doença,
20 substancialmente em indivíduos cuja ocupação se dá sob altas doses de exposição
21 solar (SCHMITT et al., 2017).

22 O CBC é composto por células que se assemelham às células basais
23 da epiderme, que se originam de células epiteliais imaturas da camada basal e mais
24 raramente do complexo cutâneo pilo-sebáceo ou outros apêndices cutâneos. É um
25 tumor que possui características infiltrativas nos tecidos adjacentes por expansões
26 irregulares digitiformes (Figura 2B e 2C) (TÂNȚU et al., 2014). Por meio de suas
27 características histológicas e clínicas, o CBC pode ser dividido nos seguintes subtipos:
28 nodular, superficial, infundibulocísticos, fibroepitelial, esclerodermiforme e infiltrativo,
29 sendo estes dois últimos os mais agressivos e o nodular e superficial os mais comuns
30 (CAMERON et al., 2019; MARZUKA; BOOK, 2015).

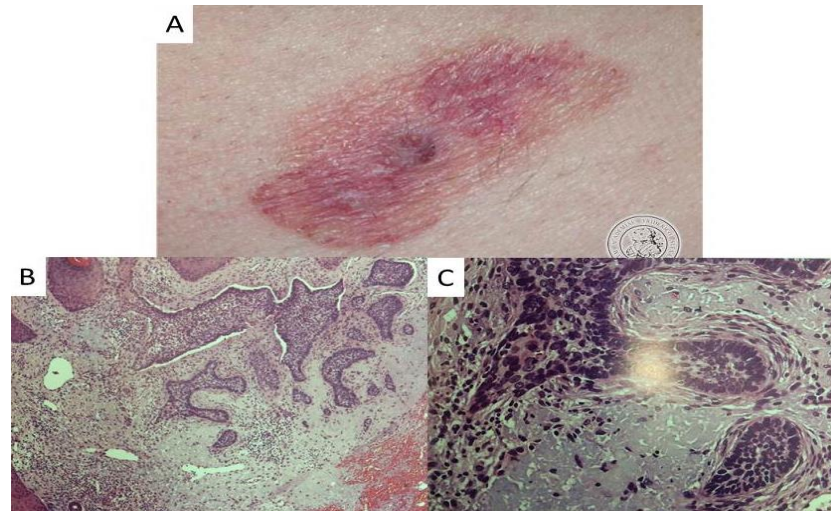


Figura 2. Carcinoma basocelular (CBC). (A) CBC em face. (B) Lâmina de CBC corado por hematoxilina e eosina (H&E) mostrando que o CBC é caracterizado histologicamente pela presença de agregados esféricos ou infiltrantes formados por células neoplásicas em azul-escuro e que se conectam com a epiderme. Artefatos de separação entre os nódulos neoplásicos e o estroma adjacente é um achado que auxilia o diagnóstico. (C) As células nos limites dos agregados neoplásicos se alinham em paliçada. O CBC apresenta um estroma fibromixóide azul peculiar, periférico aos agregados neoplásicos. Esse aspecto pode facilitar a distinção entre os nódulos neoplásicos do CBC e folículos pilosos inocentes. Fonte: Atlas de Histopatologia - Dr. Ivan Damjanov.

1.3 CARCINOMA DE CÉLULAS ESCAMOSAS OU ESPINOCELULAR (CEC)

O CEC é o segundo tipo mais frequente de CPNM, detendo aproximadamente 25% dos casos, surgindo de queratinócitos displásicos na camada espinhosa da epiderme (Figura 3) (BRANDT; MOORE, 2019).

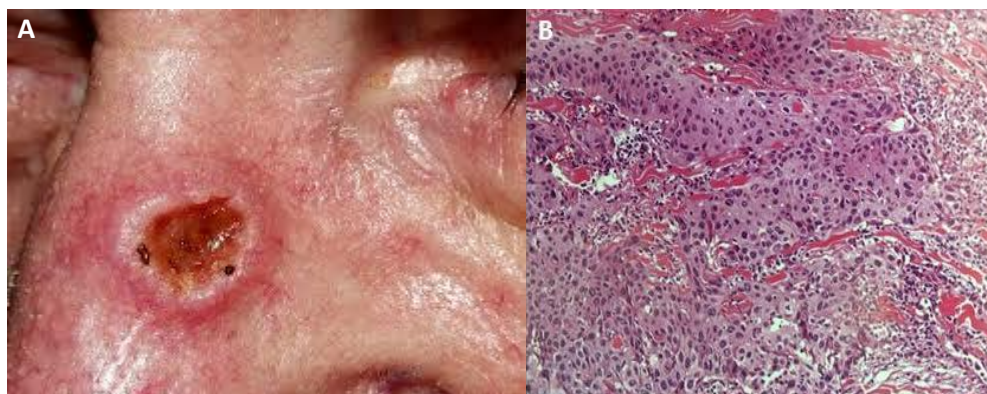


Figura 3. Carcinoma espino celular em paciente (A) e histopatologia em H&E (B). (A) Paciente com diagnóstico de carcinoma espino celular antes da remoção cirúrgica. Fonte: Dr. Julio Bergmann. Disponível em: <https://clinicabergmann.com.br/tratamento/fotos-cancer-de-pele-espino-celular-no-nariz-labio-orelhas-e-pernas-em-porto-alegre/>. (B) O tumor é composto de células com núcleos grandes e pleomórficos, organizadas em agregados com formas variadas e que invadem a derme. Fonte: Atlas de Histopatologia - Dr. Ivan Damjanov.

1 A apresentação clínica mais frequente de CEC é a presença de placas
2 bem diferenciadas, queratinocíticas e eritematosas. Pode ser endurecido,
3 apresentando pápulas firmes, placas ou nódulos com hiperqueratose e ulceração
4 (lesões bem diferenciadas), podendo também apresentar pápulas granulomatosas,
5 carnudas e macias ou nódulos sem queratinização (lesões pouco diferenciadas). O
6 CEC acomete principalmente as regiões do corpo mais expostas aos raios UV, como
7 a face, cabeça, pescoço, antebraços, dorso das mãos e a parte superior do tórax
8 (WARSZAWIK-HENDZEL et al., 2015).

9 A exposição crônica à radiação UV do sol com subsequente dano no
10 DNA é o mais importante fator que predispõe o desenvolvimento do CEC. Para além
11 deste, a idade avançada e o fotótipo de pele claro (Classificação de Fitzpatrick I-III)
12 também estão associados (BAILEY et al., 2019; FARTASCH et al., 2012; SCHMITT;
13 BORDEAUX, 2013). A população de maior risco inclui trabalhadores com exposição
14 ocupacional à luz ultravioleta (FARTASCH et al., 2012; ROTHENBERG; ELLISEN,
15 2012). Dentre as lesões precursoras clinicamente associadas ao CEC, inclui a
16 Queratose Actínica (QA), uma lesão pré-maligna, com alto potencial de progressão
17 (WARSZAWIK-HENDZEL et al., 2015).

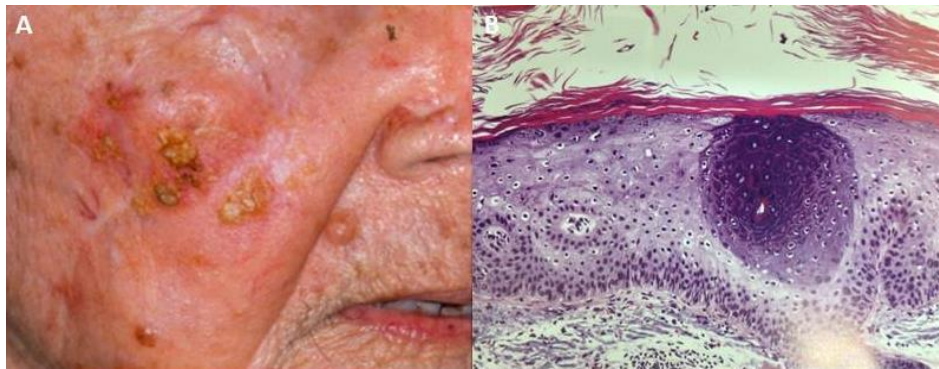
18

19 1.4 QUERATOSE ACTÍNICA (QA)

20 A QA é uma lesão na pele formada pela proliferação de queratinócitos
21 displásicos em áreas com maior foto-exposição, comum em indivíduos adultos e
22 idosos de pele clara, resultante de um longo período de exposição aos raios UV, tendo
23 esse último como o principal mecanismo pela indução de inflamação, estresse
24 oxidativo, imunossupressão, desregulação do ciclo celular e apoptose, além de
25 proliferação celular e remodelamento tecidual (REINEHR; BAKOS, 2019; SIEGEL;
26 KORGAVKAR; WEINSTOCK, 2017). Os principais fatores de risco para o
27 desenvolvimento da QA incluem a exposição cumulativa à radiação solar, idade
28 avançada, fotótipo de pele clara (Classificação de Fitzpatrick I-III), histórico de
29 queimaduras na infância e habitar em regiões de baixas latitudes (regiões onde há
30 maior incidência dos raios solares) (SCHMITT; BORDEAUX, 2013; SCHMITT; MIOT,
31 2012).

32 A QA é uma lesão eritematosa que comumente aparece como
33 múltiplas lesões (Figura 4), além de ser considerada uma lesão pré-cancerosa (tem a

1 capacidade de progredir para um carcinoma espinocelular, que depende do grau de
 2 displasia), que apresenta expressões fenotípicas de fotoenvelhecimento cutâneo,
 3 juntamente com perda de elasticidade da pele, atrofia e mudanças na pigmentação
 4 (SCHMITT; BORDEAUX, 2013; SCHMITT; MIOT, 2012; WARSZAWIK-HENDZEL et
 5 al., 2015). Histologicamente, a QA pode apresentar hiperqueratose que geralmente
 6 contém áreas de paraqueratose, indicando queratinização anormal e infiltrado
 7 linfocítico, que pode ter um papel na progressão da doença (FERNANDEZ
 8 FIGUERAS, 2017).



9

10 **Figura 4** - Queratose actínica. A: Queratose actínica com característica elevada,
 11 esbranquiçada, áspera e escamosa em face de paciente. B: Histopatologia em H&E, a
 12 epiderme comprometida se queratiniza anormalmente com paraqueratose. Aumento de 100X.
 13 Fonte: Atlas de Histopatologia - Dr. Ivan Damjanov, 2013.

14

15 1.5 CÂNCER DE PELE NÃO-MELANOMA E MARCADORES SISTÊMICOS DE ESTRESSE OXIDATIVO 16 E PROCESSO INFLAMATÓRIO

17 Sabe-se que a radiação UV é um carcinógeno ambiental que atua
 18 como iniciador e promotor do processo de carcinogênese, através da lesão direta ao
 19 DNA formando dímeros de pirimidina, causando imunossupressão, inflamação e
 20 indução de estresse oxidativo (CARR et al., 2012; DAMIANI; ULLRICH, 2016;
 21 WILLIAMS et al., 2014).

22 O espectro da radiação UV é subdividido em três bandas: UVA (320-
 23 400 nm), UVB (290-320 nm) e UVC (100-280 nm). A UVC é absorvida pela camada
 24 de ozônio, a radiação UVA penetra profundamente até a derme, enquanto os raios da
 25 UVB são retidos na epiderme da pele. Sabe-se que ambos os espectros UVA e UVB
 26 levam ao dano oxidativo, uma vez que são capazes de induzir e gerar espécies
 27 reativas de oxigênio (ERO) (WILLIAMS et al., 2014). Entretanto, os mecanismos da

1 fotocarcinogênese do CPNM induzida por estresse fotooxidativo são complexos, a
2 constante e intensa exposição à radiação UV leva à produção elevada de ERO pelos
3 queratinócitos com consequente redução nas defesas antioxidantes, caracterizando o
4 EO (FEEHAN; SHANTZ, 2016). As ERO rapidamente interagem com as
5 macromoléculas como lipídeos de membrana, proteínas e DNA, levando ao dano
6 oxidativo (FEEHAN; SHANTZ, 2016). Em adição a isso, os danos ao DNA em sua
7 maioria não são reparados, favorecendo a propagação de mutações em genes
8 específicos, levando a alterações em vias de sinalização relacionadas a proliferação
9 celular e apoptose, favorecendo o desenvolvimento de câncer (FEEHAN; SHANTZ,
10 2016; SCHEUER, 2017).

11 Foi demonstrado em estudos anteriores algumas alterações em
12 marcadores sistêmicos de EO no CPNM. Foi demonstrado uma diminuição
13 significativa nos antioxidantes plasmáticos, como o α -tocoferol e ácido ascórbico, e
14 glutathiona nos eritrócitos de pacientes com QA e CBC (VURAL; CANBAZ; SEKÇUKI,
15 1999), e aumento sérico de um subproduto da oxidação lipídica, o malondialdeído
16 (MDA), em pacientes com CBC quando comparado a indivíduos controle (MAJIDI;
17 DJALALI; JAVANBAKHT, 2017), demonstrando que alterações redox sistêmicas
18 também estão presentes no CPNM. Entretanto, alguns marcadores sistêmicos de
19 estresse oxidativo são pouco investigados tanto na queratose actínica quanto em
20 CPNM, sejam essas lesões de um único tipo ou lesões de múltiplos tipos.

21 A resposta inflamatória é um componente que está muito relacionado
22 com a carcinogênese e progressão tumoral. Isso pode ser devido à geração de
23 espécies reativas e fatores de crescimento pelo ambiente inflamatório, que contribui
24 com as alterações que culminam na carcinogênese (FEKECS et al., 2010; OLTEANU
25 et al., 2012). A radiação UV é capaz de induzir resposta inflamatória e
26 imunossupressão na pele, efeitos caracterizados pela indução da produção de
27 mediadores inflamatórios e citocinas, alteração das respostas vasculares e de
28 moléculas de adesão, além de recrutamento de células inflamatórias, criando um
29 microambiente favorável para o tumor (FEEHAN; SHANTZ, 2016). A perda da
30 capacidade apresentadora de antígenos pode contribuir com a dificuldade do sistema
31 imunológico em eliminar as células cancerosas, e também favorece a carcinogênese
32 na pele e sua possível progressão (FEEHAN; SHANTZ, 2016). Por conseguinte, a
33 determinação de marcadores sistêmicos do processo inflamatório em pacientes com

1 CPNM, seja de um único tipo ou de múltiplos tipos é muito importante, uma vez que
2 pode auxiliar na compreensão da resposta do organismo frente ao tumor.

3 Sabe-se que o estresse oxidativo é capaz de interferir em diferentes
4 vias de sinalização intracelular e algumas delas podem relacionar-se com o
5 desenvolvimento de neoplasias. Uma das vias que o estresse oxidativo pode alterar é
6 a do fator transformador do crescimento beta 1 (TGF- β 1), sendo o estresse oxidativo
7 capaz de aumentar a sua expressão e ativação (KRSTIĆ et al., 2015). O TGF- β 1 é
8 um potente supressor tumoral em células pré-malignas, em contrapartida, um
9 favorecedor da progressão e metástase em células de tumores mais avançados
10 (MASSAGUÉ, 2012). O TGF- β 1 é capaz de induzir a produção de ERO, regular sua
11 atividade, não apenas na produção, mas também na modulação de antioxidantes
12 enzimáticos, como a catalase e a SOD, e também não enzimáticos, como a GSH,
13 através da inibição da expressão de genes importantes para sua formação (KRSTIĆ
14 et al., 2015). Entretanto ainda não foi investigado a concentração sistêmica dessa
15 citocina no CPNM, e se essa citocina a nível sistêmico exerce um papel importante no
16 desenvolvimento de mais de um tipo de CPNM nos pacientes.

17 Dessa forma, sabe-se que tanto o EO quanto o processo inflamatório
18 exercem papéis importantes no desenvolvimento do CPNM, e estudos prévios
19 sugerem algumas particularidades com relação ao EO nos diferentes tipos de CPNM.
20 Além disso, sabendo-se que tanto o EO quanto o processo inflamatório são capazes
21 de interferir na ativação de vias sinalizadoras críticas para o desenvolvimento e
22 progressão tumoral, a hipótese desse trabalho é que existem diferenças no perfil
23 oxidativo e inflamatório sistêmico nos pacientes com queratose actínica e com CPNM,
24 que possam estar associados com a presença de mais de um tipo de CPNM nos
25 pacientes.

1 2 OBJETIVOS

2

3 2.1 OBJETIVO GERAL

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5 Avaliar marcadores sistêmicos de estresse oxidativo e de processo
6 inflamatório em pacientes com queratose actínica e com câncer de pele não-
7 melanoma e sua associação a presença de múltiplos tipos de câncer de pele não-
8 melanoma em um mesmo paciente.

9

10 2.2 OBJETIVOS ESPECÍFICOS

11

- 12 • Avaliar marcadores de EO sistêmico em pacientes com queratose actínica e
13 com um ou mais tipos de CPNM;
- 14 • Quantificar parâmetros inflamatórios no sangue de pacientes com queratose
15 actínica e com um ou mais tipos de CPNM;
- 16 • Investigar se parâmetros sistêmicos de EO e de processo inflamatório são
17 capazes de diferenciar os dois tipos de CPNM, controlando os fatores
18 confundidores e fatores de risco que podem influenciar os resultados obtidos;
- 19 • Analisar se os marcadores sistêmicos de EO e do processo inflamatório são
20 capazes de diferenciar a lesão pré-cancerosa do câncer estabelecido, fazendo
21 o controle das variáveis confundidoras e fatores de risco;
- 22 • Avaliar se os marcadores de EO e do processo inflamatório sistêmicos estão
23 associados com a presença de múltiplos tipos de CPNM em um mesmo
24 paciente, controlando os fatores confundidores e fatores de risco que possam
25 estar influenciando nos resultados obtidos.

1 **3 ARTIGO PARA PUBLICAÇÃO**

2 O presente trabalho, realizado na Universidade Estadual de Londrina,
3 nos laboratórios de Patologia Molecular, de Fisiopatologia e Radicais Livres, de
4 Pesquisa em Imunologia Aplicada, e no Ambulatório de Dermatologia do Hospital
5 Universitário da UEL e no Hospital Norte Paranaense de Araçongas, originou um
6 artigo científico.

7 O artigo será submetido para publicação em revista internacional a ser definida e
8 intitula-se “Evaluation of systemic parameters of oxidative stress and inflammation in
9 patients with single and multiple types of nonmelanoma skin cancer”.

1 **SYSTEMIC PARAMETERS OF OXIDATIVE STRESS AND INFLAMMATION IN**
2 **SINGLE AND MULTIPLE TYPES OF NONMELANOMA SKIN CANCER**

3 Walison Augusto da Silva Brito ^a, Poliana Camila Marinello ^a, Natália Medeiros Dias
4 Lopes ^a, Larissa Juliani Sanches ^a, André Armani ^b, Airton dos Santos Gon ^c, Andréa
5 Name Colado Simão ^d, Rodrigo Cabral Luiz ^a, Rubens Cecchini ^e, Alessandra
6 Lourenço Cecchini ^a.

7

8 ^a Laboratory of Molecular Pathology, State University of Londrina, Londrina, PR, Brazil.

9 ^b Head and Neck Surgeon, Department of Surgical Clinic, State University of Londrina,
10 Londrina, PR, Brazil.

11 ^c Dermatologist, Department of Internal Medicine, State University of Londrina,
12 Londrina, PR, Brazil.

13 ^d Laboratory of Research in Applied Immunology, State University of Londrina,
14 Londrina, PR, Brazil.

15 ^e Laboratory of Physiopathology and Free Radicals, State University of Londrina,
16 Londrina, PR, Brazil.

17

18 Corresponding author: Alessandra Lourenço Cecchini Armani

19 E-mail: alcecchini@uel.br

20 Fax: +55 43 3371 42 67

21 Phone: +55 43 3371 45 29

22 Laboratório de Patologia Molecular, Universidade Estadual de Londrina, Rodovia
23 Celso Garcia Cid, PR445, km 380, Campus Universitário, Londrina, CEP 86051-990,
24 Paraná, Brasil.

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1 Abstract

2 The research purpose is to investigate the systemic parameters of oxidative stress
3 (OS) and inflammatory process in patients with actinic keratosis (AK) and
4 nonmelanoma skin cancer (NMSC) and if those parameters differ between groups of
5 patients with multiple types of NMSC and with pre-malign and malign lesions. Basal
6 cell carcinoma (BCC; n=90), Squamous cell carcinoma (SCC; n=24), AK (n=19),
7 SCC+BCC (n=13) and SCC+BCC+AK (n=36) groups were analyzed in which blood
8 samples were collected immediately before surgery to resect skin lesions. From
9 controls (without cancer, n=75) only blood was collected. Reduced (GSH) and oxidized
10 glutathione (GSSG), catalase, superoxide dismutase, lipid hydroperoxides (CL-
11 LOOH), total thiols, malondialdehyde, advanced oxidized protein products, TNF- α , IL-
12 10, TGF- β 1, ferritin, C-reactive protein and gamma-glutamyl transpeptidase (GGT)
13 were evaluated. BCC and SCC presented reduced levels of GSSG/GSH ratio and CL-
14 LOOH compared to control. GGT activity was lower in BCC than in SCC and, the higher
15 activities were found in patients from BCC+SCC. GGT results lost statistical difference
16 after control by confounding factors. Remaining evaluated parameters of OS and
17 inflammation did not alter. In conclusion, single type NMSC patients present low levels
18 of systemic OS (independently associated with NMSC); GGT activity is different in
19 patients with single and multiple types of NMSC. However, GGT is influenced by
20 confounding factors. Besides, others OS and inflammatory markers used are not good
21 parameters to differ systemically between the types of NMSC, nor in patients with
22 single and multiple types of NMSC, as well as none evaluated parameter differed
23 between pre-malign and malign lesions.

24 **Keywords:** Gamma-glutamyl transpeptidase. Lipid peroxidation. Antioxidant
25 defenses. Basal cell carcinoma. Squamous cell carcinoma.

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1 Introduction

2 Nonmelanoma skin cancer (NMSC) is the most incident cancer
3 worldwide. This incidence is constantly raising due to the increase in population age
4 and sun exposure throughout life (GLOBAL BURDEN OF DISEASE CANCER
5 COLLABORATION, 2019; WHO, 2019). NMSC is a group of cutaneous malignancies
6 that mainly comprises keratinocytes carcinomas: basal cell carcinoma (BCC),
7 corresponding to approximately 75% of cases, and squamous cell carcinoma (SCC),
8 25% of the remaining cases (BAILEY et al., 2019). NMSC presents low mortality rates
9 and rarely metastasize, however, it presents high morbidity, since most lesions occur
10 in visible regions of the body such as face, head and neck (BAILEY et al., 2019; WHO,
11 2019). According to the severity, the lesions can even cause disfigurement, generating
12 social and psychological impact on patients (BAILEY et al., 2019; WHO, 2019). Actinic
13 keratosis (AK) is a skin lesion formed by proliferating dysplastic keratinocytes in areas
14 with greater photo-exposure, and is considered a precursor of SCC (REINEHR;
15 BAKOS, 2019).

16 Ultraviolet (UV) radiation from the sun is the greatest environmental
17 risk factor for developing NMSC. It is able to induce direct DNA damage, inflammation,
18 immunosuppression and formation of reactive oxygen species (ROS). Patients with fair
19 skin and high sunlight exposure throughout life are the most affected (FEEHAN;
20 SHANTZ, 2016). In addition to UV exposure and skin pigmentation, other risk factors
21 can contribute to the development of NMSC, such as immunosuppression, genetic
22 factors, infection by some HPV subtypes and smoking (BELBASIS et al., 2016;
23 TESSARI; GIROLOMONI, 2012). Further, previously history of cutaneous neoplasms
24 confers a greater risk of new skin lesions development, including the same type of the
25 primary lesion or multiple types of NMSC lesions (DUARTE et al., 2018).

26 UV radiation-induced oxidative stress (OS) is one of the main
27 mechanisms of skin carcinogenesis (HAY et al., 2014; OLTEANU et al., 2012). Some
28 studies demonstrated systemic OS markers changed in patients with AK and NMSC,
29 such as decreased antioxidant defenses (SRIVASTAVA et al., 2012; VURAL;
30 CANBAZ; SEKÇUKI, 1999), and increased lipid peroxidation (MAJIDI; DJALALI;
31 JAVANBAKHT, 2017; SRIVASTAVA et al., 2012). The inflammatory process can also
32 be modulated by exposure to UV radiation, culminating in greater cell proliferation,
33 angiogenesis and decreased apoptosis, favoring tumor development (FEEHAN;

1 SHANTZ, 2016; OLTEANU et al., 2012). Some authors found differences in some
2 serum cytokines levels in SCC patients, like increased interferon gamma (INF- γ) and
3 transforming growth factor beta1 (TGF- β 1) and decreased interleukin-6 (IL-6) in
4 comparison with individuals without NMSC (YAMADA et al., 2016). In relation to BCC,
5 it was reported increased serum levels of interleukin-10 (IL-10) and decreased
6 interleukin-2 when compared to control (SOBJANEK et al., 2016), in addition
7 decreased TGF- β 1 when compared to BCC patients after photodynamic therapy
8 (ADAMEK et al., 2005).

9 As observed, the literature has presenting some results indicating that
10 besides the participation of OS and inflammation in skin carcinogenesis, they are also
11 systemically different modulated in the different types of NMSC. However, despite the
12 knowledge that previous story of NMSC can increase the risk for the development of
13 other skin lesions, the involvement of OS and inflammation with the presence of
14 multiple types of NMSC in the same patient have never been investigated. Based on
15 this, the objective of this study was to evaluate systemic parameters of OS and
16 inflammatory process in patients with AK and NMSC and to investigate if they could
17 differ in patients with multiple types of NMSC and with pre-malign and malign lesions.

18

19 **Methods and materials**

20 *Ethical approval*

21 This study was approved by the Ethics in Research Committee
22 involving human beings of the State University of Londrina (CEP/UEL), under registers
23 CAAE: 44235015.3.0000.5231 in 20/05/2015 and CAAE: 00745118.7.0000.5231 in
24 14/09/2019. All the practices were approved by the institutional board and were in
25 accordance with the ethical standards of the Brazilian National Commission on Ethics
26 in Research (CONEP) and with the Brazilian National Health Council Resolution
27 466/12.

28 *Sample collection and study design*

29 A total of 182 patients aged between 29 and 87 years old were
30 recruited from Dermatology outpatients of the University Hospital of Londrina, Paraná,
31 Brazil, and from Northern Paranaense Hospital (HONPAR) of Arapongas, Paraná,

1 Brazil, from June 2015 to December 2019. Before surgery for removal of skin lesions,
2 patients were invited to participate in the study and in the case of acceptance, they
3 signed the informed consent form (ICF) after complete explanation of the study, its
4 risks and benefits. After histopathological diagnostic of lesions, 90 patients were
5 grouped in basal cell carcinoma (BCC) group, 24 patients were grouped in squamous
6 cell carcinoma (SCC) group, and 19 patients were grouped in actinic keratosis (AK)
7 group. Some patients presented more than one type of skin lesion, then, those who
8 developed SCC and BCC were grouped in SCC+BCC group (13 patients), and those
9 who developed SCC, BCC and AK were grouped in SCC+BCC+AK group (36
10 patients). The control group comprised 75 individuals and the inclusion criteria were
11 both sex volunteers without any type of cancer history. All participants were not under
12 antioxidant therapy and answered a questionnaire to obtain information about sex, skin
13 phototype (according to Fitzpatrick's classification), sun exposure, location of tumor,
14 smoking, the presence of chronic diseases and the use of medications (insulin,
15 antihypertensive, antilipemic and antihyperglycemic drugs). Peripheral blood were
16 collected with and without heparin and centrifuged at 1331 x G during 5 minutes at 4
17 °C to obtain plasma and serum, respectively. Separated plasma and serum were
18 frozen at -20 °C until analysis, while erythrocytes were used fresh to the evaluation of
19 oxidative stress parameters.

20 *Evaluation of oxidative stress in erythrocytes*

21 Erythrocytes were obtained from heparinized blood and washed three
22 times with 0.9 % saline solution at 4 °C. An aliquot of heparinized blood was used to
23 determine total protein by Lowry technique (LOWRY; ROSEBROUGH; FARR, 1951),
24 modified by Miller, using bovine serum albumin (Sigma Aldrich®) as standard
25 (MILLER, 1956).

26 Total (GT) and reduced glutathione (GSH) levels were determined
27 according to the method previously described by Locatelli et al (2009). The
28 quantification of GT was based on the conversion of oxidized glutathione (GSSG)
29 present in the medium to GSH by the action of the enzyme glutathione reductase, in
30 the presence of NADPH (LOCATELLI et al., 2009). Erythrocytes were diluted at a ratio
31 of 1:800 in deionized water and incubated with 5,5'-Dithiobis-(2-acid-nitrobenzoic)
32 (DTNB). GSH reacts with DTNB forming a yellow compound (5-nitrothiobenzoate)
33 detectable at 412 nm using a Multiskan GO® microplate reader (Thermo Fisher

1 Scientific®, USA). After GT and GSH quantification, the levels of GSSG were
2 estimated taking into account the stoichiometry of the reaction, when two GSH are
3 required to form one GSSG. Therefore, GSSG was determined by the equation: $GSSG$
4 $= (GT-GSH/2)$. The results were expressed in $\mu\text{M/g}$ protein. It was also calculated
5 GSSG/GSH ratio and the results were expressed in percentage (%).

6 The activity of catalase was measured using erythrocytes at a dilution
7 of 1:80 in deionized water (PANIS et al., 2012). The method is based on the
8 decomposition of hydrogen peroxide (H_2O_2) by catalase, directly related to its
9 absorption at 240 nm (AEBI, 1984). The enzymatic kinetics was monitored using a UV-
10 1650 PC UV-Vis spectrophotometer (Shimadzu, Kyoto, Japan). The results were
11 expressed in velocity of absorbance decreased in 1 min per milligram of total protein
12 (Vabs/min/mg protein).

13 Superoxide dismutase (SOD) activity was determined using different
14 volumes of diluted erythrocytes at a ratio of 1:20 in deionized water (MARKLUND;
15 MARKLUND, 1974). This method is based on the inhibition of pyrogallol autoxidation
16 in aqueous solution by SOD, which catalyzes the dismutation of superoxide anion to
17 hydrogen peroxide. This autoxidation inhibition was monitored at 420 nm in a UV-1650
18 PC UV-Vis spectrophotometer (Shimadzu, Kyoto, Japan) during 6 minutes.
19 Autoxidation of pyrogallol alone was used as control. The amount of SOD capable of
20 inhibit in 50 % pyrogallol autoxidation was defined as a unit of enzymatic activity (U).
21 Results were expressed in USOD/g total protein.

22 Lipid peroxidation in erythrocytes was evaluated by
23 chemiluminescence stimulated by tert-butyl hydroperoxide (CL-LOOH) (GONZALEZ
24 FLECHA; LLESUY; BOVERIS, 1991). For this method, erythrocytes were diluted at a
25 ratio of 1:1200 in 10 mM phosphate buffer and the reaction was assessed in a
26 luminometer Lumat³ LB 9508 (Berthold Technologies Bioanalytic) during 40 minutes.
27 After measurement, the results were plotted and area under curve (AUC) were
28 calculated.

29 *Oxidative stress parameters in plasma and serum*

30 Total thiols levels in plasma were quantified by the method based on
31 the reaction between sulfhydryl group (thiol) and DTNB, resulting in a
32 spectrophotometrically detectable color alteration at 412 nm (HU, 1994). Total thiol

1 group was calculated using a calibration curve prepared with GSH. The results were
2 expressed in μM .

3 Plasma malondialdehyde (MDA) was measured by high performance
4 liquid chromatography (HPLC) in a LC-20AT-HPLC system (Shimadzu, Kyoto, Japan)
5 (VICTORINO et al., 2013). The readings were made during 11 minutes, at a flow rate
6 of 0.8 mL/min at 35 °C. The results were expressed in nM of MDA.

7 Serum concentrations of ferritin and the activity of gamma-glutamyl
8 transpeptidase (GGT) were evaluated using a biochemical auto-analyzer Dimension
9 RxL® (Dade Behring/ Siemens Healthcare Diagnostics, Dade Behring, Deerfield, IL,
10 USA).

11 Plasma advanced oxidation protein products (AOPP) levels were
12 measured in a microplate reader. AOPP results from oxidation of protein residues of
13 tyrosine by inflammatory response, leading to the formation of dityrosine-containing
14 protein cross-linking products detected by spectrophotometry at 340 nm (WITKO-
15 SARSAT et al., 1996). AOPP concentrations were expressed as $\mu\text{mol/L}$ of chloramine-
16 T equivalents.

17 *Inflammatory parameters*

18 Plasma cytokine levels of tumor necrosis factor alpha (TNF- α), IL-10
19 and TGF- β 1 were measured by a sandwich enzyme-linked immunosorbent assay
20 (ELISA), following the manufacturers' instructions (eBioscience®, San Diego, CA,
21 USA). TNF- α and IL-10 concentrations were expressed as pg/mL, and TGF- β 1
22 concentrations were expressed as ng/mL.

23 Serum C-reactive protein (CRP) levels were evaluated using a high
24 sensitivity turbidimetry assay (Architect C8000, Abbott Laboratory, Abbott Park, IL,
25 USA), with limit of detection of 0.175 mg/L (hsCRP).

26 *Statistical analysis*

27 In this study, three comparisons were made between the groups: 1)
28 Single type of NMSC lesions: control group vs BCC group vs SCC group; 2) Pre-malign
29 lesion and malign lesion: control group vs AK group vs SCC group; 3) Single and
30 multiple types of NMSC lesions: BCC group vs SCC group vs SCC+BCC group vs
31 SCC+BCC+AK group. The chi-square test and z test for proportion comparison were

1 applied in categorical variables, and were expressed as absolute (n) and relative (%)
2 numbers. The normality of data distribution was assessed using Shapiro-Wilk test, and
3 when the variables were not normally distributed, the natural logarithmic (\ln) of
4 continuous data were used. Univariate analyses of continuous data were performed
5 by ANOVA (one-way or two-way) with Tukey's post-hoc test, and expressed as mean
6 \pm standard error of mean (SEM) when data presented normal distribution. When data
7 present nonparametric distribution, the univariate analysis was performed by Kruskal-
8 Wallis with Dunn's post-hoc test, and results were expressed as median with
9 interquartile range. Multinomial logistic regression analysis was made to evaluate
10 whether the variables that presented $p < 0.1$ in the first analysis were independently
11 associated in all comparisons performed. The results were considered statistically
12 significant when $p < 0.05$. All the statistical analyses were performed in SPSS version
13 24.0 (SPSS, Chicago, IL, USA).

14

15 **Results**

16 *Single type of NMSC lesions comparison*

17 The characteristics of the 114 patients and 75 controls are presented
18 in supplementary table 1. We found that BCC and SCC patients were older than
19 controls ($p < 0.001$). There were no differences between groups in relation to sex
20 ($p = 0.471$), smoking ($p = 0.063$), and chronic diseases ($p = 0.927$). The most reported
21 chronic diseases were systemic arterial hypertension, diabetes mellitus, and
22 hypothyroidism, followed by neurological diseases such as depression, epilepsy, and
23 chronic renal insufficiency. Regarding the use of medications: insulin ($p = 0.403$),
24 antihyperglycemic ($p = 0.340$) and antihypertensive ($p = 0.869$) drugs did not shown any
25 difference between groups, however, BCC and control group differed in relation to the
26 use of antilipemic drugs ($p = 0.015$). Regarding skin phototype ($p < 0.001$), sun exposure
27 ($p < 0.001$) and location of tumor ($p < 0.001$), known risk factors for NMSC, all of them
28 presented significant difference between groups. All variables with $p < 0.1$ (age, use of
29 antilipemic drugs, skin phototype and sun exposure) were used in multinomial logistic
30 regression model as confounding factors, to understand how they can be associated
31 with the results obtained in systemic analysis.

1 Table 1 presents all systemic variables analyzed by univariate
 2 analysis. In erythrocytes samples, GSH levels ($p=0.004$) and GSSG levels ($p<0.001$)
 3 decreased in both NMSC groups when compared to control group. GSSG/GSH ratio,
 4 a marker known as oxidative stress index, also decreased in both NMSC groups
 5 comparing to control group ($p<0.001$). The activity of catalase and SOD did not shown
 6 differences between groups ($p=0.213$ and $p=0.461$, respectively). Lipid peroxidation in
 7 erythrocytes (CL-LOOH) reduced in both NMSC groups when compared to control
 8 group ($p<0.001$). The remaining parameters (total thiols, MDA, GGT, AOPP, ferritin,
 9 TNF- α , IL-10, TGF- β 1 and CRP) evaluated in serum and plasma did not show
 10 differences between groups.

11 Table 2 shows multinomial logistic regression analysis with the two
 12 NMSC (BCC and SCC) groups and controls as dependent variables and systemic
 13 parameters with $p < 0.01$ as explanatory variables. The confounding factors obtained
 14 in the supplementary table 1 were used in different models of multinomial logistic
 15 regression to identify if the explanatory variables were independently associated with
 16 the presence of NMSC. In the first model, the explanatory variables were controlled by
 17 age, smoking and use of antilipemic drugs. In this model, patients with BCC and SCC
 18 presented reduced GSSG/GSH ratio ($p=0.014$ and $p=0.042$, respectively) in
 19 comparison to control. In addition, there was a significant decrease in CL-LOOH in
 20 BCC and SCC groups when compared to control group ($p = 0.002$ and $p=0.010$,
 21 respectively). After the addition of only skin phototype (model 2) or only sun exposure
 22 (model 3) or both (model 4) as confounding factors in the regression models, the
 23 differences in GSSG/GSH ratio in BCC and SCC group ($p=0.012$ and $p=0.025$,
 24 respectively) and in lipid peroxidation (CL-LOOH) in BCC and SCC group ($p=0.004$
 25 and $p=0.002$, respectively) compared to control, continued. None of the explanatory
 26 variables differed between NMSC types. Because control group did not presented any
 27 tumor, the variable location of tumor was not included in any regression model in this
 28 comparison.

29 *Pre-malign and malign lesion*

30 Supplementary Table 2 demonstrates the characteristics of the 39
 31 patients (SCC: $n=24$; AK: $n=15$) and 75 controls included in this comparison. SCC and
 32 AK group were older than control group ($p<0.001$). Characteristics such as sex, skin
 33 phototype, sun exposure, and location of tumor, presence of chronic diseases and the

1 use of medications (data not shown) did not alter between groups. Smoking were
2 higher in patients with SCC when compared to AK and Control ($p=0.022$). Variables
3 with $p<0.1$ (age, smoking and skin phototype) were used in multinomial logistic
4 regression model as confounding factors, to understand how they can be associated
5 with the results obtained in systemic analysis.

6 Table 3 shows a decrease in GSH and GSSG levels decrease in SCC
7 and AK groups compared to control group ($p=0.009$ and $p<0.001$, respectively).
8 GSSG/GSH ratio and lipid peroxidation (CL-LOOH) also decreased in these groups in
9 relation to control ($p =0.002$ and $p<0.001$, respectively; Table 3). The remaining
10 parameters evaluated (catalase, SOD, total thiols, MDA, GGT, AOPP, ferritin, TNF- α ,
11 IL-10, TGF- β 1 and CRP) did not shown differences between groups.

12 Table 4 shows multinomial logistic regression analysis with SCC, AK
13 and control groups as dependent variables and systemic parameters of oxidative
14 stress and inflammation with $p<0.1$ as explanatory variables. The confounding factors
15 obtained in the supplementary table 2 were used in different models of multinomial
16 logistic regression to identify if the explanatory variables were independently
17 associated with the presence of NMSC. In the first model, the explanatory variables
18 were controlled by age and smoking and in the second model, the explanatory
19 variables were controlled by model 1 and skin phototype. The multinomial logistic
20 regression analysis shows that lipid peroxidation is lower in SCC group when
21 compared to control in both models 1 ($p=0.022$) and 2 ($p=0.037$) of regression,
22 indicating that this difference was independently associated with SCC. In addition,
23 none of the other explanatory variables (GSH, GSSG, GSSG/GSH ratio, TNF- α and
24 TGF- β 1) presented differences between pre-malign lesion and malignant lesion (Table
25 4).

26 27 *Single and multiple types of NMSC lesions*

28 Supplementary Table 3 presents the characteristics of 163 patients
29 that were included in this comparison (BCC; SCC; SCC+BCC and SCC+BCC+AK
30 groups). SCC+BCC+AK group was older than the other groups ($p<0.001$). Sun
31 exposure (mainly intense) and location of tumor (mainly lesions located in head/neck)
32 were different between groups ($p=0.019$ and $p<0.001$, respectively). The other
33 characteristics (sex, skin phototype, smoking and chronic diseases) did not shown

1 statistical differences in this comparison. Variables with $p < 0.1$ (age, sun exposure and
2 tumor location) were used in multinomial logistic regression model as confounding
3 factors, to understand how they can be associated with the results obtained in systemic
4 analysis.

5 Table 5 demonstrates the results obtained from the analysis of
6 systemic parameters of OS and inflammation. The only parameter that altered between
7 groups was GGT activity ($p = 0.009$). Decreased activity of GGT was found in SCC
8 group when compared to both SCC+BCC ($p = 0.001$) and SCC+BCC+AK ($p = 0.036$)
9 groups. In addition, it was observed increased activity of GGT in SCC+BCC group
10 compared to BCC group ($p = 0.016$). The other parameters did not present differences
11 between groups.

12 Table 6 shows multinomial logistic regression analysis with BCC,
13 SCC, SCC+BCC and SCC+BCC+AK as dependent variables, GGT activity as
14 explanatory variable. The confounding factors obtained in the supplementary table 3
15 were used in different models of multinomial logistic regression to identify if the
16 explanatory variables were independently associated with the presence of single and
17 multiple types of NMSC. In the first model, the explanatory variables were controlled
18 by age. In the second model they were controlled by model 1 and sun exposure. In the
19 third model, the control was performed by model 1 and tumor location and model 1, 2
20 and 3 were used in the control of explanatory variables in the fourth model. In the
21 models 1 and 2, the activity of GGT were different in BCC when compared to SCC
22 ($p = 0.015$), in addition, SCC+BCC presented different activity of GGT when compared
23 to SCC ($p = 0.008$) and SCC+BCC+AK ($p = 0.043$). However, when both sun exposure
24 and location of tumor were added in regression model (models 3 and 4), the activity of
25 GGT lost significance, indicating that this variable was not independently associated
26 with NMSC, and probably GGT activity was associated with both risk factors.

27

28 **Discussion**

29 The main findings of the present study are that patients with single
30 type NMSC lesions present low levels of systemic OS and that GGT activity is the only
31 evaluated parameter that differ between patients with only BCC or SCC. Furthermore,
32 GGT activity was also the only evaluated parameter that is different between patients

1 that only present SCC from those that present both SCC and BCC. These results were
2 based on a rigorous statistical analysis, by the multinomial logistic regression model
3 that were controlled the interference of potential confounding factors. Our findings must
4 be highlighted since, to the best of our knowledge, we are the first to investigate if
5 systemic parameters of OS and inflammation could be different modulated in patients
6 with single and multiple types of NMSC lesions. Previous studies related alteration in
7 more parameters of OS and inflammation in patients with single type NMSC and AK
8 lesions than we found in this study (ADAMEK et al., 2005; MAJIDI; DJALALI;
9 JAVANBAKHT, 2017; SOBJANEK et al., 2016; SRIVASTAVA et al., 2012; VURAL;
10 CANBAZ; SEKÇUKI, 1999; YAMADA et al., 2016). However, these parameters may
11 be suffering the interference from NMSC risk factors and lifestyle habits, since the
12 control of these confounding factors were not performed.

13 All patients included in this research presented more than 60 years
14 old, fair skin (Fitzpatrick phototype I-III), high exposure to UV radiation and developed
15 lesions in body regions with high sun exposure. This is in accordance with the main
16 risk factors previously described for the development of NMSC and AK (BAILEY et al.,
17 2019; FARTASCH et al., 2012; SCHMITT; BORDEAUX, 2013). It is known that aging
18 (ZHANG; DAVIES; FORMAN, 2015), smoking habits (PASINI et al., 2019), and use of
19 medications such as antilipemic drugs (RASMUSSEN et al., 2016), modulate in
20 different ways systemic OS and inflammation markers. For this reason, it is important
21 to control the confounding factors to minimize the biases in the analysis.

22 OS and chronic inflammation processes were previously related to
23 cancer initiation, promotion, progression and metastasis (REUTER et al., 2011). In our
24 study, we observed that patients with BCC and SCC demonstrated lower levels of
25 GSSG/GSH ratio than control volunteers, which corroborates with a previous study
26 that observed low GSSG/GSH ratio in tissues of patients with head and neck SCC, a
27 carcinoma that mainly affects mucosas (DEQUANTER; DOK; NUYTS, 2017). GSH is
28 a tripeptide with great antioxidant capacity, belongs to the group of non-enzymatic
29 antioxidants and acts capturing ROS, and also as cofactor of enzymes such as
30 glutathione peroxidase (GPx) and glutathione S-transferase (GST), which are
31 important enzymes in detoxification of hydrogen peroxide, lipid hydroperoxides and
32 electrophilic compounds (METGUD; BAJAJ, 2014; REISCHL et al., 2007). The GSH
33 consumed can be converted to the oxidized form, GSSG, or used for protein S-
34 glutathiolation, protecting proteins from oxidative damage (MALVEZZI et al., 2012).

1 The assessment of GSSG/GSH ratio can provide an estimation of intracellular OS and
2 is a well-described marker used to analyze oxidative balance (SÁNCHEZ-
3 RODRÍGUEZ; MENDOZA-NÚÑEZ, 2019). In addition, it is considered to be a great
4 marker of systemic OS in different clinical conditions, such as metabolic,
5 cardiovascular and neurological diseases and cancer (SÁNCHEZ-RODRÍGUEZ;
6 MENDOZA-NÚÑEZ, 2019). In our study, GSSG/GSH ratio presented decreased
7 values in both BCC and SCC patients compared to control volunteers, indicating low
8 systemic intracellular OS. In addition, reduction in GSSG/GSH ratio was independently
9 associated with the presence of single type NMSC lesions.

10 Lipids from membranes are one of the main macromolecules that
11 suffer from oxidative damage caused by interaction with ROS, generating product
12 derived from oxidation, such as lipid hydroperoxides and MDA (WILLIAMS et al.,
13 2014). In the current study, patients with only one type of NMSC (BCC or SCC) showed
14 significant decrease in lipid hydroperoxides, measured by erythrocytic
15 chemiluminescence method (CL-LOOH), and this result were different from other
16 studies that demonstrated increased systemic lipid peroxidation, but using other
17 methods (MAJIDI; DJALALI; JAVANBAKHT, 2017; SRIVASTAVA et al., 2012). These
18 studies measured thiobarbituric acid reactive species (TBARS), which is a method
19 used to measure MDA levels that possess several interferents (GUTTERIDGE, 1986).
20 In our study, we evaluated lipid peroxidation using two methods considered highly
21 sensitive, the measurement of lipid hydroperoxides by chemiluminescence and MDA
22 using HPLC, both products of lipid peroxidation formed in different stages. Besides
23 that, these analysis were performed in two distinct biological compartments, in
24 erythrocytes and plasma, reflecting intracellular and extracellular compartments,
25 respectively. For this reason, our analysis allow a better understanding of the systemic
26 importance of this process in NMSC.

27 We found the greatest activity of serum GGT in patients with the two
28 types of NMSC (SCC+BCC), and observed that the activity of this enzyme differed in
29 patients that present only BCC or only SCC. Previously, it was demonstrated that
30 patients with esophageal SCC present increased levels of serum GGT (HUANG et al.,
31 2017). However, after the control by risk factors by logistic regression model in our
32 study, we observed that GGT activity was associated with tumor location in patients
33 with SCC+BCC and by tumor location combined with sun exposure in patients with
34 single NMSC. GGT is an enzyme present in plasma membrane of all cells, but can

1 also be found in serum, being considered a marker of liver diseases (NICOLAE et al.,
2 2017; POMPELLA et al., 2006). The interference of liver diseases in our GGT results
3 was discarded because our patients did not present hepatic alterations. This enzyme
4 is involved in the metabolism of extracellular GSH and it conjugates and provide an
5 adequate recovery of cysteine, it also play a role in OS, generating ROS in low levels
6 of activity and participates in leukotrienes formation (NICOLAE et al., 2017;
7 POMPELLA et al., 2006). GSH is the main intracellular antioxidant enrolled in
8 detoxification of ROS and lipid hydroperoxides, protecting the cells against lipid
9 peroxidation-induced damage, however GSH can also be found in extracellular
10 medium (POMPELLA et al., 2006). During tumor cell proliferation, cysteine needs to
11 be recovered in intracellular compartment, thereby, GGT present in plasma membrane
12 of cells catalyzes the hydrolysis of extracellular GSH resulting in posterior transport of
13 cysteine into the cell (POMPELLA et al., 2006). The activity of GGT is influenced by
14 the stability of membranes; in equilibrium GGT initiates GSH metabolism culminating
15 in release of cysteine, but low activity of this enzyme leads to OS through generation
16 of pro-oxidant molecules (NICOLAE et al., 2017). In our study, patients presented low
17 GGT activity and low levels of systemic OS. However, we could not exclude the
18 possibility of this enzyme interfere in OS in tumor tissue.

19 Interestingly none of systemic markers of inflammatory process
20 analyzed presented any differences between groups in all comparisons performed in
21 our study, and these results were different from others studies (ADAMEK et al., 2005;
22 SOBJANEK et al., 2016; YAMADA et al., 2016). However, although the methodological
23 procedure to measure the inflammatory mediators was similar, the experimental
24 design was different, since we analyzed the role clinical history of patients, ensuring
25 that patients allocated in single type of NMSC groups (SCC and BCC groups) did not
26 present other type of skin lesions. This kind of analysis is not specified in most of
27 studies. Our results indicate that TNF- α , IL-10, TGF- β 1 and C-reactive protein are not
28 good parameters to evaluate systemically the inflammatory process in patients with
29 single and multiple NMSC lesions.

30 The main limitation of the present study is the small number of
31 individuals that present both NMSC (SCC+BCC group). However, this limitation is a
32 consequence of the rigor adopted in the experimental design, which was fundamental
33 to bring a new approach on NMSC. For the first time, systemic differences between

1 OS and inflammatory parameters have been investigated in patients who have one or
2 more types of NMSC.

3 In conclusion, the analysis of the results allow us to conclude that
4 patients with single type of NMSC present low systemic oxidative stress. Furthermore,
5 GGT activity was the only evaluated parameter that is different in patients with single
6 and multiple NMSC lesions. However, it is influenced by tumor location associated with
7 sun exposure. The results also allow us to conclude that none of the systemic
8 parameters evaluated were able to differentiate the pre-malign lesion (AK) from the
9 malign lesion (SCC). Besides, the inflammatory markers evaluated in this study are
10 not good parameters to systemically investigate inflammation in patients with single
11 and multiple NMSC lesions. These findings enable a better comprehension of systemic
12 alterations related with oxidative stress and inflammation in patients with single and
13 multiple types of NMSC lesions that may contribute to the reduction of morbidity and
14 mortality due to this type of cancer in the future.

15

1 **References**

- 2 ADAMEK, M. et al. Topical ALA-PDT modifies neutrophils' chemiluminescence,
3 lymphocytes' interleukin-1beta secretion and serum level of transforming growth
4 factor beta1 in patients with nonmelanoma skin malignancies: A clinical study.
5 **Photodiagnosis and Photodynamic Therapy**, v. 2, n. 1 SPEC. ISS., p. 65–72,
6 2005.
- 7 AEBI, H. Catalase in Vitro. In: **Methods in Enzymology**. [s.l.] Academic Press, Inc,
8 1984. v. 105p. 121–126.
- 9 BAILEY, A. et al. Management of keratinocyte carcinoma - Special considerations in
10 the elderly. **International Journal of Women's Dermatology**, v. 5, n. 4, p. 235–245,
11 2019.
- 12 BELBASIS, L. et al. Non-genetic risk factors for cutaneous melanoma and
13 keratinocyte skin cancers: An umbrella review of meta-analyses. **Journal of**
14 **Dermatological Science**, v. 84, n. 3, p. 330–339, 2016.
- 15 DEQUANTER, D.; DOK, R.; NUYTS, S. Basal oxidative stress ratio of head and neck
16 squamous cell carcinomas correlates with nodal metastatic spread in patients under
17 therapy. **OncoTargets and Therapy**, v. 10, p. 259–263, 2017.
- 18 DUARTE, A. F. et al. Risk factors for development of new skin neoplasms in patients
19 with past history of skin cancer: A survival analysis. **Scientific Reports**, v. 8, n. 1, p.
20 6–11, 2018.
- 21 FARTASCH, M. et al. The relationship between occupational sun exposure and non-
22 melanoma skin cancer: clinical basics, epidemiology, occupational disease
23 evaluation, and prevention. **Deutsches Ärzteblatt international**, v. 109, n. 43, p.
24 715–20, 2012.
- 25 FEEHAN, R. P.; SHANTZ, L. M. Molecular signaling cascades involved in
26 nonmelanoma skin carcinogenesis. **Biochemical Journal**, v. 473, n. 19, p. 2973–
27 2994, 2016.
- 28 GLOBAL BURDEN OF DISEASE CANCER COLLABORATION. Global, regional,
29 and national cancer incidence, mortality, years of life lost, years lived with disability,
30 and disability-Adjusted life-years for 29 cancer groups, 1990 to 2017: A systematic

- 1 analysis for the global burden of disease study. **JAMA Oncology**, v. 5, n. 12, p.
2 1749–1768, 2019.
- 3 GONZALEZ FLECHA, B.; LLESUY, S.; BOVERIS, A. Hydroperoxide-initiated
4 chemiluminescence: An assay for oxidative stress in biopsies of heart, liver, and
5 muscle. **Free Radical Biology and Medicine**, v. 10, n. 2, p. 93–100, 1991.
- 6 GUTTERIDGE, J. M. C. Aspects to consider when detecting and measuring lipid
7 peroxidation. **Free Radical Research**, v. 1, n. 3, p. 173–184, 1986.
- 8 HAY, R. J. et al. The global burden of skin disease in 2010: An analysis of the
9 prevalence and impact of skin conditions. **Journal of Investigative Dermatology**, v.
10 134, n. 6, p. 1527–1534, 2014.
- 11 HU, M. L. Measurement of protein thiol groups and glutathione in plasma. **Methods**
12 **in Enzymology**, v. 233, n. 1987, p. 380–385, 1994.
- 13 HUANG, H. et al. Prognostic value of pretreatment serum alanine
14 aminotransferase/aspartate aminotransferase (ALT/AST) ratio and gamma
15 glutamyltransferase (GGT) in patients with esophageal squamous cell carcinoma.
16 **BMC Cancer**, v. 17, n. 1, p. 1–11, 2017.
- 17 LOCATELLI, C. et al. Gallic acid ester derivatives induce apoptosis and cell adhesion
18 inhibition in melanoma cells: The relationship between free radical generation,
19 glutathione depletion and cell death. **Chemico-Biological Interactions**, v. 181, n. 2,
20 p. 175–184, 2009.
- 21 LOWRY, O. H.; ROSEBROUGH, N. J.; FARR. Protein measurement with the folin
22 phenol reagent. **Journal of Biological Chemistry**, v. 193, p. 265–72, 1951.
- 23 MAJIDI, Z.; DJALALI, M.; JAVANBAKHT, M. H. Evaluation of the Level of Zinc and
24 Malondialdehyde in Basal Cell Carcinoma. **Iranian Journal of Public Health**, v. 46,
25 n. 8, p. 1104–1109, 2017.
- 26 MALVEZZI, A. et al. The cysteine-rich protein Thimet oligopeptidase as a model of
27 the structural requirements for S-glutathiolation and oxidative oligomerization. **PLoS**
28 **ONE**, v. 7, n. 6, p. 1–11, 2012.
- 29 MARKLUND, S.; MARKLUND, G. Involvement of the superoxide anion radical in the
30 autoxidation of pyrogallol and a convenient assay for superoxide dismutase.

- 1 **European Journal of Biochemistry**, v. 47, p. 469–474, 1974.
- 2 METGUD, R.; BAJAJ, S. Evaluation of salivary and serum lipid peroxidation, and
3 glutathione in oral leukoplakia and oral squamous cell carcinoma. **Journal of oral**
4 **science**, v. 56, n. 2, p. 135–42, 2014.
- 5 MILLER, G. L. Protein determination for large numbers of samples. **Analytical**
6 **Chemistry**, v. 31, p. 964, 1956.
- 7 NICOLAE, I. et al. GAMMA-GLUTAMYL TRANSPEPTIDASE ALTERATION AS A
8 BIOMARKER OF OXIDATIVE STRESS IN PATIENTS WITH HUMAN
9 PAPILLOMAVIRUS LESIONS FOLLOWING TOPICAL TREATMENT WITH
10 SINECATECHINS. **Farmacia**, v. 65, n. 4, p. 617–623, 2017.
- 11 OLTEANU, E. D. et al. Photochemoprotective effect of calluna vulgaris extract on
12 skin exposed to multiple doses of ultraviolet b in SKH-1 hairless mice. **Journal of**
13 **Environmental Pathology, Toxicology and Oncology**, v. 31, n. 3, p. 233–243,
14 2012.
- 15 PANIS, C. et al. Differential oxidative status and immune characterization of the early
16 and advanced stages of human breast cancer. **Breast Cancer Research and**
17 **Treatment**, v. 133, n. 3, p. 881–888, 2012.
- 18 PASINI, A. M. F. et al. Oxidative Stress and Nrf2 Gene Expression in Peripheral
19 Blood Mononuclear Cells Derived From Copd Patients: a Longitudinal Study. **Chest**,
20 v. 155, n. 6, p. A363, 2019.
- 21 POMPELLA, A. et al. Expression of γ -glutamyltransferase in cancer cells and its
22 significance in drug resistance. **Biochemical Pharmacology**, v. 71, n. 3, p. 231–
23 238, 2006.
- 24 RASMUSSEN, S. T. et al. Simvastatin and oxidative stress in humans: A
25 randomized, Double-blinded, Placebo-controlled clinical trial. **Redox Biology**, v. 9, p.
26 32–38, 2016.
- 27 REINEHR, C. P. H.; BAKOS, R. M. Actinic keratoses: review of clinical, dermoscopic,
28 and therapeutic aspects. **Anais Brasileiros de Dermatologia**, v. 94, n. 6, p. 637–
29 657, 2019.
- 30 REISCHL, E. et al. Distribution, adaptation and physiological meaning of thiols from

- 1 vertebrate hemoglobins. **Comparative Biochemistry and Physiology - C**
2 **Toxicology and Pharmacology**, v. 146, n. 1- 2 SPEC. ISS., p. 22–53, 2007.
- 3 REUTER, S. et al. Oxidative stress, inflammation, and cancer: How are they linked?
4 **Free Radical Biology and Medicine**, v. 49, n. 11, p. 1603–1616, 2011.
- 5 SÁNCHEZ-RODRÍGUEZ, M. A.; MENDOZA-NÚÑEZ, V. M. Oxidative stress indexes
6 for diagnosis of health or disease in humans. **Oxidative Medicine and Cellular**
7 **Longevity**, v. 2019, 2019.
- 8 SCHMITT, A. R.; BORDEAUX, J. S. Actinic Neoplasia Syndrome and an Update on
9 the Epidemiology of Basal Cell Carcinoma, Squamous Cell Carcinoma, and Actinic
10 Keratosis. **Current Dermatology Reports**, v. 2, n. 1, p. 42–47, 2013.
- 11 SOBJANEK, M. et al. Clinical significance of IL-2 and IL-10 gene polymorphisms and
12 serum levels in patients with basal-cell carcinoma. **Biomarkers in Medicine**, v. 10, n.
13 2, p. 185–195, 2016.
- 14 SRIVASTAVA, K. C. et al. A Case control study to evaluate oxidative stress in
15 plasma samples of oral malignancy. **Contemporary clinical dentistry**, v. 3, n. 4, p.
16 271–6, 2012.
- 17 TESSARI, G.; GIROLOMONI, G. Nonmelanoma skin cancer in solid organ transplant
18 recipients: Update on epidemiology, risk factors, and management. **Dermatologic**
19 **Surgery**, v. 38, n. 10, p. 1622–1630, 2012.
- 20 VICTORINO, V. J. et al. Decreased oxidant profile and increased antioxidant
21 capacity in naturally postmenopausal women. **Age**, v. 35, n. 4, p. 1411–1421, 2013.
- 22 VURAL, P.; CANBAZ, M.; SEKÇUKI, D. Plasma antioxidant defence in actinic
23 keratosis and basal cell carcinoma. **Journal of the European Academy of**
24 **Dermatology and Venereology : JEADV**, v. 38, n. 6, p. 439–442, 1999.
- 25 WHO. **Skin cancers**. Disponível em:
26 <<https://www.who.int/uv/faq/skincancer/en/index1.html>>.
- 27 WILLIAMS, J. D. et al. Malondialdehyde-Derived Epitopes In Human Skin Result
28 From Acute Exposure To Solar UV And Occur In Nonmelanoma Skin Cancer Tissue.
29 **Journal of Photochemistry and Photobiology B: Biology**, v. 10, n. 1, p. 54–56,
30 2014.

- 1 WITKO-SARSAT, V. et al. Advanced oxidation protein products as a novel marker of
2 oxidative stress in uremia. **Kidney International**, v. 49, p. 1304–1313, 1996.
- 3 YAMADA, S. et al. Cytokine expression profiles in the sera of cutaneous squamous
4 cell carcinoma patients. **Drug discoveries & therapeutics**, v. 10, n. 3, p. 172–176,
5 2016.
- 6 ZHANG, H.; DAVIES, K. J. A.; FORMAN, H. J. Oxidative stress response and Nrf2
7 signaling in aging. **Free Radical Biology and Medicine**, v. 88, n. Part B, p. 314–
8 336, 2015.
- 9

Table 1. Univariate analysis of systemic oxidative stress and systemic inflammatory markers in patients with basal cell carcinoma (BCC), squamous cell carcinoma (SCC) and control group.

| Systemic markers | Control (N=75) | BCC (N=90) | SCC (N=24) | p value |
|---|-------------------------------------|------------------------------------|-----------------------------------|------------------|
| GSH ($\mu\text{M/g}$ protein) | 650.4 (513.0-820.5) ^a | 564.9 (399.1-701.2) ^b | 436.9 (357.8-621.7) ^b | 0.004 |
| GSSG ($\mu\text{M/g}$ protein) | 271.1 (227.4-375.2) ^a | 134.1 (92.30-244.7) ^b | 116.0 (81.45-297.3) ^b | <0.001 |
| GSSG/GSH ratio (%) | 31.56 (25.52-37.74) ^a | 20.54 (14.95-30.13) ^b | 23.51 (16.65-37.14) ^b | <0.001 |
| Catalase activity (Vabs/min . mg protein ⁻¹) | 35.25 (29.39-41.49) | 30.98 (22.61-41.56) | 29.78 (18.35-41.92) | 0.213 |
| SOD activity (U/g protein) | 0.102 (0.082-0.116) | 0.093 (0.072-0.112) | 0.097 (0.074-0.111) | 0.461 |
| Total Thiols (μM) | 460.2 (386.0-551.4) | 487.8 (360.0-592.9) | 467.2 (331.4-608.2) | 0.725 |
| MDA (nM) | 472.6 (146.9-785.2) | 311.8 (107.6-764.8) | 302.6 (173.4-489.7) | 0.374 |
| CL-LOOH (AUC) | 177072 (115205-211164) ^a | 100903 (75316-137046) ^b | 91248 (64502-133374) ^b | <0.001 |
| GGT activity (U/L) | 34 (24-49) | 36 (28-55) | 27 (21-44.5) | 0.148 |
| AOPP (μM chloramine-T/L) | 129.9 (96.10-180.4) | 121.1 (84.5-147.0) | 119.5 (79.15-192.9) | 0.180 |
| Ferritin (ng/mL) | 133.1 (76.15-228.6) | 161.7 (91.60-294.6) | 111.3 (98.20-274.0) | 0.456 |
| TNF- α (pg/mL) | 186.5 \pm 58.64 | 43.87 \pm 17.55 | 61.16 \pm 32.40 | 0.160 |
| IL-10 (pg/mL) | 3.051 (2.258-4.790) | 2.686 (1.916-3.670) | 2.625 (1.664-5.817) | 0.301 |
| TGF- β 1 (ng/mL) | 4.082 \pm 0.675 | 2.660 \pm 0.242 | 3.252 \pm 0.701 | 0.262 |
| CRP (mg/L) | 3.617 \pm 0.588 | 5.777 \pm 1.249 | 2.523 \pm 0.603 | 0.483 |

Results were expressed as median with interquartile range and were statistically analyzed by Kruskal-Wallis with Dunn's post-hoc test, or were expressed as mean \pm SEM (standard error of mean) and were analyzed by one-way ANOVA with Tukey's post-hoc test. Natural logarithmic transformation were used in TNF- α , TGF- β 1 and CRP data. Different letters were considered statistically different between groups. GSH: reduced glutathione; GSSG: oxidized glutathione; SOD: superoxide dismutase; MDA: malondialdehyde; CL-LOOH: chemiluminescence stimulated by tert-butyl hydroperoxide; AUC: area under curve; GGT: gamma-glutamyl transpeptidase; AOPP: advanced oxidative protein products; TNF- α : tumor necrosis factor-alpha; IL-10: interleukin-10; TGF- β 1: transforming growth factor-beta 1; CRP: C-reactive protein. p values < 0.05 are highlighted.

Table 2. Multinomial logistic regression analyses in patients with basal cell carcinoma (BCC) and squamous cell carcinoma (SCC), and control group as dependent variables and systemic parameters of oxidative stress as explanatory variables.

| Explanatory Variables | Group comparison | p value | | | |
|------------------------------------|------------------|--------------|--------------|--------------|--------------|
| | | Model 1 | Model 2 | Model 3 | Model 4 |
| GSH ($\mu\text{M/g}$ protein) | Control vs BCC | 0.238 | 0.283 | 0.222 | 0.168 |
| | Control vs SCC | 0.162 | 0.203 | 0.123 | 0.091 |
| | BCC vs SCC | 0.616 | 0.619 | 0.562 | 0.496 |
| GSSG ($\mu\text{M/g}$ protein) | Control vs BCC | 0.624 | 0.622 | 0.657 | 0.558 |
| | Control vs SCC | 0.497 | 0.546 | 0.509 | 0.537 |
| | BCC vs SCC | 0.830 | 0.866 | 0.844 | 0.949 |
| GSSG/GSH ratio (%) | Control vs BCC | 0.014 | 0.019 | 0.014 | 0.012 |
| | Control vs SCC | 0.042 | 0.061 | 0.032 | 0.025 |
| | BCC vs SCC | 0.855 | 0.856 | 0.896 | 0.899 |
| CL-LOOH (AUC) | Control vs BCC | 0.002 | 0.002 | 0.008 | 0.004 |
| | Control vs SCC | 0.010 | 0.006 | 0.015 | 0.002 |
| | BCC vs SCC | 0.956 | 0.741 | 0.839 | 0.495 |

Model 1: controlled by age, smoking and the use of antilipemic drugs; Model 2: controlled by model 1 and skin phototype; Model 3: controlled by model 1 and sun exposure; Model 4: controlled by model 1, skin phototype and sun exposure. GSH: reduced glutathione; GSSG: oxidized glutathione; CL-LOOH: chemiluminescence stimulated by tert-butyl hydroperoxide; AUC: area under curve. p values <0.05 are highlighted.

Table 3. Univariate analysis of systemic oxidative stress and systemic inflammatory markers in patients with squamous cell carcinoma (SCC), actinic keratosis (AK) and control group.

| Systemic markers | Control (N=75) | SCC (N=24) | AK (N=15) | p value |
|---|-------------------------------------|-----------------------------------|-----------------------------------|------------------|
| GSH ($\mu\text{M/g}$ protein) | 650.4 (513.0-820.5) ^a | 436.9 (357.8-621.7) ^b | 567.4 (450.2-706.1) ^b | 0.009 |
| GSSG ($\mu\text{M/g}$ protein) | 271.1 (227.4-375.2) ^a | 116.0 (81.45-297.3) ^b | 182.5 (88.80-253.0) ^b | <0.001 |
| GSSG/GSH ratio (%) | 31.56 (25.52-37.74) ^a | 23.51 (16.65-37.14) ^b | 23.24 (16.76-31.22) ^b | 0.002 |
| Catalase activity (Vabs/min . mg protein ⁻¹) | 33.95 \pm 1.197 | 29.81 \pm 2.608 | 33.33 \pm 4.134 | 0.310 |
| SOD activity (U/g protein) | 0.102 (0.082-0.116) | 0.097 (0.074-0.111) | 0.089 (0.081-0.114) | 0.696 |
| Total Thiols (μM) | 460.2 (386.0-551.4) | 467.2 (331.4-608.2) | 513.6 (395.9-724.2) | 0.425 |
| MDA (nM) | 472.6 (146.9-785.2) | 302.6 (173.4-489.7) | 769.1 (229.2-1004) | 0.149 |
| CL-LOOH (AUC) | 177072 (115205-211164) ^a | 91248 (64502-133374) ^b | 95249 (70687-153682) ^b | <0.001 |
| GGT activity (U/L) | 34 (24-49) | 27 (21-44.5) | 40 (25-75.5) | 0.378 |
| AOPP (μM chloramine-T/L) | 144.9 \pm 7.718 | 136.6 \pm 17.35 | 127.5 \pm 15.14 | 0.438 |
| Ferritin (ng/mL) | 133.1 (76.15-228.6) | 111.3 (98.20-274.0) | 93.51 (46.36-154.7) | 0.338 |
| TNF- α (pg/mL) | 8.301 (1.994-74.94) | 4.407 (0.899-86.49) | 1.999 (0.887-6.160) | 0.050 |
| IL-10 (pg/mL) | 3.051 (2.258-4.790) | 2.625 (1.664-5.817) | 2.138 (2.403-5.664) | 0.708 |
| TGF- β 1 (ng/mL) | 4.082 \pm 0.675 | 3.252 \pm 0.701 | 1.691 \pm 0.224 | 0.061 |
| CRP (mg/L) | 3.617 \pm 0.588 | 2.523 \pm 0.603 | 4.450 \pm 1.238 | 0.437 |

Results were expressed as median with interquartile range and were statistically analyzed by Kruskal-Wallis with Dunn's post-hoc test, or were expressed as mean \pm SEM (standard error of mean) and were analyzed by one-way ANOVA with Tukey's post-hoc test. Natural logarithmic transformation were used in AOPP, TGF- β 1 and CRP data. Different letters were considered statistically different between groups. GSH: reduced glutathione; GSSG: oxidized glutathione; SOD: superoxide dismutase; MDA: malondialdehyde; CL-LOOH: chemiluminescence stimulated by tert-butyl hydroperoxide; AUC: area under curve; GGT: gamma-glutamyl transpeptidase; AOPP: advanced oxidative protein products; TNF- α : tumor necrosis factor-alpha; IL-10: interleukin-10; TGF- β 1: transforming growth factor-beta 1; CRP: C-reactive protein. p values < 0.05 are highlighted.

Table 4. Multinomial logistic regression analyses in patients with squamous cell carcinoma (SCC) and actinic keratosis (AK) and control group as dependent variables and systemic parameters of oxidative stress and inflammation listed as explanatory variables.

| Explanatory Variables | Group comparisons | p value | |
|------------------------------------|-------------------|--------------|--------------|
| | | Model 1 | Model 2 |
| GSH ($\mu\text{M/g}$ protein) | Control vs SCC | 0.267 | 0.474 |
| | Control vs AK | 0.154 | 0.275 |
| | SCC vs AK | 0.196 | 0.525 |
| GSSG ($\mu\text{M/g}$ protein) | Control vs SCC | 0.963 | 0.899 |
| | Control vs AK | 0.213 | 0.387 |
| | SCC vs AK | 0.286 | 0.643 |
| GSSG/GSH ratio (%) | Control vs SCC | 0.148 | 0.151 |
| | Control vs AK | 0.276 | 0.442 |
| | SCC vs AK | 0.354 | 0.690 |
| CL-LOOH (AUC) | Control vs SCC | 0.022 | 0.037 |
| | Control vs AK | 0.413 | 0.610 |
| | SCC vs AK | 0.832 | 0.964 |
| TNF- α (pg/mL) | Control vs SCC | 0.722 | 0.850 |
| | Control vs AK | 0.110 | 0.114 |
| | SCC vs AK | 0.157 | 0.125 |
| TGF- β 1 (ng/mL) | Control vs SCC | 0.741 | 0.953 |
| | Control vs AK | 0.157 | 0.201 |
| | SCC vs AK | 0.183 | 0.377 |

Model 1: controlled by age; model 2: controlled by model 1 and smoking. GSH: reduced glutathione; GSSG: oxidized glutathione; CL-LOOH: chemiluminescence stimulated by tert-butyl hydroperoxide; AUC: area under curve; TNF- α : tumor necrosis factor-alpha; TGF- β 1: transforming growth factor-beta 1. p values <0.05 are highlighted.

Table 5. Univariate analysis of systemic oxidative stress and inflammatory markers in patients with single and multiple types of nonmelanoma skin cancer.

| Systemic markers | BCC (N=90) | SCC (N=24) | SCC+BCC (N=13) | SCC+BCC+AK (N=36) | p value |
|--|--------------------------|---------------------------|-----------------------------------|---------------------------------|--------------|
| GSH ($\mu\text{M/g}$ protein) | 564.9 (399.1-701.2) | 436.9 (357.8-621.7) | 472.9 (454.4-713.8) | 489.8 (388.3-625.3) | 0.208 |
| GSSG ($\mu\text{M/g}$ protein) | 184.1 \pm 15.63 | 202.7 \pm 43.57 | 191.4 \pm 27.46 | 213.6 \pm 26.66 | 0.635 |
| GSSG/GSH ratio (%) | 20.54 (14.95-30.13) | 23.51 (16.65-37.14) | 22.64 (17.40-32.49) | 27.84 (18.54-35.21) | 0.510 |
| Catalase activity (Vabs/min . mg protein ⁻¹) | 30.98 (22.61-41.56) | 29.78 (18.35-41.92) | 26.69 (12.17-37.51) | 27.72 (21.82-35.16) | 0.604 |
| SOD activity (U/g protein) | 0.094 \pm 0.003 | 0.094 \pm 0.006 | 0.079 \pm 0.010 | 0.097 \pm 0.005 | 0.349 |
| Total Thiols (μM) | 487.8 (360.0-592.9) | 467.2 (331.4-608.2) | 456.4 (393.8-558.7) | 434.1 (335.1-535.0) | 0.547 |
| MDA (nM) | 311.8 (107.6-764.8) | 302.6 (173.4-489.7) | 106.0 (51.42-490.6) | 356.5 (148.5-736.5) | 0.461 |
| CL-LOOH (AUC) | 100903 (75316-137046) | 91248 (64502-133374) | 114961 (51528-186718) | 109479 (61948-190995) | 0.834 |
| GGT activity (U/L) | 36 (28-55) ^{ab} | 27 (21-44.5) ^b | 85.50 (44.25-368.75) ^c | 50.50 (27.5-79.5) ^{ac} | 0.009 |
| AOPP (μM chloramine-T/L) | 121.1 (84.5-147.0) | 119.5 (79.15-192.9) | 123.3 (99.53-189.2) | 124.8 (85.34-162.6) | 0.868 |
| Ferritin (ng/mL) | 220.9 \pm 28.81 | 210.4 \pm 53.97 | 264.3 \pm 90.85 | 236.9 \pm 67.08 | 0.924 |
| TNF- α (pg/mL) | 4.764 (1.223-27.81) | 4.407 (0.899-86.49) | 6.248 (1.662-35.39) | 1.768 (0.554-17.15) | 0.280 |
| IL-10 (pg/mL) | 2.686 (1.916-3.670) | 2.625 (1.664-5.817) | 3.089 (1.661-6.093) | 2.191 (1.437-3.203) | 0.232 |
| TGF- β 1 (ng/mL) | 2.660 \pm 0.242 | 3.252 \pm 0.701 | 0.594 \pm 0.194 | 0.713 \pm 0.128 | 0.585 |
| CRP (mg/L) | 5.777 \pm 1.249 | 2.523 \pm 0.603 | 5.025 \pm 2.125 | 3.614 \pm 0.898 | 0.810 |

BCC: Basal cell carcinoma; SCC: Squamous cell carcinoma; SCC+BCC: Patients with SCC and BCC; SCC+BCC+AK: Patients with BCC, SCC and actinic keratosis (AK). Results were expressed as median with interquartile range and were statistically analyzed by Kruskal-Wallis with Dunn's post-hoc test, or were expressed as mean \pm SEM (standard error of mean) and were analyzed by one-way ANOVA with Tukey's post-hoc test. Natural logarithmic transformation were used in GSSG, Ferritin, TGF- β 1 and CRP data. Different letters were considered statistically different between groups. GSH: reduced glutathione; GSSG: oxidized glutathione; SOD: superoxide dismutase; MDA: malondialdehyde; CL-LOOH: chemiluminescence stimulated by tert-butyl hydroperoxide; AUC: area under curve; GGT: gamma-glutamyl transpeptidase; AOPP: advanced oxidative protein products; TNF- α : tumor necrosis factor-alpha; IL-10: interleukin-10; TGF- β 1: transforming growth factor-beta 1; CRP: C-reactive protein. p values < 0.05 are highlighted.

Table 6. Multinomial logistic regression analyses in patients with single and multiples types of nonmelanoma skin cancer divided into those with only basal cell carcinoma (BCC) and only squamous cells carcinoma (SCC) and patients who developed BCC and SCC (SCC+BCC) and those who developed BCC, SCC and actinic keratosis lesions (SCC+BCC+AK) as dependent variables and systemic gamma-glutamyl transpeptidase (GGT) as explanatory variable.

| Explanatory Variables | Group comparison | p value | | | |
|-----------------------|-----------------------|--------------|--------------|---------|---------|
| | | Model 1 | Model 2 | Model 3 | Model 4 |
| GGT (U/L) | BCC vs SCC | 0.015 | 0.031 | 0.089 | 0.210 |
| | BCC vs SCC+BCC | 0.091 | 0.099 | 0.404 | 0.382 |
| | BCC vs SCC+BCC+AK | 0.078 | 0.121 | 0.131 | 0.338 |
| | SCC vs SCC+BCC | 0.008 | 0.018 | 0.075 | 0.165 |
| | SCC vs SCC+BCC+AK | 0.224 | 0.287 | 0.364 | 0.691 |
| | SCC+BCC vs SCC+BCC+AK | 0.043 | 0.070 | 0.101 | 0.264 |

Model 1: controlled by age; Model 2: controlled by Model 1 and sun exposure; Model 3: controlled by Model 1 and location of tumor; Model 4: controlled by Model 1, sun exposure and location of tumor. p values <0.05 are highlighted.

Supplementary Table 1. Characteristics of participants with single type of nonmelanoma skin cancer and control without cancer.

| Characteristics | Control (N=75) | BCC (N=90) | SCC (N=24) | p value |
|--|-------------------------|-------------------------|---------------------------|------------------|
| Age | 60 (53-70) ^a | 68 (60-75) ^b | 69 (60.5-75) ^b | <0.001 |
| Median (25%-75%) | | | | |
| Sex | | | | 0.471 |
| Male | 37 (49.3%) | 53 (58.9%) | 13 (54.2%) | |
| Female | 38 (50.7%) | 37 (41.1%) | 11 (45.8%) | |
| Skin Phototype (Fitzpatrick's Classification) | | | | <0.001 |
| Type I/II/III | 59 (78.7%) ^a | 88 (97.8%) ^b | 22 (91.7%) ^{ab} | |
| Type IV/V/VI | 16 (21.3%) ^a | 2 (2.2%) ^b | 2 (8.3%) ^{ab} | |
| Sun Exposure | | | | <0.001 |
| Low | 33 (44.0%) ^a | 11 (12.2%) ^b | 9 (37.5%) ^a | |
| Intermediate | 16 (21.3%) ^a | 17 (18.9%) ^a | 6 (25.0%) ^a | |
| Intense | 26 (34.7%) ^a | 62 (68.9%) ^b | 9 (37.5%) ^a | |
| Location of tumor | | | | <0.001 |
| Head/Neck | NA | 74 (82.2%) ^a | 12 (50.0%) ^b | |
| Trunk | NA | 9 (10.0%) ^a | 2 (8.3%) ^a | |
| Upper Limbs | NA | 2 (2.2%) ^a | 6 (25.0%) ^b | |
| Lower Limbs | NA | 1 (1.1%) ^a | 2 (8.3%) ^b | |
| Associated Locations | NA | 4 (4.4%) ^a | 2 (8.3%) ^a | |
| Smoking | | | | 0.063 |
| Yes | 6 (8.0%) | 9 (10.0%) | 6 (25.0%) | |
| No | 69 (92.0%) | 81 (90.0%) | 18 (75.0%) | |
| Use of insulin | | | | 0.403 |
| Yes | 5 (6.7%) | 4 (4.4%) | 0 (0.0%) | |
| No | 70 (93.3%) | 86 (95.6%) | 24 (100%) | |
| Use of antihyperglycemic drugs | | | | 0.340 |
| Yes | 9 (12.0%) | 18 (20.0%) | 5 (20.8%) | |
| No | 66 (88.0%) | 72 (80.0%) | 19 (79.2%) | |
| Use of antihypertensive drugs | | | | 0.869 |
| Yes | 36 (48.0%) | 45 (50.0%) | 13 (54.2%) | |
| No | 39 (52.0%) | 45 (50.0%) | 11 (45.8%) | |
| Use of antilipemic drugs | | | | 0.015 |
| Yes | 20 (26.7%) ^a | 9 (10.0%) ^b | 3 (12.5%) ^{ab} | |
| No | 55 (73.3%) ^a | 81 (90.0%) ^b | 21 (87.5%) ^{ab} | |
| Chronic diseases | | | | 0.927 |
| Yes | 53 (70.7%) | 62 (68.9%) | 16 (66.7%) | |
| No | 22 (29.3%) | 28 (31.1%) | 8 (33.3%) | |

Qualitative data were evaluated by chi-square and z test for proportion comparison, and quantitative data were evaluated by Kruskal-Wallis test with Dunn's post-hoc test. Different letters were considered statistically different between groups. BCC, Basal cell carcinoma; SCC, Squamous cell carcinoma; NA, Not applicable. p values <0.05 are highlighted.

Supplementary Table 2. Characteristics of participants with squamous cell carcinoma (SCC), actinic keratosis (AK) and control without cancer.

| Characteristics | Control (N=75) | SCC (N=24) | AK (N=15) | p value |
|--|-------------------------|---------------------------|-------------------------|------------------|
| Age | | | | |
| Median (25%-75%) | 60 (53-70) ^a | 69 (60.5-75) ^b | 72 (68-75) ^b | <0.001 |
| Sex | | | | 0.900 |
| Male | 37 (49.3%) | 13 (54.2%) | 8 (53.3%) | |
| Female | 38 (50.7%) | 11 (45.8%) | 7 (46.7%) | |
| Skin Phototype (Fitzpatrick's Classification) | | | | 0.062 |
| Type I/II/III | 59 (78.7%) | 22 (91.7%) | 15 (100%) | |
| Type IV/V/VI | 16 (21.3%) | 2 (8.3%) | 0 (0.0%) | |
| Sun Exposure | | | | 0.537 |
| Low | 33 (44.0%) | 9 (37.5%) | 3 (20.0%) | |
| Intermediate | 16 (21.3%) | 6 (25.0%) | 5 (33.3%) | |
| Intense | 26 (34.7%) | 9 (51.3%) | 7 (46.7%) | |
| Location of tumor | | | | 0.490 |
| Head/Neck | NA | 12 (50.0%) | 7 (46.7%) | |
| Trunk | NA | 2 (8.3%) | 4 (26.7%) | |
| Upper Limbs | NA | 6 (25.0%) | 3 (20.0%) | |
| Lower Limbs | NA | 2 (8.3%) | 0 (0.0%) | |
| Associated Locations | NA | 2 (8.3%) | 1 (6.7%) | |
| Smoking | | | | 0.022 |
| Yes | 6 (8.0%) ^a | 6 (25.0%) ^b | 0 (0.0%) ^a | |
| No | 69 (92.0%) ^a | 18 (75.0%) ^b | 15 (100%) ^a | |
| Chronic diseases | | | | 0.606 |
| Yes | 53 (70.7%) | 15 (62.5%) | 9 (60.0%) | |
| No | 22 (29.3%) | 9 (37.5%) | 6 (40.0%) | |

Qualitative data were evaluated by using a chi-square and z test for proportion comparison. Quantitative data were evaluated by using Kruskal-Wallis test with Dunn's post-hoc test. Different letters were considered statistically different between groups. SCC: Squamous cell carcinoma; AK: actinic Keratosis; NA. Not applicable. p values <0.05 are highlighted.

1 **Supplementary Table 3.** Characteristics of participants who presented single and
 2 multiple types of nonmelanoma skin cancer (NMSC).

| Characteristics | BCC (N=90) | SCC (N=24) | SCC+BCC (N=13) | SCC+BCC+AK (N=36) | p value |
|--|------------------------------|----------------------------|-------------------------|-------------------------|------------------|
| Age | 68 | 69 | 68 | 76.5 | <0.001 |
| Median (25%-75%) | (60-75) ^a | (60.5-75) ^a | (59.5-74) ^a | (71.25-80) ^b | |
| Sex | | | | | 0.389 |
| Male | 53 (58.9%) | 13 (54.2%) | 10 (76.9%) | 18 (50.0%) | |
| Female | 37 (41.1%) | 11 (45.8%) | 3 (23.1%) | 18 (50.0%) | |
| Skin Phototype (Fitzpatrick's Classification) | | | | | 0.210 |
| Type I/II/III | 88 (97.8%) | 22 (91.7%) | 12 (92.3%) | 36 (100%) | |
| Type IV/V/VI | 2 (2.2%) | 2 (8.3%) | 1 (7.7%) | 0 (0.0%) | |
| Sun Exposure | | | | | 0.019 |
| Low | 11 (12.2%) ^a | 9 (37.5%) ^b | 3 (23.1%) ^{ab} | 4 (11.1%) ^a | |
| Intermediate | 17 (18.9%) ^{abc} | 6 (25.0%) ^b | 0 (0.0%) ^c | 9 (25.0%) ^{ab} | |
| Intense | 62 (68.9%) ^a | 9 (37.5%) ^b | 10 (76.9%) ^a | 23 (63.9%) ^a | |
| Location of tumor | | | | | <0.001 |
| Head/Neck | 74 (82.2%) ^a | 12 (50.0%) ^b | 9 (69.2%) ^{ab} | 22 (61.1%) ^b | |
| Trunk | 9 (10.0%) ^a | 2 (8.3%) ^{ab} | 2 (15.4%) ^a | 0 (0.0%) ^b | |
| Upper Limbs | 2 (2.2%) ^a | 6 (25.0%) ^b | 0 (0.0%) ^{ac} | 6 (16.7%) ^{bc} | |
| Lower Limbs | 1 (1.1%) ^a | 2 (8.3%) ^b | 0 (0.0%) ^{ab} | 0 (0.0%) ^{ab} | |
| Associated Locations | 4 (4.4%) ^a | 2 (8.3%) ^{ab} | 2 (15.4%) ^{ab} | 8 (22.2%) ^b | |
| Smoking | | | | | 0.264 |
| Yes | 9 (10.0%) | 6 (25.0%) | 2 (15.4%) | 4 (11.1%) | |
| No | 81 (90.0%) | 18 (75.0%) | 11 (84.6%) | 32 (88.9%) | |
| Chronic diseases | | | | | 0.498 |
| Yes | 62 (68.9%) | 15 (62.5%) | 7 (53.8%) | 27 (75.0%) | |
| No | 28 (31.1%) | 9 (37.5%) | 6 (46.2%) | 9 (25.0%) | |

3 Qualitative data were evaluated by using a chi-square and z test for proportion comparison.
 4 Quantitative data were evaluated by using Kruskal-Wallis test with Dunn's post-hoc test.
 5 Different letters were considered statistically different between groups. BCC: Basal cell
 6 carcinoma; SCC: Squamous cell carcinoma; AK: Actinic keratosis. SCC+BCC: patients with
 7 BCC and SCC. SCC+BCC+AK: patients with BCC, SCC and actinic keratosis.

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1 4 CONCLUSÃO

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- 4 • Os pacientes com CBC e CEC possuem baixos níveis de EO sistêmico,
5 observados por níveis reduzidos na razão entre glutathiona oxidada e reduzida
6 e na lipoperoxidação, e essa redução deve-se à presença do câncer, não sendo
7 influenciada por fatores de risco como fotótipo de pele clara e grau de
8 exposição solar;
 - 9 • A atividade da GGT sistêmica é o único parâmetro de EO, dentre os avaliados,
10 que difere entre os dois tipos de CPNM, além de estar associada com o
11 desenvolvimento dos dois tipos de CPNM em um mesmo paciente, entretanto,
12 a localização do tumor interfere nessas diferenças;
 - 13 • Nenhum dos marcadores de estresse oxidativo e inflamação analisados difere
14 sistemicamente entre quem possui a lesão pré-maligna (QA) e maligna (CEC);
 - 15 • Os parâmetros de inflamação avaliados não diferem sistemicamente nos
pacientes, independente de possuírem um ou mais tipos de CPNM.

1 **5 CONSIDERAÇÕES FINAIS**

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Esse trabalho apresenta como principal limitação o número reduzido de participantes em alguns grupos. Isso se deve principalmente ao rigor adotado no delineamento experimental, que foi fundamental para trazer uma nova abordagem sobre o câncer de pele não-melanoma. Pela primeira vez, diferenças sistêmicas entre parâmetros de estresse oxidativo e de inflamação foram investigadas em pessoas que apresentam um ou mais de um tipo de câncer de pele não-melanoma. Além disso, o trabalho inovou ao considerar características que podem influenciar nos níveis de estresse oxidativo e mediadores inflamatórios sistêmicos na análise estatística dos dados obtidos, proporcionando maior robustez e relevância aos resultados.

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REFERÊNCIAS

ADAMEK, M. et al. Topical ALA-PDT modifies neutrophils' chemiluminescence, lymphocytes' interleukin-1beta secretion and serum level of transforming growth factor beta1 in patients with nonmelanoma skin malignancies: A clinical study. **Photodiagnosis and Photodynamic Therapy**, v. 2, n. 1 SPEC. ISS., p. 65–72, 2005.

AEBI, H. Catalase in Vitro. In: **Methods in Enzymology**. [s.l.] Academic Press, Inc, 1984. v. 105p. 121–126.

AROUN, A. et al. Iron, oxidative stress and the example of solar ultraviolet A radiation. **Photochemical and Photobiological Sciences**, v. 11, n. 1, p. 118–134, 2012.

BAILEY, A. et al. Management of keratinocyte carcinoma - Special considerations in the elderly. **International Journal of Women's Dermatology**, v. 5, n. 4, p. 235–245, 2019.

BALPANDE, A. R.; SATHAWANE, R. Estimation and Comparative Evaluation of Serum Iron, Copper, Zinc and Copper/Zinc Ratio in Oral Leukoplakia, Submucous Fibrosis and Squamous Cell Carcinoma. **Journal of Indian Academy of Oral Medicine and Radiology**, v. 22, n. June, p. 73–76, 2010.

BELBASIS, L. et al. Non-genetic risk factors for cutaneous melanoma and keratinocyte skin cancers: An umbrella review of meta-analyses. **Journal of Dermatological Science**, v. 84, n. 3, p. 330–339, 2016.

BRANDT, M. G.; MOORE, C. C. Nonmelanoma Skin Cancer. **Facial Plastic Surgery Clinics of North America**, v. 27, n. 1, p. 1–13, 2019.

BRENNER, M.; HEARING, V. J. The Protective Role of Melanin Against UV Damage in Human Skin. **Photochemistry & Photobiology**, v. 84, n. 3, p. 539–549, 2008.

CAMERON, M. C. et al. Basal cell carcinoma: Epidemiology; pathophysiology; clinical and histological subtypes; and disease associations. **Journal of the American Academy of Dermatology**, v. 80, n. 2, p. 303–317, 2019.

CARR, T. D. et al. Inhibition of mammalian target of rapamycin (mTOR) suppresses UVB-induced keratinocyte proliferation and survival. **Cancer prevention research**, v. 5, n. 12, p. 1394–1404, 2012.

DAMIANI, E.; ULLRICH, S. E. Understanding the connection between platelet-

- 1 activating factor, a UV-induced lipid mediator of inflammation, immune suppression
2 and skin cancer. **Progress in Lipid Research**, v. 63, p. 14–27, 2016.
- 3 DEQUANTER, D.; DOK, R.; NUYTS, S. Basal oxidative stress ratio of head and neck
4 squamous cell carcinomas correlates with nodal metastatic spread in patients under
5 therapy. **OncoTargets and Therapy**, v. 10, p. 259–263, 2017.
- 6 DUARTE, A. F. et al. Risk factors for development of new skin neoplasms in patients
7 with past history of skin cancer: A survival analysis. **Scientific Reports**, v. 8, n. 1, p.
8 6–11, 2018.
- 9 FAJUYIGBE, D.; YOUNG, A. R. The impact of skin colour on human photobiological
10 responses. **Pigment Cell and Melanoma Research**, v. 29, n. 6, p. 607–618, 2016.
- 11 FARTASCH, M. et al. The relationship between occupational sun exposure and non-
12 melanoma skin cancer: clinical basics, epidemiology, occupational disease
13 evaluation, and prevention. **Deutsches Ärzteblatt international**, v. 109, n. 43, p.
14 715–20, 2012.
- 15 FEEHAN, R. P.; SHANTZ, L. M. Molecular signaling cascades involved in
16 nonmelanoma skin carcinogenesis. **Biochemical Journal**, v. 473, n. 19, p. 2973–
17 2994, 2016.
- 18 FEKECS, T. et al. Changes in oxidative stress in patients screened for skin cancer
19 after solid-organ transplantation. **Transplantation Proceedings**, v. 42, n. 6, p. 2336–
20 2338, 2010.
- 21 FERNANDEZ FIGUERAS, M. T. From actinic keratosis to squamous cell carcinoma:
22 pathophysiology revisited. **Journal of the European Academy of Dermatology and**
23 **Venereology**, v. 31, p. 5–7, 2017.
- 24 GLOBAL BURDEN OF DISEASE CANCER COLLABORATION. Global, regional,
25 and national cancer incidence, mortality, years of life lost, years lived with disability,
26 and disability-Adjusted life-years for 29 cancer groups, 1990 to 2017: A systematic
27 analysis for the global burden of disease study. **JAMA Oncology**, v. 5, n. 12, p.
28 1749–1768, 2019.
- 29 GONZALEZ FLECHA, B.; LLESUY, S.; BOVERIS, A. Hydroperoxide-initiated
30 chemiluminescence: An assay for oxidative stress in biopsies of heart, liver, and
31 muscle. **Free Radical Biology and Medicine**, v. 10, n. 2, p. 93–100, 1991.
- 32 GORODETSKY, R.; SHESKIN, J.; WEINREB, A. Iron, Copper, and Zinc
33 Concentrations in Normal Skin and in Various Nonmalignant and Malignant Lesions.
34 **International Journal of Dermatology**, v. 25, n. 7, p. 440–445, 1986.

- 1 HAY, R. J. et al. The global burden of skin disease in 2010: An analysis of the
2 prevalence and impact of skin conditions. **Journal of Investigative Dermatology**, v.
3 134, n. 6, p. 1527–1534, 2014.
- 4 HOFBAUER, G. F. L.; BAVINCK, J. N. B.; EUVRARD, S. Organ transplantation and
5 skin cancer: Basic problems and new perspectives. **Experimental Dermatology**, v.
6 19, n. 6, p. 473–482, 2010.
- 7 HU, M. L. Measurement of protein thiol groups and glutathione in plasma. **Methods**
8 **in Enzymology**, v. 233, n. 1987, p. 380–385, 1994.
- 9 HUANG, H. et al. Prognostic value of pretreatment serum alanine
10 aminotransferase/aspartate aminotransferase (ALT/AST) ratio and gamma
11 glutamyltransferase (GGT) in patients with esophageal squamous cell carcinoma.
12 **BMC Cancer**, v. 17, n. 1, p. 1–11, 2017.
- 13 INCA, I. N. DO C. J. A. G. DA SI. **Tipos de câncer: Pele não melanoma**. Disponível
14 em:
15 <http://www2.inca.gov.br/wps/wcm/connect/tiposdecancer/site/home/pele_ao_melanoma>.
16
- 17 KOH, H. K. et al. Prevention and Early detection strategies for melanoma and skin
18 cancer. **Archives of Dermatology**, v. 132, p. 436–443, 1996.
- 19 KRSTIĆ, J. et al. Transforming growth factor-beta and oxidative stress interplay:
20 Implications in tumorigenesis and cancer progression. **Oxidative Medicine and**
21 **Cellular Longevity**, v. 2015, 2015.
- 22 LOCATELLI, C. et al. Gallic acid ester derivatives induce apoptosis and cell adhesion
23 inhibition in melanoma cells: The relationship between free radical generation,
24 glutathione depletion and cell death. **Chemico-Biological Interactions**, v. 181, n. 2,
25 p. 175–184, 2009.
- 26 LOWRY, O. H.; ROSEBROUGH, N. J.; FARR. Protein measurement with the folin
27 phenol reagent. **Journal of Biological Chemistry**, v. 193, p. 265–72, 1951.
- 28 LUCAS, R. et al. Solar Ultraviolet Radiation: Global burden of disease from solar
29 ultraviolet radiation. **World Health Organization**, v. 55, n. 13, p. 987–999, 2006.
- 30 MAJIDI, Z.; DJALALI, M.; JAVANBAKHT, M. H. Evaluation of the Level of Zinc and
31 Malondialdehyde in Basal Cell Carcinoma. **Iranian Journal of Public Health**, v. 46,
32 n. 8, p. 1104–1109, 2017.
- 33 MALVEZZI, A. et al. The cysteine-rich protein Thimet oligopeptidase as a model of
34 the structural requirements for S-glutathiolation and oxidative oligomerization. **PLoS**

- 1 **ONE**, v. 7, n. 6, p. 1–11, 2012.
- 2 MARKLUND, S.; MARKLUND, G. Involvement of the superoxide anion radical in the
3 autoxidation of pyrogallol and a conveniente assay for superoxide dismutase.
4 **European Journal of Biochemistry**, v. 47, p. 469–474, 1974.
- 5 MARZUKA, A. G.; BOOK, S. E. Basal Cell Carcinoma: Pathogenesis, Epidemiology,
6 Clinical Features, Diagnosis, Histopathology, and Management. **Yale Journal of**
7 **Biology and Medicine**, v. 88, p. 167–179, 2015.
- 8 MASSAGUÉ, J. TGF β signalling in context. **Nature Reviews Molecular Cell**
9 **Biology**, v. 13, n. 10, p. 616–630, 2012.
- 10 METGUD, R.; BAJAJ, S. Evaluation of salivary and serum lipid peroxidation, and
11 glutathione in oral leukoplakia and oral squamous cell carcinoma. **Journal of oral**
12 **science**, v. 56, n. 2, p. 135–42, 2014.
- 13 MILLER, G. L. Protein determination for large numbers of samples. **Analytical**
14 **Chemistry**, v. 31, p. 964, 1956.
- 15 NICOLAE, I. et al. GAMMA-GLUTAMYL TRANSPEPTIDASE ALTERATION AS A
16 BIOMARKER OF OXIDATIVE STRESS IN PATIENTS WITH HUMAN
17 PAPILOMAVIRUS LESIONS FOLLOWING TOPICAL TREATMENT WITH
18 SINECATECHINS. **Farmacía**, v. 65, n. 4, p. 617–623, 2017.
- 19 OLTEANU, E. D. et al. Photochemoprotective effect of calluna vulgaris extract on
20 skin exposed to multiple doses of ultraviolet b in SKH-1 hairless mice. **Journal of**
21 **Environmental Pathology, Toxicology and Oncology**, v. 31, n. 3, p. 233–243,
22 2012.
- 23 PANDEYA, N.; OLSEN, C. M.; WHITEMAN, D. C. The incidence and multiplicity
24 rates of keratinocyte cancers in Australia. **Medical Journal of Australia**, v. 207, n. 8,
25 p. 339–343, 2017.
- 26 PANIS, C. et al. Differential oxidative status and immune characterization of the early
27 and advanced stages of human breast cancer. **Breast Cancer Research and**
28 **Treatment**, v. 133, n. 3, p. 881–888, 2012.
- 29 PASINI, A. M. F. et al. Oxidative Stress and Nrf2 Gene Expression in Peripheral
30 Blood Mononuclear Cells Derived From Copd Patients: a Longitudinal Study. **Chest**,
31 v. 155, n. 6, p. A363, 2019.
- 32 PAYETTE, M. J.; WHALEN, J.; GRANT-KELS, J. M. Nutrition and nonmelanoma skin
33 cancers. **Clinics in Dermatology**, v. 28, n. 6, p. 650–662, 2010.
- 34 POMPELLA, A. et al. Expression of γ -glutamyltransferase in cancer cells and its

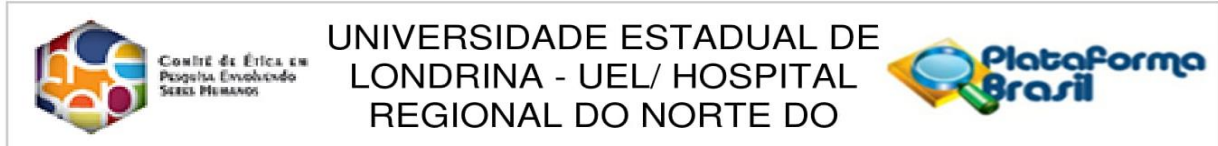
- 1 significance in drug resistance. **Biochemical Pharmacology**, v. 71, n. 3, p. 231–
2 238, 2006.
- 3 RASMUSSEN, S. T. et al. Simvastatin and oxidative stress in humans: A
4 randomized, Double-blinded, Placebo-controlled clinical trial. **Redox Biology**, v. 9, p.
5 32–38, 2016.
- 6 REINEHR, C. P. H.; BAKOS, R. M. Actinic keratoses: review of clinical, dermoscopic,
7 and therapeutic aspects. **Anais Brasileiros de Dermatologia**, v. 94, n. 6, p. 637–
8 657, 2019.
- 9 REISCHL, E. et al. Distribution, adaptation and physiological meaning of thiols from
10 vertebrate hemoglobins. **Comparative Biochemistry and Physiology - C**
11 **Toxicology and Pharmacology**, v. 146, n. 1- 2 SPEC. ISS., p. 22–53, 2007.
- 12 REUTER, S. et al. Oxidative stress, inflammation, and cancer: How are they linked?
13 **Free Radical Biology and Medicine**, v. 49, n. 11, p. 1603–1616, 2011.
- 14 ROLLISON, D. E. et al. Case-control study of smoking and non-melanoma skin
15 cancer. **Cancer Causes Control**, v. 23, n. 2, p. 245–254, 2012.
- 16 ROTHENBERG, S. M.; ELLISEN, L. W. The molecular pathogenesis of head and
17 neck squamous cell carcinoma. **The Journal of clinical investigation**, v. 122, n. 6,
18 p. 1951–7, 2012.
- 19 SÁNCHEZ-RODRÍGUEZ, M. A.; MENDOZA-NÚÑEZ, V. M. Oxidative stress indexes
20 for diagnosis of health or disease in humans. **Oxidative Medicine and Cellular**
21 **Longevity**, v. 2019, 2019.
- 22 SCHEUER, C. Melatonin for prevention of erythema and oxidative stress in response
23 to ultraviolet radiation. n. 47, p. 1–15, 2017.
- 24 SCHMITT, A. R.; BORDEAUX, J. S. Actinic Neoplasia Syndrome and an Update on
25 the Epidemiology of Basal Cell Carcinoma, Squamous Cell Carcinoma, and Actinic
26 Keratosis. **Current Dermatology Reports**, v. 2, n. 1, p. 42–47, 2013.
- 27 SCHMITT, J. et al. Occupational UV-exposure is a major risk factor for basal cell
28 carcinoma: Results of the population-based case-control study FB-181. **Journal of**
29 **Occupational and Environmental Medicine**, v. 60, n. 1, p. 36–43, 2017.
- 30 SCHMITT, J. V.; MIOT, H. A. Actinic keratosis: a clinical and epidemiological
31 revision. **Anais Brasileiros de Dermatologia**, v. 87, n. 3, p. 425–434, 2012.
- 32 SIEGEL, J. A.; KORGAVKAR, K.; WEINSTOCK, M. A. Current perspective on actinic
33 keratosis: a review. **British Journal of Dermatology**, v. 177, n. 2, p. 350–358, 2017.
- 34 SILVERBERG, J. I.; RATNER, D. Associations of non-melanoma skin cancer and

- 1 melanoma, extra-cutaneous cancers and smoking in adults: a US population-based
2 study. **Journal of the European Academy of Dermatology and Venereology :**
3 **JEADV**, v. 29, n. 7, p. 1389–1397, 2015.
- 4 SOBJANEK, M. et al. Clinical significance of IL-2 and IL-10 gene polymorphisms and
5 serum levels in patients with basal-cell carcinoma. **Biomarkers in Medicine**, v. 10, n.
6 2, p. 185–195, 2016.
- 7 SRIVASTAVA, K. C. et al. A Case control study to evaluate oxidative stress in
8 plasma samples of oral malignancy. **Contemporary clinical dentistry**, v. 3, n. 4, p.
9 271–6, 2012.
- 10 ȚÂNȚU, M. M. et al. Prevalence and histopathological types of skin carcinomas in
11 arges county, Romania. **Romanian Journal of Morphology and Embryology**, v.
12 55, n. 3, p. 803–809, 2014.
- 13 TESSARI, G.; GIROLOMONI, G. Nonmelanoma skin cancer in solid organ transplant
14 recipients: Update on epidemiology, risk factors, and management. **Dermatologic**
15 **Surgery**, v. 38, n. 10, p. 1622–1630, 2012.
- 16 VICTORINO, V. J. et al. Decreased oxidant profile and increased antioxidant
17 capacity in naturally postmenopausal women. **Age**, v. 35, n. 4, p. 1411–1421, 2013.
- 18 VURAL, P. et al. Lipid profile in actinic keratosis and basal cell carcinoma.
19 **International Journal of Dermatology**, v. 38, n. 6, p. 439–442, 1999.
- 20 VURAL, P.; CANBAZ, M.; SEKÇUKI, D. Plasma antioxidant defence in actinic
21 keratosis and basal cell carcinoma. **Journal of the European Academy of**
22 **Dermatology and Venereology : JEADV**, v. 38, n. 6, p. 439–442, 1999.
- 23 WARSZAWIK-HENDZEL, O. et al. Non-invasive diagnostic techniques in the
24 diagnosis of squamous cell carcinoma. **Journal of Dermatological Case Reports**,
25 v. 9, n. 4, p. 89–97, 2015.
- 26 WHO. **Skin cancers**. Disponível em:
27 <<https://www.who.int/uv/faq/skincancer/en/index1.html>>.
- 28 WILLIAMS, J. D. et al. Malondialdehyde-Derived Epitopes In Human Skin Result
29 From Acute Exposure To Solar UV And Occur In Nonmelanoma Skin Cancer Tissue.
30 **Journal of Photochemistry and Photobiology B: Biology**, v. 10, n. 1, p. 54–56,
31 2014.
- 32 WITKO-SARSAT, V. et al. Advanced oxidation protein products as a novel marker of
33 oxidative stress in uremia. **Kidney International**, v. 49, p. 1304–1313, 1996.
- 34 YAMADA, S. et al. Cytokine expression profiles in the sera of cutaneous squamous

- 1 cell carcinoma patients. **Drug discoveries & therapeutics**, v. 10, n. 3, p. 172–176,
- 2 2016.
- 3 ZHANG, H.; DAVIES, K. J. A.; FORMAN, H. J. Oxidative stress response and Nrf2
- 4 signaling in aging. **Free Radical Biology and Medicine**, v. 88, n. Part B, p. 314–
- 5 336, 2015.
- 6

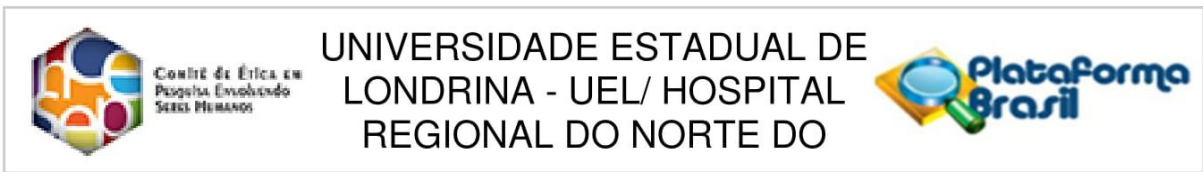
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ANEXOS

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3**ANEXO A**Cópia do parecer do Comitê de Ética em Pesquisa Envolvendo Seres Humanos da
UEL - 2015**PARECER CONSUBSTANCIADO DO CEP****DADOS DO PROJETO DE PESQUISA****Título da Pesquisa:** PERFIL OXIDATIVO SISTÊMICO E DO MICROAMBIENTE TUMORAL DE PACIENTES PORTADORES DE CÂNCER DE PELE NÃO MELANOMA**Pesquisador:** Alessandra Lourenço Cecchini Armani**Área Temática:****Versão:** 4**CAAE:** 44235015.3.0000.5231**Instituição Proponente:** Programa de PG em Patologia Experimental**Patrocinador Principal:** Financiamento Próprio**DADOS DO PARECER****Número do Parecer:** 1.077.557**Data da Relatoria:** 20/05/2015**Apresentação do Projeto:**

No documento intitulado "PB_RELATORIO_PESQUISA_475300.pdf", item Resumo, lê-se: Sendo assim compreender os aspectos oxidativos da doença pode adicionar novos conhecimentos sobre sua etiopatogênese e auxiliar na investigação de novas estratégias terapêuticas no seu tratamento. Com isso pretende-se caracterizar biomarcadores oxidativos no sangue e de crescimento celular no tecido tumoral de pacientes com câncer de pele não-melanoma. Pacientes com diagnóstico clínico de CPNM e com indicação cirúrgica de remoção do tumor serão recrutados de Hospitais de Londrina-PR e Região durante aproximadamente 36 meses. Será retirado sangue antes do procedimento cirúrgico e amostras da biópsia do tumor com margem serão encaminhadas a patologia para o diagnóstico histopatológico da doença e processadas para pesquisa de estresse oxidativo e nitrosativo no laboratório de Patologia Molecular da Universidade. O sangue será submetido a análises bioquímicas para -glutamiltanspeptidase, proteína C reativa, ácido úrico, TNF- e IL-1b. Para a análise de estresse oxidativo plasmático serão realizados as análises de capacidade antioxidante total, catalase, glutatona, superóxido dismutase, malondialdeído e tiol total. A análise de 3-nitrotirosina por imunohistoquímica do tecido tumoral e da margem normal do tecido para verificar a participação de espécies reativas de oxigênio in situ. Os tipos de tumores

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Continuação do Parecer: 1.077.557

serão agrupados como carcinoma basocelular (CBC) e carcinoma espinocelular (CEC) e a pele normal do paciente será considerada seu próprio controle. As comparações serão feitas da seguinte forma: grupo controle versus grupo CBC; controle versus CEC; CBC versus CEC. As análises estatísticas empregadas serão as mais adequadas quanto da distribuição da normalidade dos grupos."

Objetivo da Pesquisa:

OBJETIVO PRIMÁRIO

Caracterizar biomarcadores oxidativos no sangue e de crescimento celular no tecido tumoral de pacientes com câncer de pele não-melanoma.

OBJETIVOS SECUNDÁRIOS

- (1) Caracterizar o estresse oxidativo sistêmico e tumoral e parâmetros inflamatórios sanguíneos em pacientes com câncer de pele não-melanoma basocelular e espinocelular, antes da ressecção cirúrgica do tumor, com estadiamentos de I a IV estabelecida pelo Comitê Americano de Câncer.
- (2) Caracterizar os marcadores de crescimento tumoral p53, Ki67, PCNA e de estresse nitrosativo - 3-nitrotirosina e oxidativo - SOD e proteína carbonílica na biópsia de pacientes submetidos a cirurgia de ressecção do câncer de pele não-melanoma basocelular e espinocelular.
- (3) Caracterizar o estresse oxidativo sistêmico, marcadores plasmáticos de doença avançada (estádio IV) e os níveis de TGF- circulante em pacientes com câncer de pele espinocelular.

Avaliação dos Riscos e Benefícios:

Riscos:

O atual projeto que visa obtenção das amostras dos pacientes não trará risco adicional ao paciente. O risco ao paciente será o da própria cirurgia que este deverá ser submetido o paciente participando ou não do estudo.

Benefícios:

Quanto aos benefícios, a longo prazo este estudo beneficiará todos os pacientes que desenvolverem câncer de pele basocelular e espinocelular no que se refere às estratégias terapêuticas de escolha, quando os biomarcadores e suas correlações com o estresse oxidativo

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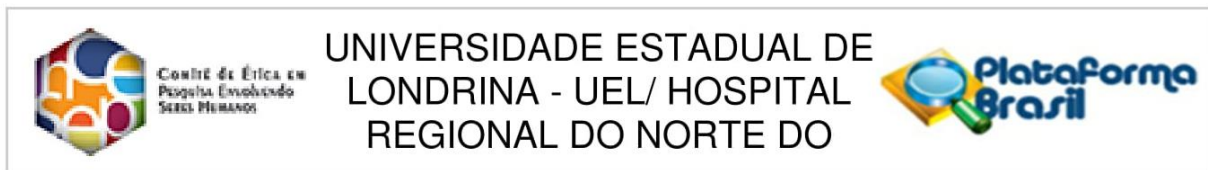
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forem estabelecidas.

Comentários e Considerações sobre a Pesquisa:

Este estudo tem como finalidade caracterizar biomarcadores oxidativos no sangue e de crescimento celular no tecido tumoral de pacientes com câncer de pele não-melanoma em pacientes com diagnóstico clínico de CPNM e com indicação cirúrgica de remoção do tumor.

Estando o mesmo de acordo com as resoluções vigentes pelo CEP e atendendo aos preceitos éticos.

Considerações sobre os Termos de apresentação obrigatória:

Os termos estão adequados e atendem as exigências deste CEP.

Recomendações:

Não existem recomendações

Conclusões ou Pendências e Lista de Inadequações:

Não existem pendências.

Situação do Parecer:

Aprovado

Necessita Apreciação da CONEP:

Não

Considerações Finais a critério do CEP:

LONDRINA, 25 de Maio de 2015

Assinado por:
Paula Mariza Zedu Alliprandini
(Coordenador)

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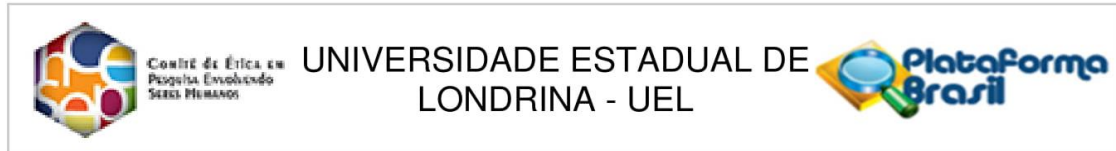
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3**ANEXO B**Cópia do parecer do Comitê de Ética em Pesquisa Envolvendo Seres Humanos da
UEL - 2019**PARECER CONSUBSTANCIADO DO CEP****DADOS DO PROJETO DE PESQUISA**

Título da Pesquisa: ANÁLISE DO ENVOLVIMENTO DE MARCADORES DE ESTRESSE OXIDATIVO E DE PROLIFERAÇÃO CELULAR SISTÊMICOS E NO MICROAMBIENTE TUMORAL NO DESENVOLVIMENTO DE MÚLTIPLOS TIPOS DE CÂNCER DE PELE NÃO-

Pesquisador: Alessandra Lourenço Cecchini Armani

Área Temática:

Versão: 3

CAAE: 00745118.7.0000.5231

Instituição Proponente: Programa de PG em Patologia Experimental

Patrocinador Principal: Financiamento Próprio

DADOS DO PARECER

Número do Parecer: 3.146.725

Apresentação do Projeto:

Trata-se de projeto de pós-graduação, que pretende verificar se existem alterações no perfil oxidativo e inflamatório sistêmico e em marcadores de estresse oxidativo e de proliferação celular no microambiente tumoral em pacientes com queratose actínica e com câncer de pele não-melanoma que possam relacionar com o desenvolvimento de múltiplos tipos de câncer de pele não-melanoma em um mesmo paciente.

Para isso, serão recrutados pacientes do Ambulatório de Especialidades do Hospital Universitário de Londrina-PR. A análise clínica dos pacientes será realizada por médicos especialistas durante as consultas encaminhadas a eles: Dr. André Armani, Cirurgião de Cabeça e Pescoço do Departamento de Clínica Cirúrgica e Dr. Airton dos Santos Gon, Dermatologista do Departamento de Clínica Médica, ambos docentes da Universidade Estadual de Londrina. Se, durante a consulta, houver a suspeita de se tratar de CEC, CBC ou de QA, estes pacientes serão convidados a participar do estudo e caso aceitem, deverão assinar o termo de consentimento livre e esclarecido (TCLE) após total e completa explicação sobre o projeto, seus riscos e benefícios.

Objetivo da Pesquisa:

Objetivo Primário:

Investigar a presença de biomarcadores no sangue e no tecido tumoral de pacientes com câncer de pele não-melanoma que possam relacionar-se com o desenvolvimento de mais de um tipo de

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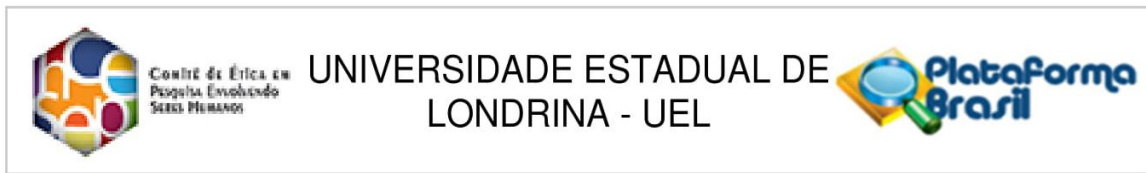
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Continuação do Parecer: 3.146.725

CPNM nos pacientes.

Objetivo Secundário:

- Avaliar o estresse oxidativo sistêmico; - Quantificar parâmetros inflamatórios sanguíneos; - Identificar marcadores de proliferação celular como o

Ki67 e o PCNA no tecido tumoral e no tecido normal adjacente de pacientes submetidos a cirurgia de ressecção do CBC, CEC ou queratose actínica;- Caracterizar o estresse oxidativo através da quantificação de SOD e proteínas carboniladas e

estresse nitrosativo (3-nitrotirosina) no tecido tumoral e no tecido normal adjacente de pacientes submetidos a cirurgia de ressecção do CBC, CEC ou queratose actínica;- Determinar os níveis de p53, Nrf2 e TGF-1 no tecido tumoral e no tecido normal adjacente de pacientes submetidos a cirurgia de ressecção do CBC, CEC ou queratose

actínica;

Avaliação dos Riscos e Benefícios:

Com relação aos riscos a pesquisadora descreve que: "O risco envolvido é inerente à cirurgia a qual será submetido, participando ou não do estudo. No caso de no momento da coleta do sangue o paciente ou controle passarem mal, ele será prontamente atendido pelos médicos e enfermeiros do Hospital de forma que este possa se recuperar prontamente."

Com relação aos benefícios relata que: "Os benefícios para o paciente serão em longo prazo cuja expectativa é de que este estudo possa fornecer ferramentas necessárias para correlacionar o estresse oxidativo com a progressão e reincidência da doença, bem como verificar se há diferenças no prognóstico do paciente com

tumor cutâneo em relação ao estresse oxidativo e se este possui alguma participação na fisiopatologia do tumor"

Comentários e Considerações sobre a Pesquisa:

Não há.

Considerações sobre os Termos de apresentação obrigatória:

Apresenta folha de rosto devidamente preenchida e assinada. Apresenta autorização do HU para realização da pesquisa. O TCLE apresentado está em acordo com a resolução.

O cronograma preve a coleta de amostras para 01/02. O orçamento na Plataforma Brasil, nas informações básicas não está detalhado, mas apresenta detalhamento no arquivo em anexo.

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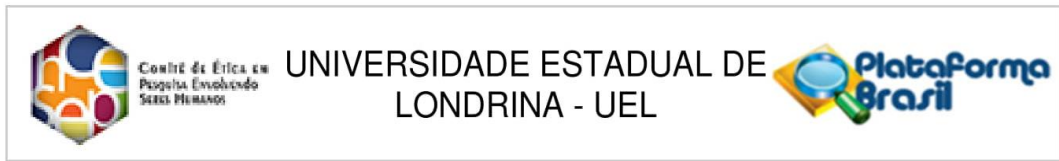
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Continuação do Parecer: 3.146.725

Conclusões ou Pendências e Lista de Inadequações:

As pendências foram atendidas, recomenda-se aprovação.

Considerações Finais a critério do CEP:

Prezado (a) Pesquisador (a),

Este é seu parecer final de aprovação, vinculado ao Comitê de Ética em Pesquisas Envolvendo Seres Humanos da Universidade Estadual de Londrina. É sua responsabilidade imprimi-lo para apresentação aos órgãos e/ou instituições pertinentes.

Coordenação CEP/UEL.

Este parecer foi elaborado baseado nos documentos abaixo relacionados:

| Tipo Documento | Arquivo | Postagem | Autor | Situação |
|---|---|------------------------|--|----------|
| Informações Básicas do Projeto | PB_INFORMAÇÕES_BÁSICAS_DO_PROJETO_1050988.pdf | 23/12/2018 12:30:30 | | Aceito |
| Outros | Resposta_Comite_de_etica_23_12_2018.docx | 23/12/2018 12:24:56 | Alessandra Lourenço Cecchini Armani | Aceito |
| TCLE / Termos de Assentimento / Justificativa de Ausência | TCLE_Pacientes_Controles.docx | 23/12/2018 12:18:14 | Alessandra Lourenço Cecchini Armani | Aceito |
| Orçamento | Orcamento_Dez_2018.docx | 07/12/2018 08:34:10 | Alessandra Lourenço Cecchini Armani | Aceito |
| Brochura Pesquisa | Projeto_30_11_2018.pdf | 07/12/2018 08:27:57 | Alessandra Lourenço Cecchini Armani | Aceito |
| Outros | Resposta_Comite_de_etica_28_11_2018.docx | 07/12/2018 08:27:08 | Alessandra Lourenço Cecchini Armani | Aceito |
| Projeto Detalhado / Brochura Investigador | Projeto_30_11_2018.doc | 07/12/2018 08:25:59 | Alessandra Lourenço Cecchini Armani | Aceito |
| Outros | Parecer_HU_Favoravel.pdf | 08/10/2018 12:07:29 | Alessandra Lourenço Cecchini Armani | Aceito |
| Cronograma | CRONOGRAMA.docx | 08/10/2018 12:06:23 | Alessandra Lourenço Cecchini Armani | Aceito |
| Declaração de Instituição e Infraestrutura | INFRAESTRUTURA.docx | 09/05/2018 10:59:03 | Alessandra Lourenço Cecchini Armani | Aceito |

Endereço: LABESC - Sala 14

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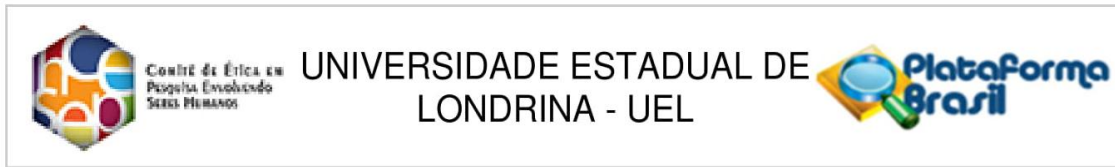
CEP: 86.057-970

UF: PR

Município: LONDRINA

Telefone: (43)3371-5455

E-mail: cep268@uel.br



Continuação do Parecer: 3.146.725

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|----------------|---------------------------|------------------------|--|--------|
| Folha de Rosto | folhaDeRosto_Assinada.pdf | 09/05/2018 10:52:18 | Alessandra Lourenço Cecchini Armani | Aceito |
|----------------|---------------------------|------------------------|--|--------|

Situação do Parecer:

Aprovado

Necessita Apreciação da CONEP:

Não

LONDRINA, 14 de Fevereiro de 2019

Assinado por:
Clisia M. Carreira
(Coordenador(a))

Endereço: LABESC - Sala 14
Bairro: Campus Universitário **CEP:** 86.057-970
UF: PR **Município:** LONDRINA
Telefone: (43)3371-5455 **E-mail:** cep268@uel.br